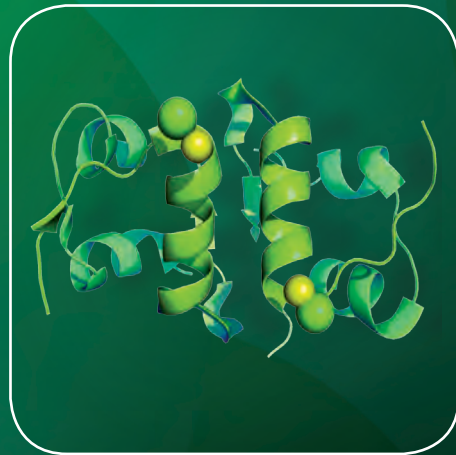
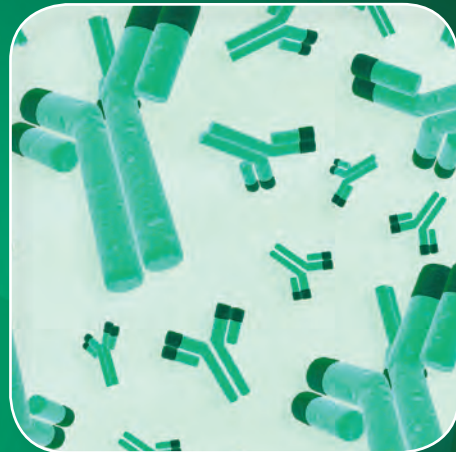


YMC

Biochromatography Columns

RP
SEC
IEX
HIC

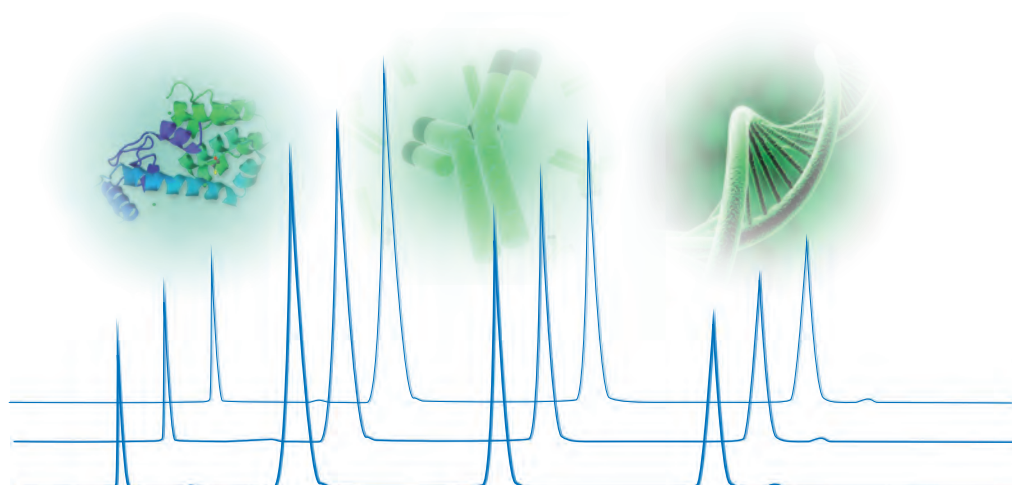


HPLC Columns for Biochromatography

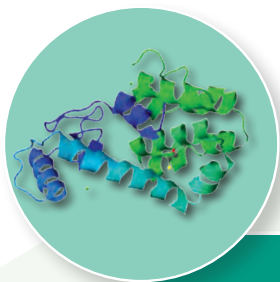
	Reversed Phase (RP)	Size Exclusion (SEC)	Ion Exchange (IEX)	Hydrophobic Interaction (HIC)
Separation principle	Hydrophobicity	Molecular weight	Electric charge	Hydrophobicity
Max. MW	Up to about 150,000 Da	Up to about 1,000,000 Da	Up to several millions Da	Up to about 1,000,000 Da
Resolution	+++	++	+++	+++
Speed	+++	+	++ / +++	+++
Loading	++	++	+++	+++
Stability	+ / ++	+++	+++	+++
Usage (e.g.)	• Peptide mapping • LC/MS • Nucleic acids and oligonucleotides	• Impurity analysis of antibody-drug conjugates • MAb separation	• Proteins/MAB • Charge variant analysis • Isoform analysis • Nucleic acids and oligonucleotides	• Drug-binding analysis of antibody-drug conjugates

New BioLC phase:
YMC-Triart Bio C18

	<i>Page</i>
Phase selection guide	04–05
Separation mechanisms	06–09
Method Validation Kits for BioLC	10
BioLC applications	11–34
Reversed Phase (RP)	35–64
Size Exclusion (SEC)	65–76
Ion Exchange (IEX)	77–92
Hydrophobic Interaction (HIC)	93–100
Substance index	102–103



Phase selection guide



Proteins / Peptides

HIC

BioPro
HIC BF

IEX

BioPro
IEX QA

BioPro
IEX QF

BioPro
IEX SP

BioPro
IEX SF

SEC

YMC-Pack
Diol-60

YMC-Pack
Diol-120

YMC-Pack
Diol-200

YMC-Pack
Diol-300

YMC-SEC MAB

RP

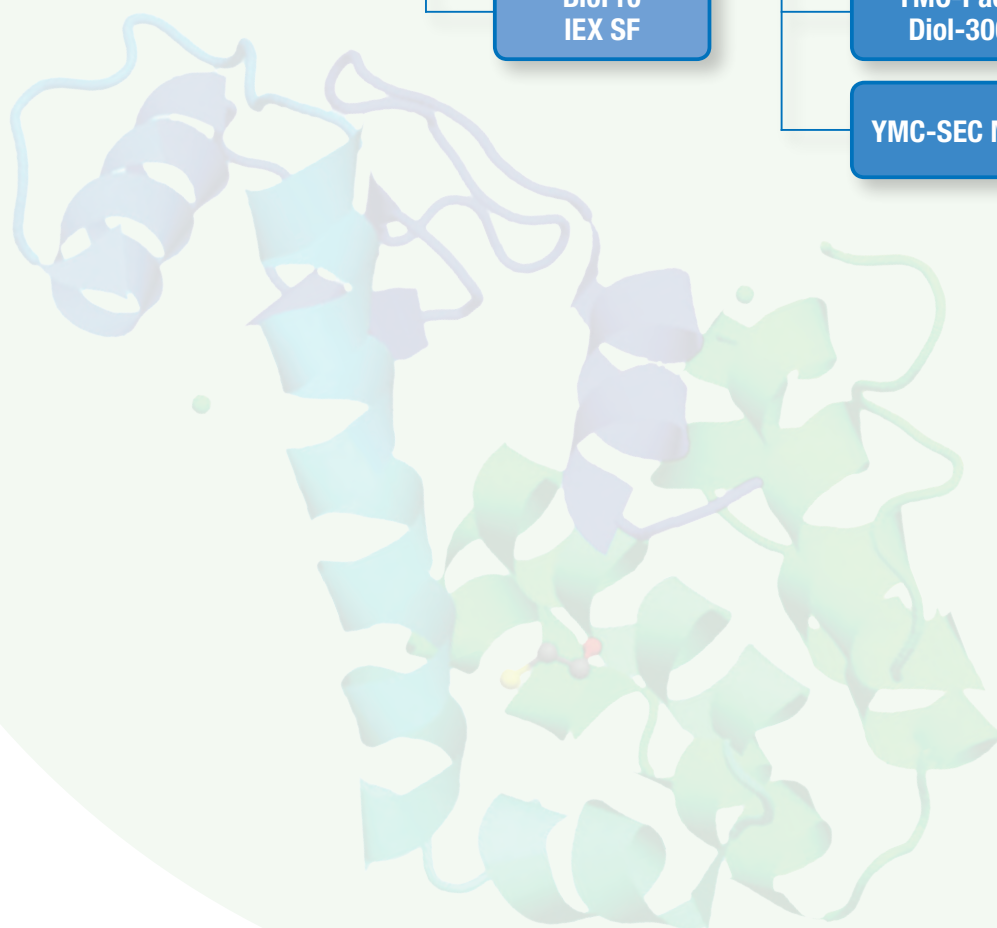
YMC-Triart
C18

YMC-Triart
Bio C18

Meteoric
Core C18 BIO

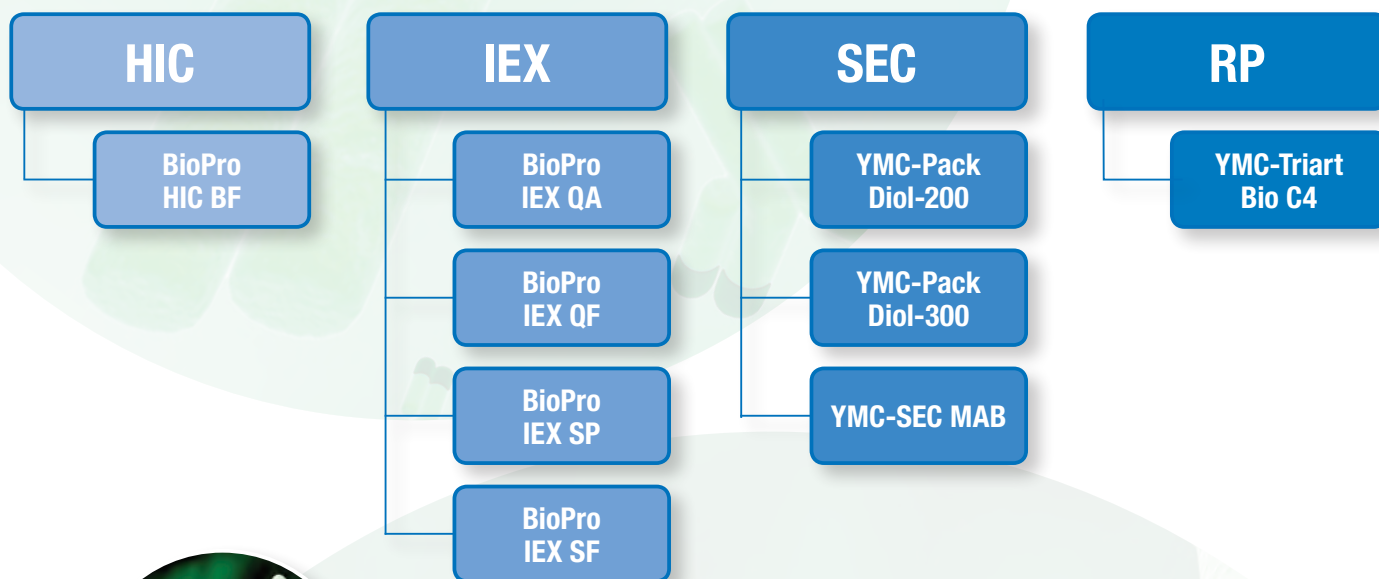
YMCbasic

YMC-Triart
Bio C4

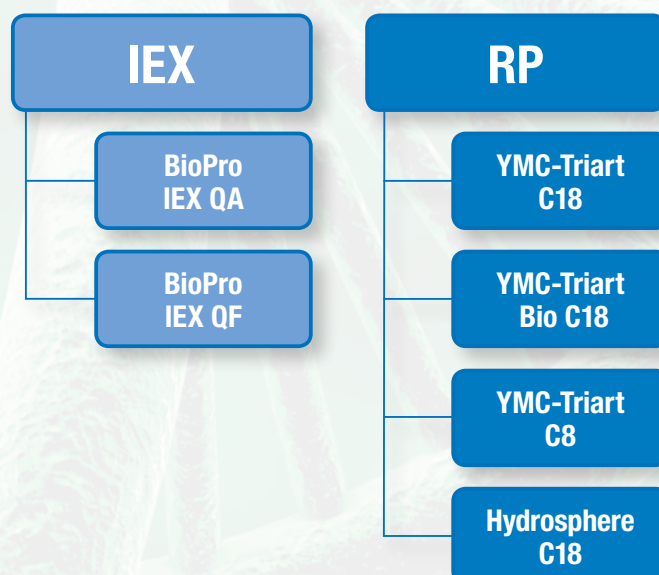




(Monoclonal) Antibodies



Oligonucleotides / Nucleic Acids



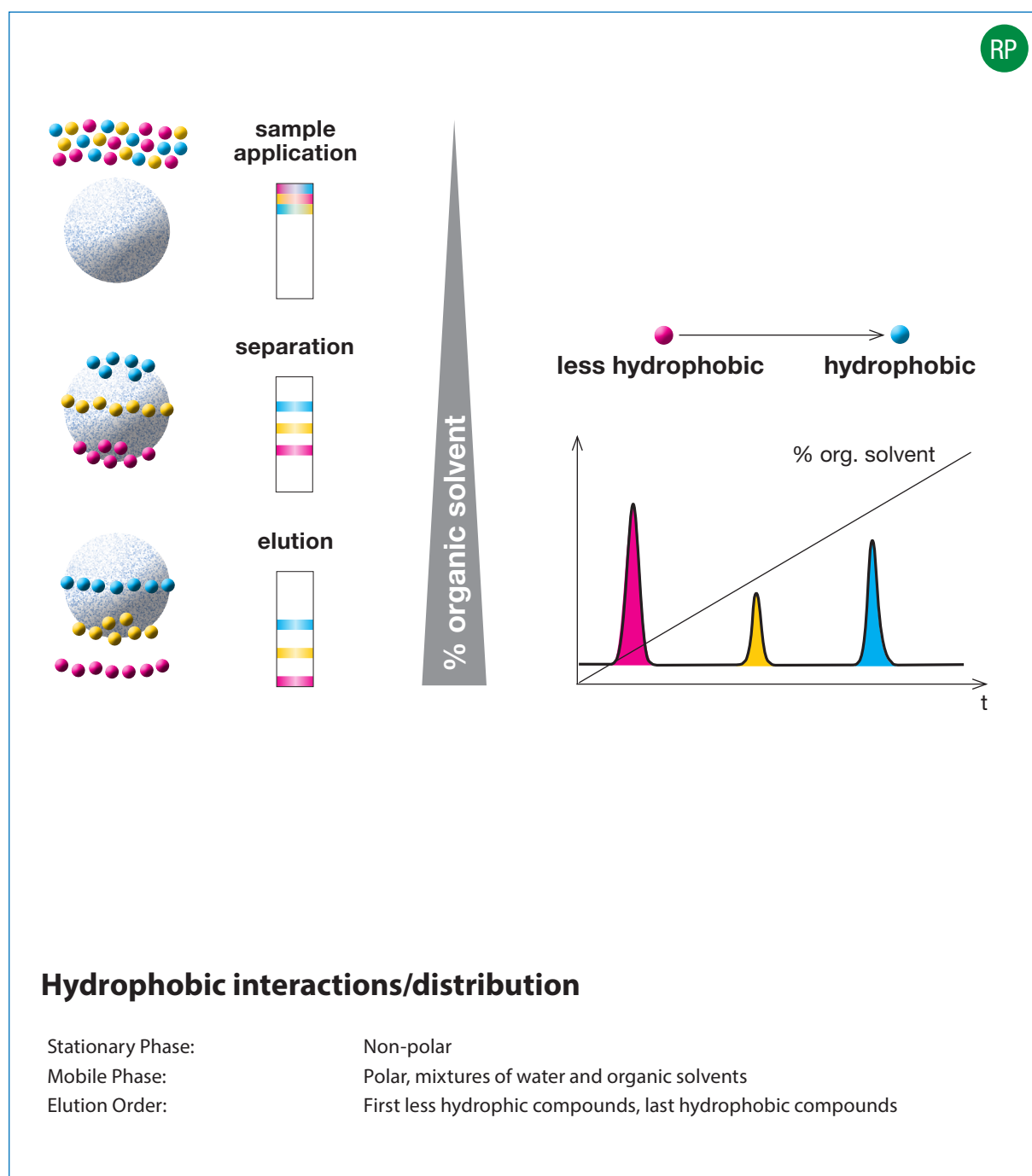
Separation mechanisms

Reversed phase chromatography

Reversed phase chromatography is a technique used to separate molecules using the hydrophobic interactions between compounds and a stationary phase and between compounds and a mobile phase. Stationary phases include silica gels and hybrid silica to which hydrophobic functional groups such as C18, C8, and C4 are

bonded. They are used with mobile phases consisting of mixtures of organic solvents and aqueous acid solutions or buffers.

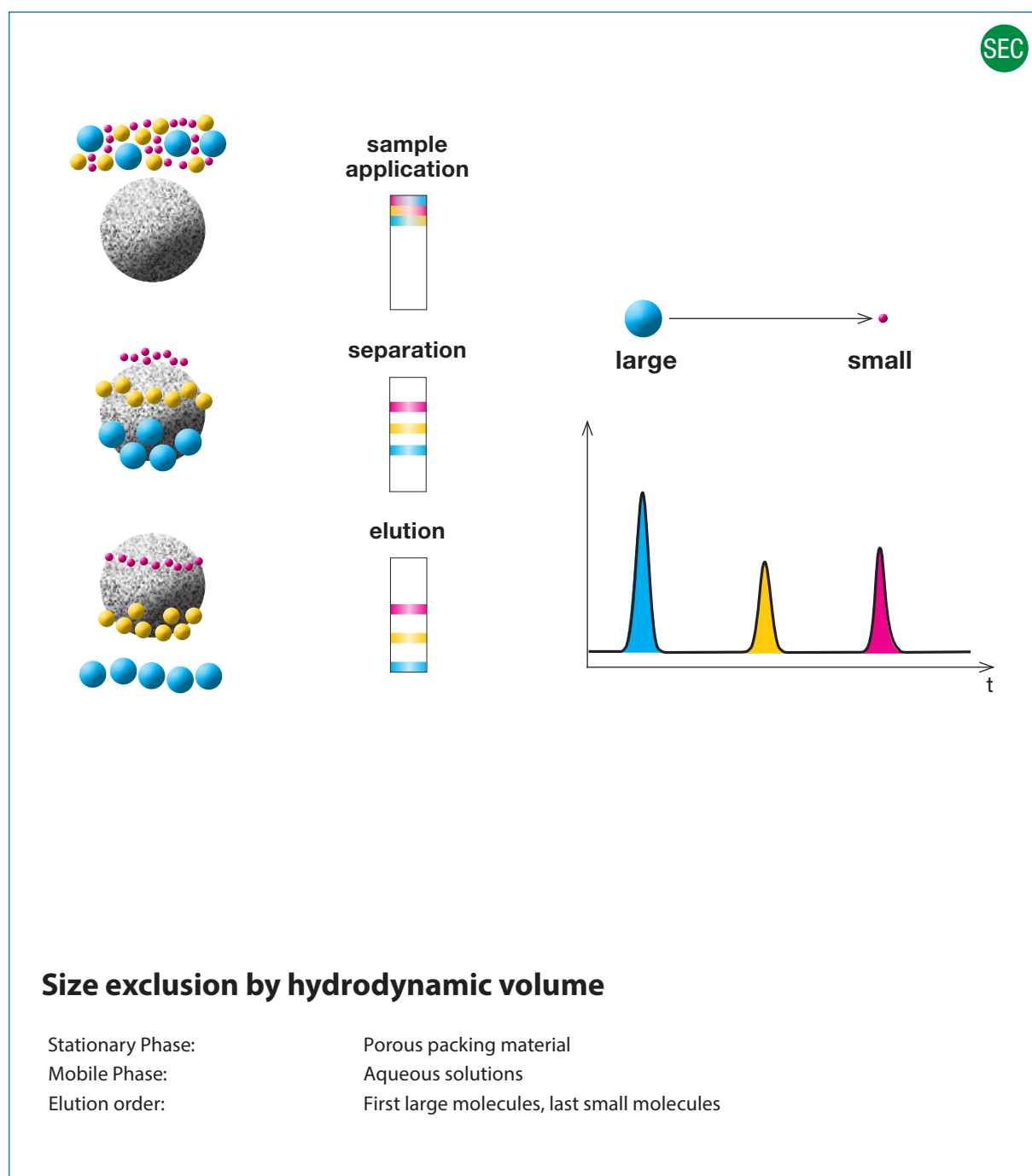
As the concentration of organic solvent in the mobile phase is increased, the molecules with lower hydrophobicity are eluted first.



Size exclusion chromatography

Size exclusion chromatography is a technique used to separate molecules by their size. Stationary phases used consist of silica gels or polymers which have closely defined pores with a network structure. Mobile phases such as buffers which have high solubility and pH stability towards the compounds are used.

Smaller molecules which can penetrate deeper into the pores are eluted later from the column while larger molecules which cannot enter the pores are eluted earlier from the column.

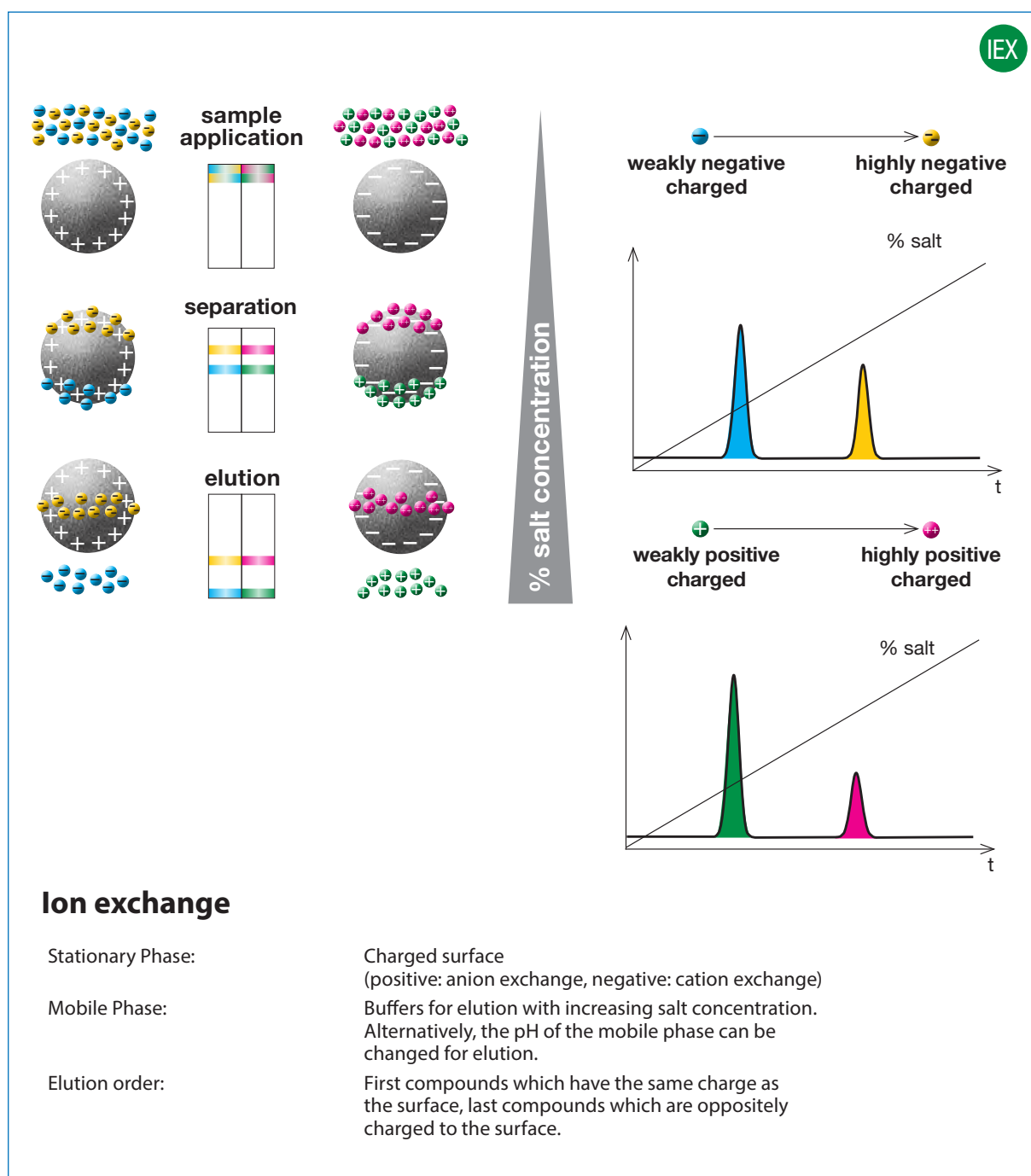


Separation mechanisms

Ion exchange chromatography

Ion exchange chromatography is a technique used to separate ionic molecules by differences in their net charge. Generally, resins (media), which have cationic or anionic groups, are used as stationary phases and buffers containing counter-ions are used as mobile phases. In the sample application step, molecules with opposite

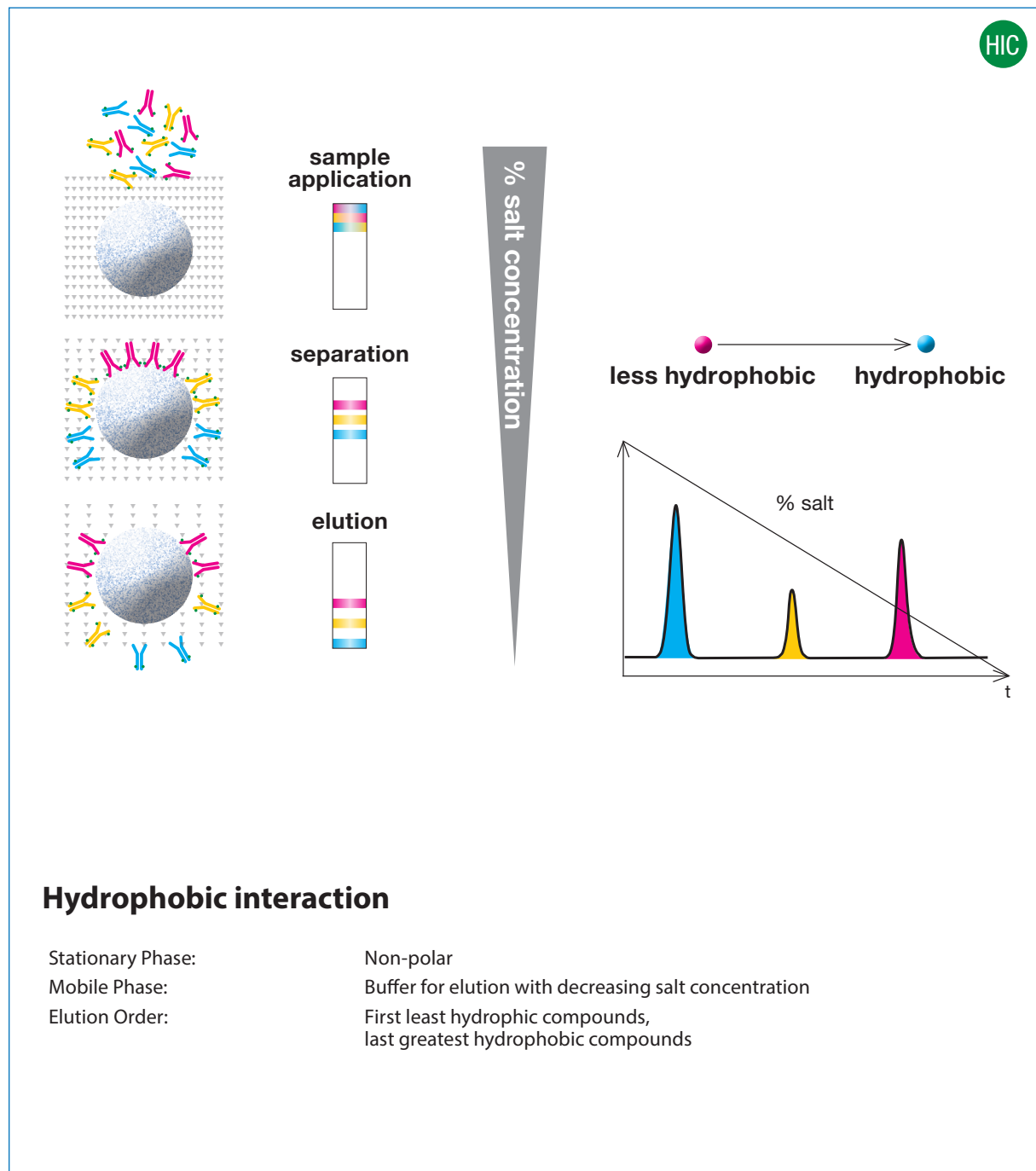
charge to the media bind to the media by ionic interaction. Elution is achieved by increasing the concentration of the counter-ions in the mobile phase. Molecules with the lowest net charge are eluted first and those with higher charge are eluted later.



Hydrophobic interaction chromatography

Hydrophobic interaction chromatography separates molecules using hydrophobic interaction between the stationary phase and compounds such as proteins. Unlike reversed phase, proteins can be separated without any denaturation, thereby maintaining its activity, with the use of ammonium sulphate as a mobile phase.

By lowering the concentration of ammonium sulphate in the mobile phase, bonding power between proteins and the mobile phase is weakened and the proteins are eluted.



Bio QC – Validation kit

Method Validation Kits for BioLC

- for documentation of robustness and reproducibility
- three analytical columns from specified lots

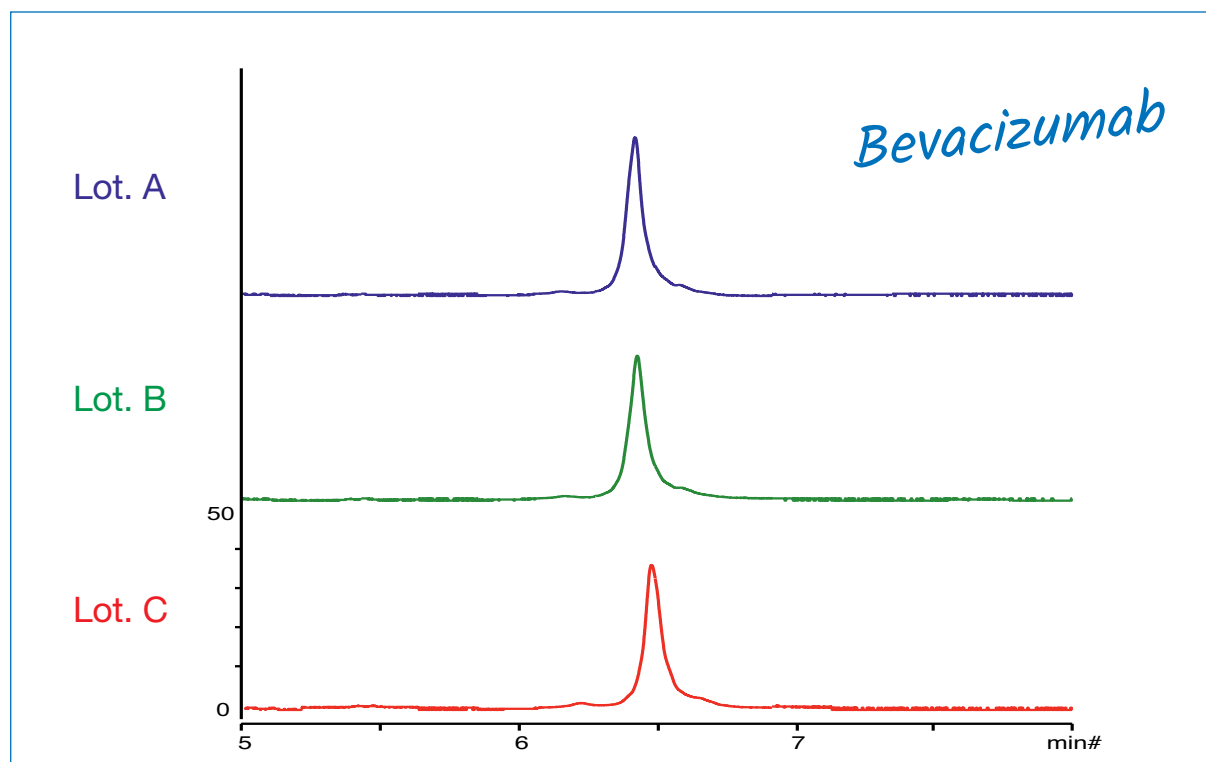
Validation kit:

contains three analytical columns packed with stationary phases from three different batches, in order to solely test the robustness of the particular method.

Available dimensions:

Length: 30 or 33, 50, 75, 100, 150, 250, 300 mm

ID: 2.0 or 2.1, 3.0, 4.0, 4.6, 8.0 mm



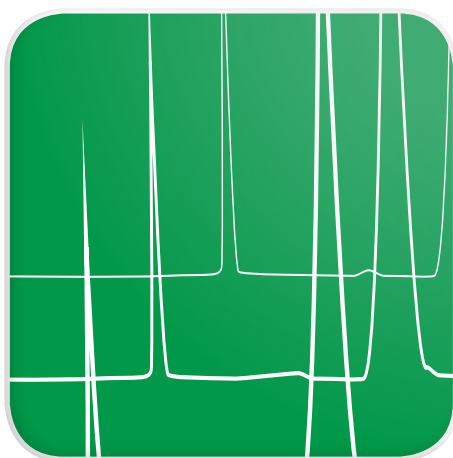
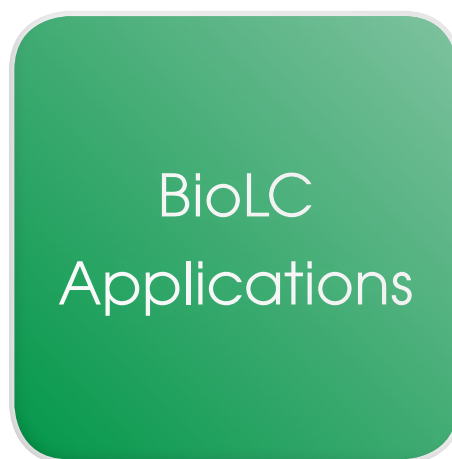
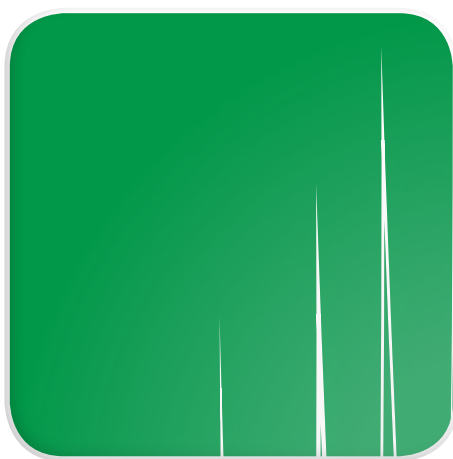
To order a validation kit simply use the ordering number for the column of interest, e.g. **TB30SP9-05Q1PT** and add V1: **TB30SP9-05Q1PTV1**.

For details on YMC selectivities and the International Product Code please refer to the specific product sections in this catalogue.

Batch Reservation Service

Occasionally, a critical analytical method may not prove as robust as you would prefer. Columns from a particular media batch may be the only way that you can, for example, isolate a critical process impurity. In such cases, YMC will reserve a specific batch of material for the use of an individual customer. YMC will also reserve prepacked columns for release according to a pre-arranged schedule.

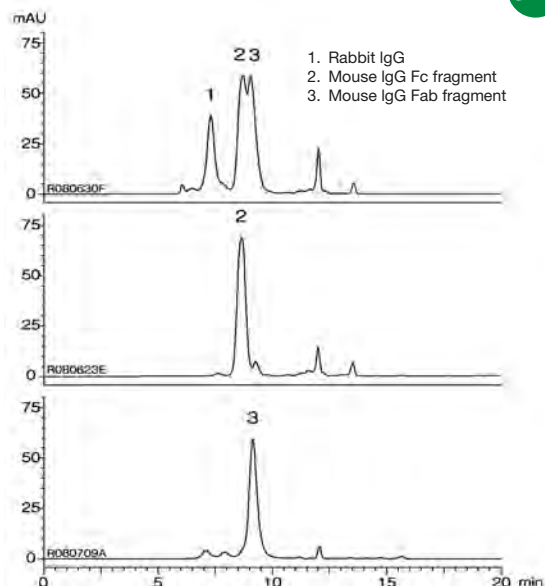
Please call YMC or contact your YMC representative for details.



BioLC applications – Antibodies

IgG, Fab and Fc fragments

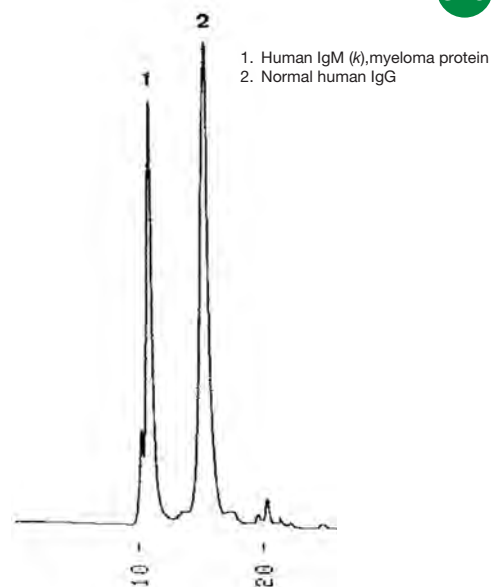
SEC



Column: YMC-Pack Diol-200 (5 μ m) 300 x 8.0 mm ID
Part No.: DL20S05-3008WT
Eluent: 0.1 M KH_2PO_4 - K_2HPO_4 (pH 6.9) containing 0.2 M NaCl
Flow rate: 1.0 mL/min
Temperature: ambient (27 °C)
Detection: UV at 220 nm
Injection: 5 μ L (0.4, 0.5 mg/mL)

Human Immunglobulin

SEC

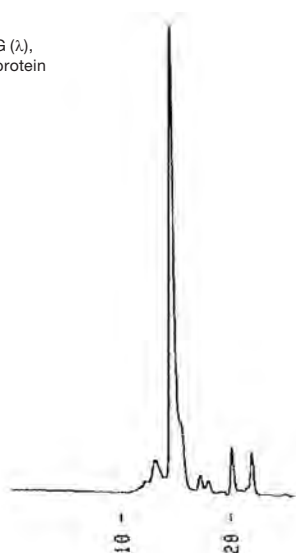


Column: YMC-Pack Diol-300 (5 μ m) 500 x 8.0 mm ID
Part No.: DL30S05-5008WT
Eluent: 0.1 M NaH_2PO_4 - Na_2HPO_4 (pH 6.8) containing 0.1 M Na_2SO_4
Flow rate: 1.0 mL/min
Temperature: ambient (24 °C)
Detection: UV at 280 nm, 0.04 AUFS
Injection: 40 μ L (0.5 mg/mL)

Human IgG (λ), myeloma protein

SEC

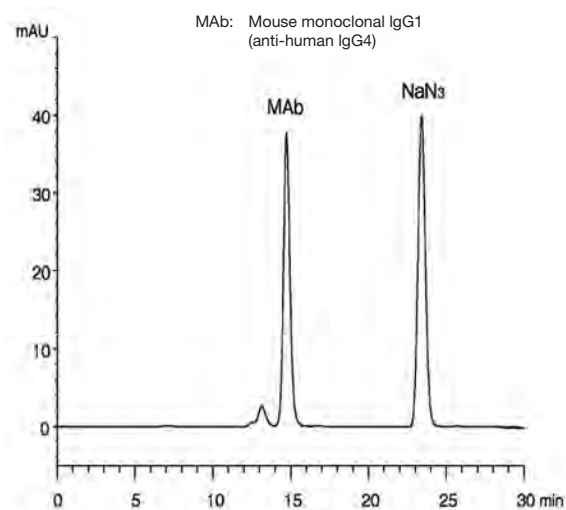
1. Human IgG (λ),
myeloma protein



Column: YMC-Pack Diol-300 (5 μ m) 500 x 8.0 mm ID
Part No.: DL30S05-5008WT
Eluent: 0.1 M NaH_2PO_4 - Na_2HPO_4 (pH 6.8) containing 0.1 M Na_2SO_4
Flow rate: 1.0 mL/min
Temperature: ambient (24 °C)
Detection: UV at 280 nm, 0.04 AUFS
Injection: 20 μ L (1.0 mg/mL)

Monoclonal antibody (MAb)

SEC

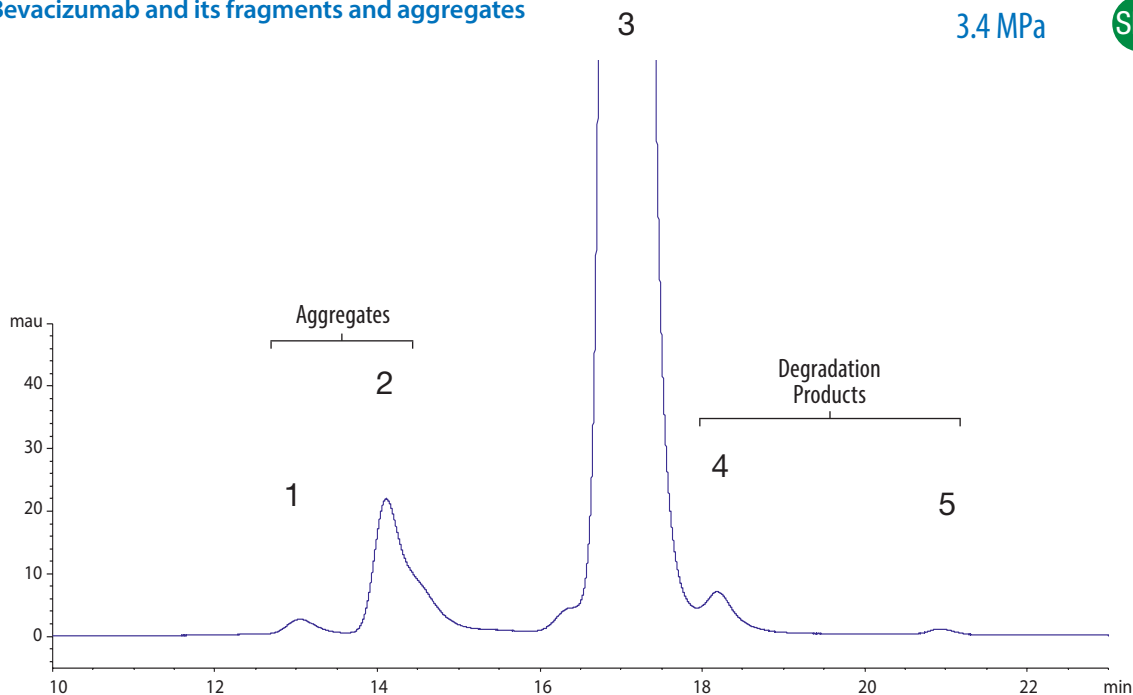


Column: YMC-Pack Diol-200 (5 μ m) 300 x 4.6 mm ID
Part No.: DL20S05-3046WT
Eluent: 0.1 M KH_2PO_4 - K_2HPO_4 (pH 7.0)
Flow rate: 0.17 mL/min
Temperature: ambient (25 °C)
Detection: UV at 220 nm
Injection: 10 μ L
Sample: a commercially available mouse monoclonal IgG1 (0.05 mg/mL)
(purified by DEAE chromatography, containing NaN_3)

Bevacizumab and its fragments and aggregates

3.4 MPa

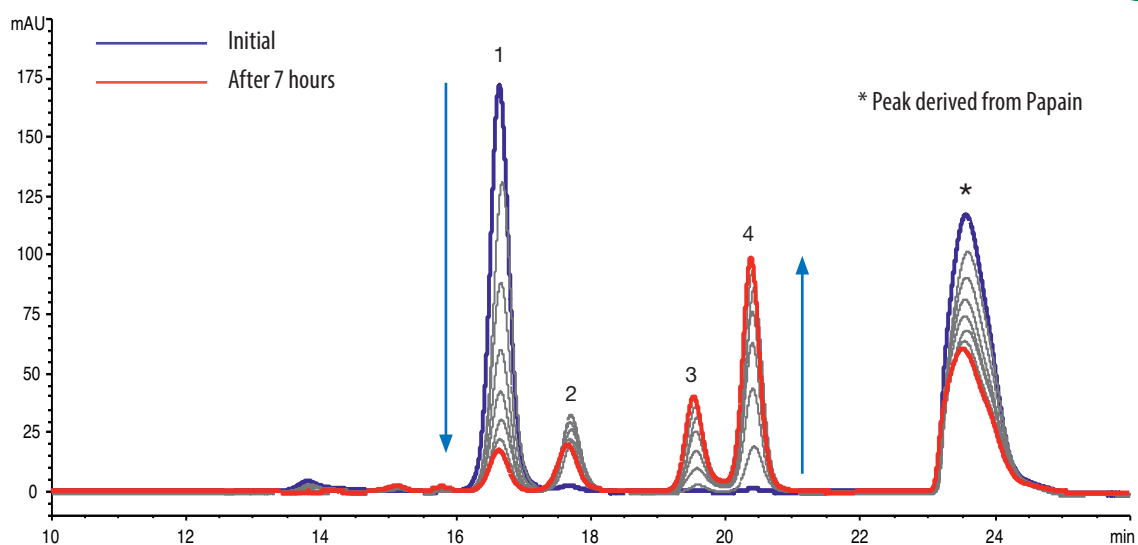
SEC



Column: YMC-SEC MAB (3 μ m, 25 nm) 300 x 4.6 mm ID
 Part No.: DLM25S03-3046WT
 Eluent: 0.1 M phosphate buffer (pH = 7) cont. 0.2 M NaCl
 Flow rate: 0.165 mL/min
 Temperature: 25 °C
 Detection: UV at 280 nm
 Cell path: 10 mm
 Injection: 10 μ L (5 mg/mL)
 Sample: Bevacizumab (Avastin®)

Immunglobulin digest

SEC

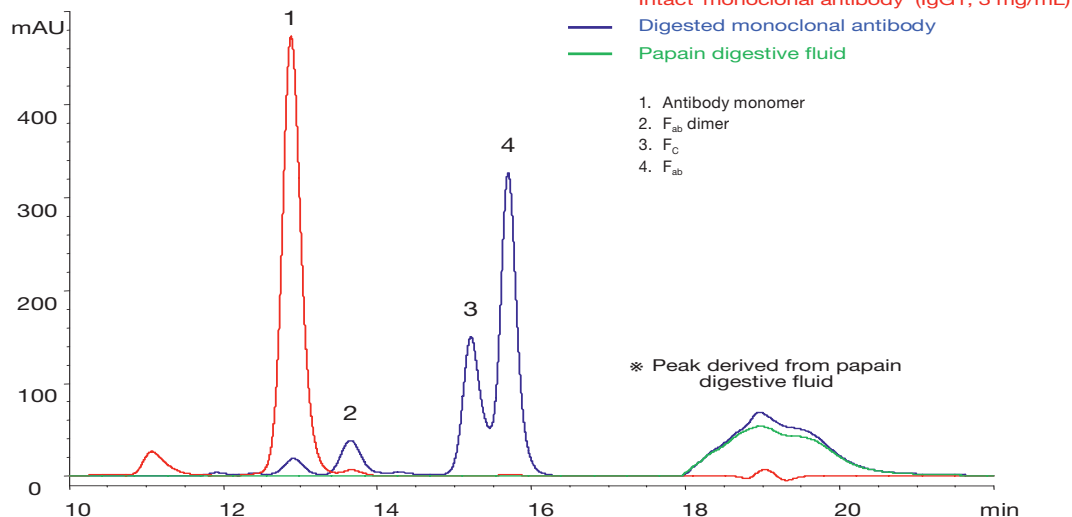


Column: YMC-SEC MAB (3 μ m, 25 nm) 300 x 4.6 mm ID
 Part No.: DLM25S03-3046WT
 Eluent: 0.1 M phosphate buffer (pH = 7) cont. 0.2 M NaCl
 Flow rate: 0.165 mL/min

Temperature: 25 °C
 Detection: UV at 280 nm
 Injection: 2 μ L (3 mg/mL)
 Sample: Humanised monoclonal IgG1 + Papain

BioLC applications – Antibodies

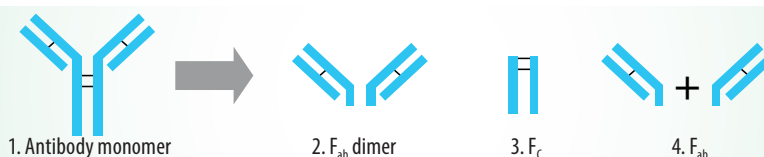
Immunoglobulin digest



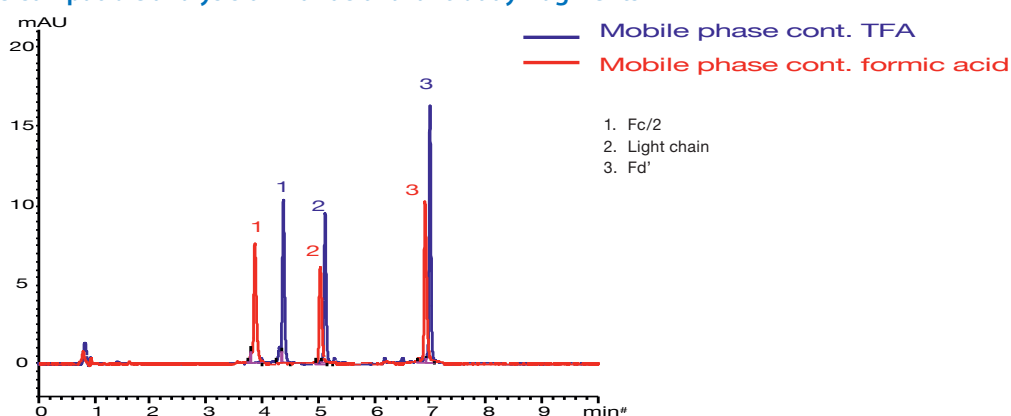
Column: YMC-Pack Diol-200 (2 µm, 20 nm) 300 x 4.6 mm ID
Part no: DL20S02-3046PTH
Eluent: 0.1 M KH₂PO₄-K₂HPO₄ (pH 7.0) containing 0.2 M NaCl
Flow rate: 0.2 mL/min

Temperature: ambient
Detection: UV at 280 nm
Sample: IgG1 (3 mg/ml)

SEC



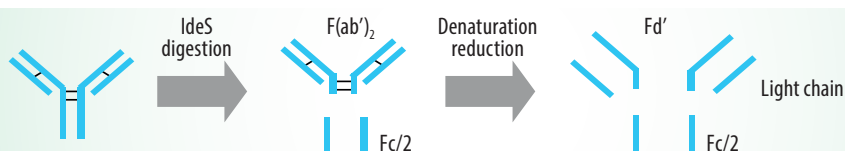
LC/MS compatible analysis of monoclonal antibody fragments



Column size: YMC-Triart Bio C4 (1.9 µm, 30 nm) 150 x 2.1 mm ID
Part no: TB30SP9-15Q1PT
Eluent <TFA>: A) water/TFA (100/0.1)
B) acetonitrile/TFA (100/0.1)
Gradient <TFA>: 25-50%B (0-10 min), 90%B (10-12.5 min)
Eluent <Formic acid>: A) water/formic acid (100/0.1)
B) acetonitrile/formic acid (100/0.1)
Gradient <Formic acid>: 20-45%B (0-10 min), 90%B (10-12.5 min)

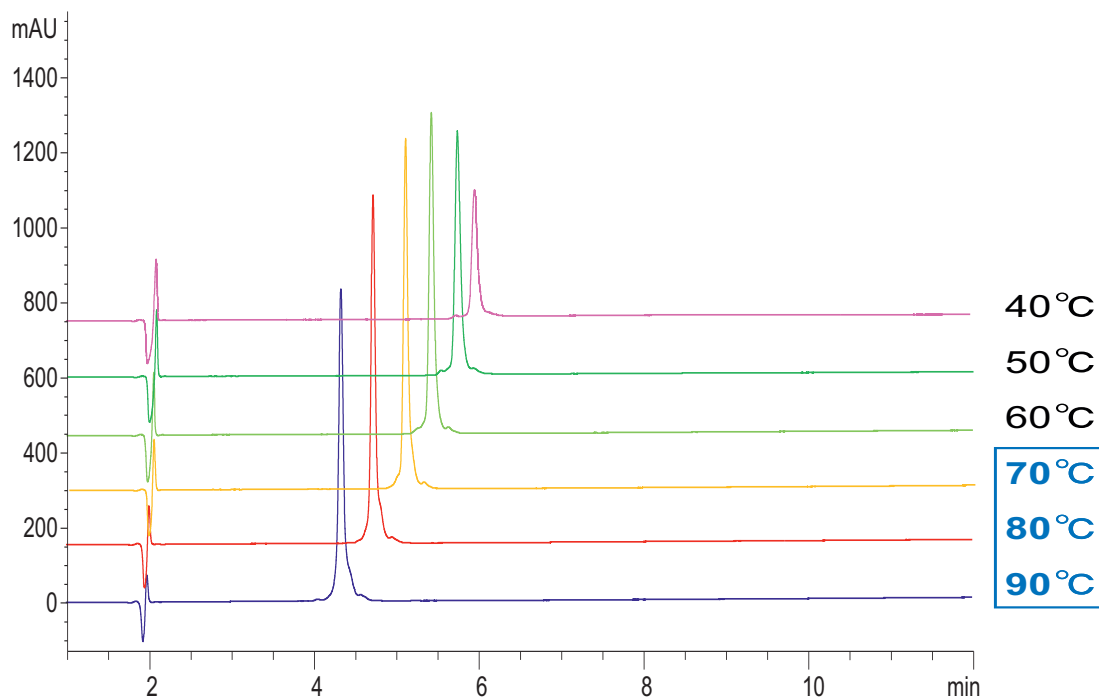
Detection: UV at 280 nm
Flow rate: 0.4 mL/min
Temperature: 80 °C
Injection: 4 µL (0.25 mg/mL)
Sample: mAb Subunit Standard (Waters Corp.)

RP

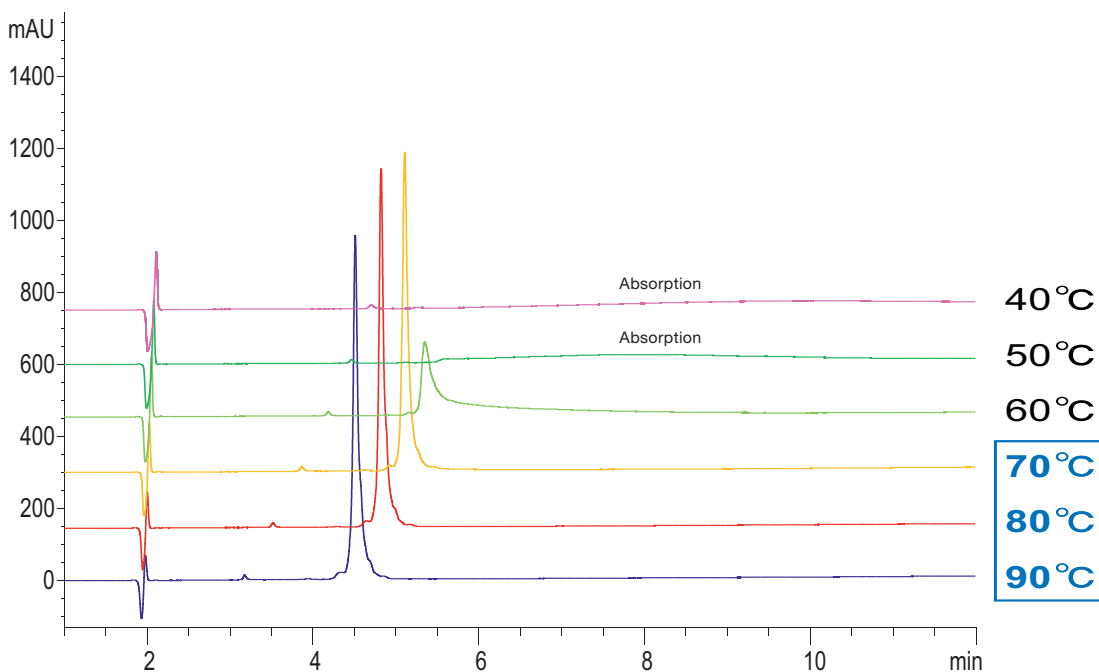


Adalimumab (Humira®, MW: ca. 148 kDa)

RP



Bevacizumab (Avastin®, MW: ca. 148 kDa)

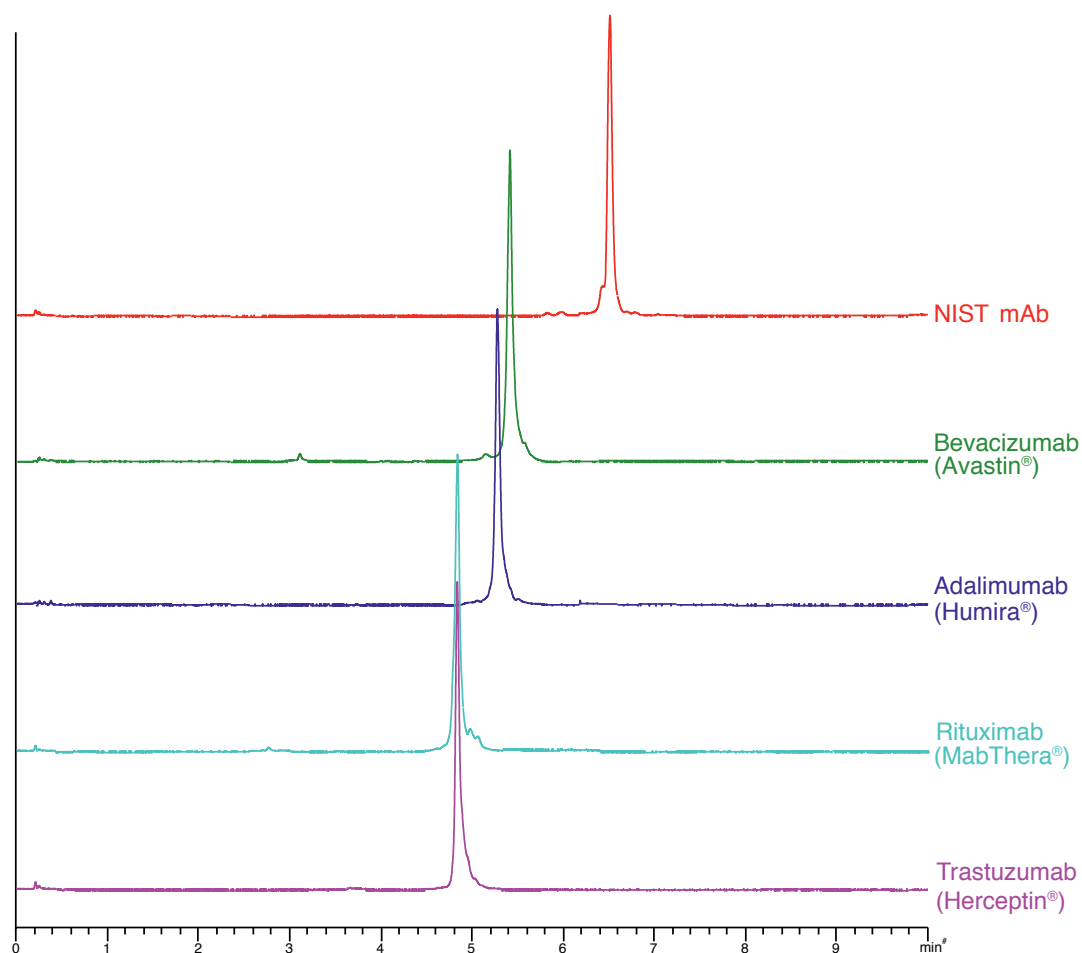


Column: YMC-Triart Bio C4 (3 μ m, 30 nm) 150 x 3.0 mm ID
 Eluent: A) water/TFA (100/0.1)
 B) acetonitrile/TFA (100/0.1)
 Gradient: 30-60%B (0-15 min), 90%B (15-30min)
 Detection: UV at 220 nm
 Flow rate: 0.4 mL/min
 Sample: Adalimumab (0.5 mg/mL) or Bevacizumab (0.5 mg/mL)
 Injection: 4 μ L

BioLC applications – Antibodies

Analysis of different monoclonal antibodies

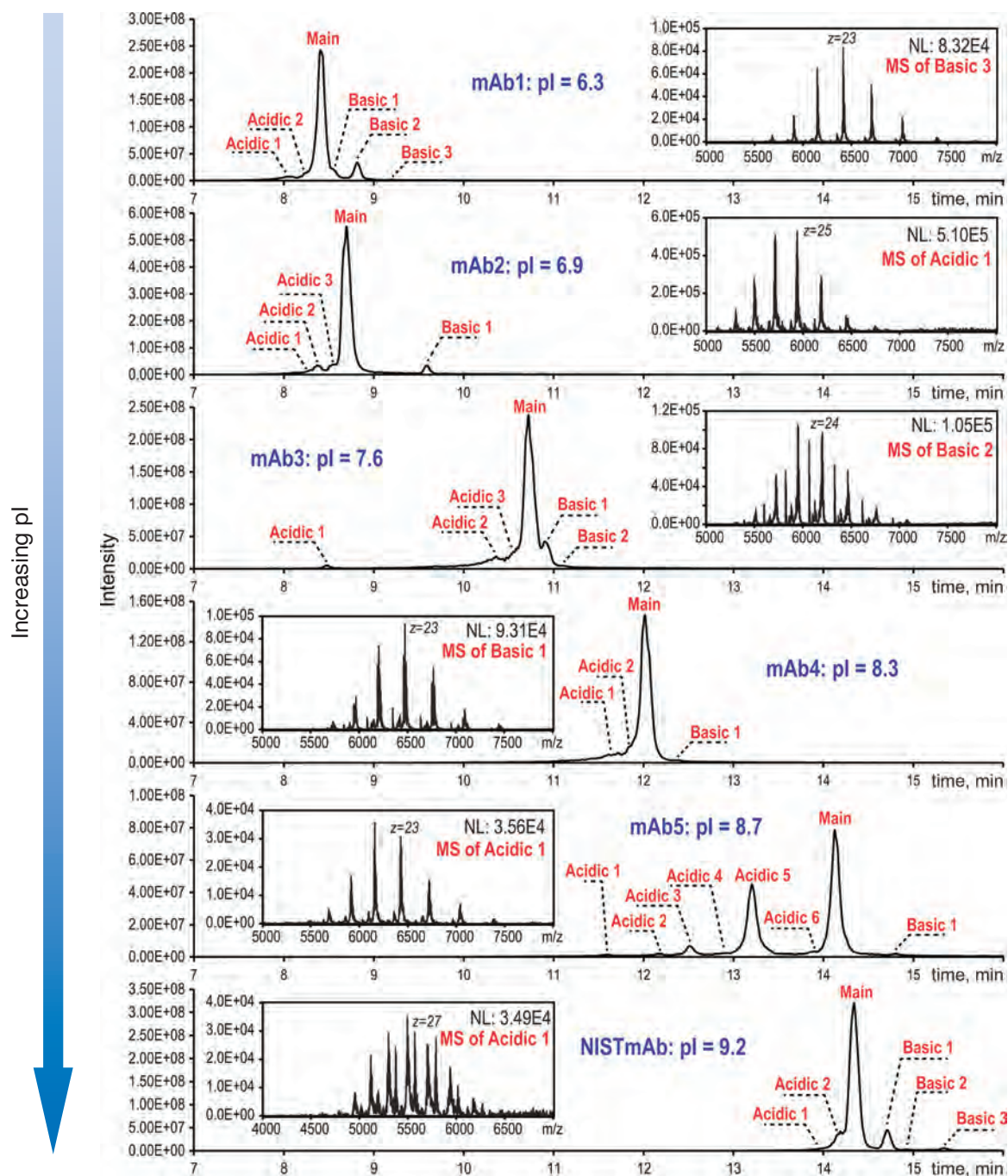
RP



Column: YMC-Triart Bio C4 (1.9 μ m, 30 nm) 50 x 2.1 mm ID
Part No.: TB30SP9-05Q1PT
Eluent: A) water/TFA (100/0.1)
B) acetonitrile/TFA (100/0.1)
Gradient: 25-45%B (0-10 min)
Detection: UV at 280 nm (0.13s, 40Hz)
Flow rate: 0.4 mL/min
Temperature: 80 °C
Sample conc.: 0.5 mg/mL
Injection: 2 μ L

Separation of different charge heterogenic mAbs via SCX-MS

IEX



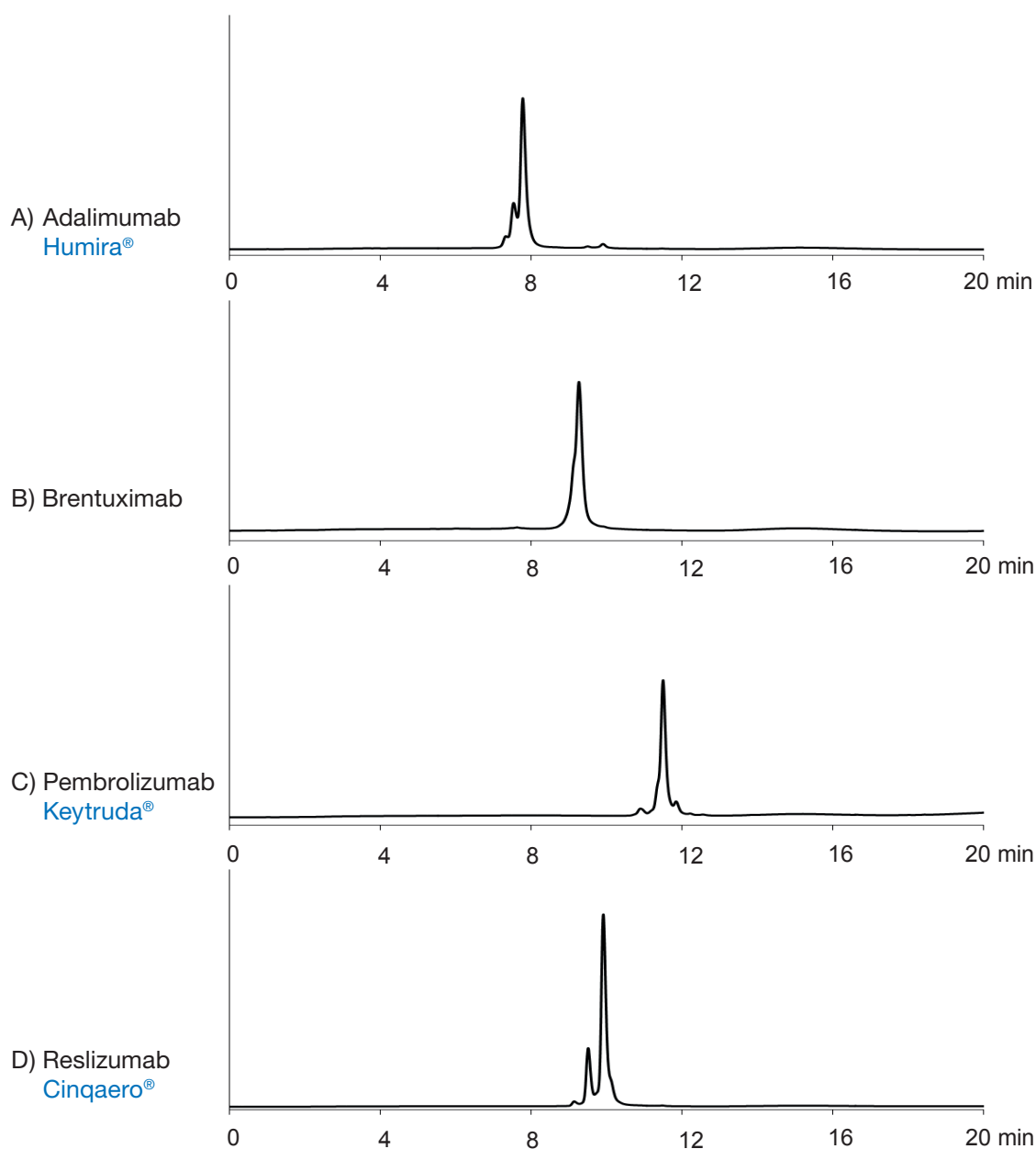
Column: BioPro IEX SF (5 μ m) 100 x 4.6 mm ID
Part. No.: SF00S05-1046WP
Eluent: A) 20 mM $\text{CH}_3\text{COONH}_4$ pH 5.6 adjusted with 20 mM CH_3COOH
B) 140 mM $\text{CH}_3\text{COONH}_4$, 10 mM $(\text{NH}_4)_2\text{CO}_3$, pH 7.6
Gradient: 0% B (0–2 min), 0–100% B (2–16 min), 100% B (16–20 min), 0% B (20–27 min)
Flow rate: 0.4 mL/min
Temperature: 45 $^\circ\text{C}$
Equipment: Post column analytical splitter (~ 400:1) to reduce analytical flow to ca. 1 $\mu\text{L}/\text{mi}$
Detector: PDA (UV)
PicoTip Emitter (NSI-MS)

Application data taken from Y. Yan, A. P. Liu, S. Wang, T. J. Daly und N. Li, "Ultrasensitive Characterization of Charge Heterogeneity of Therapeutic Monoclonal Antibodies," Anal. Chem., 2018, 90, 13013-13020.

BioLC applications – Antibodies

HIC Analysis of different monoclonal antibodies using isopropanol as modifier

HIC

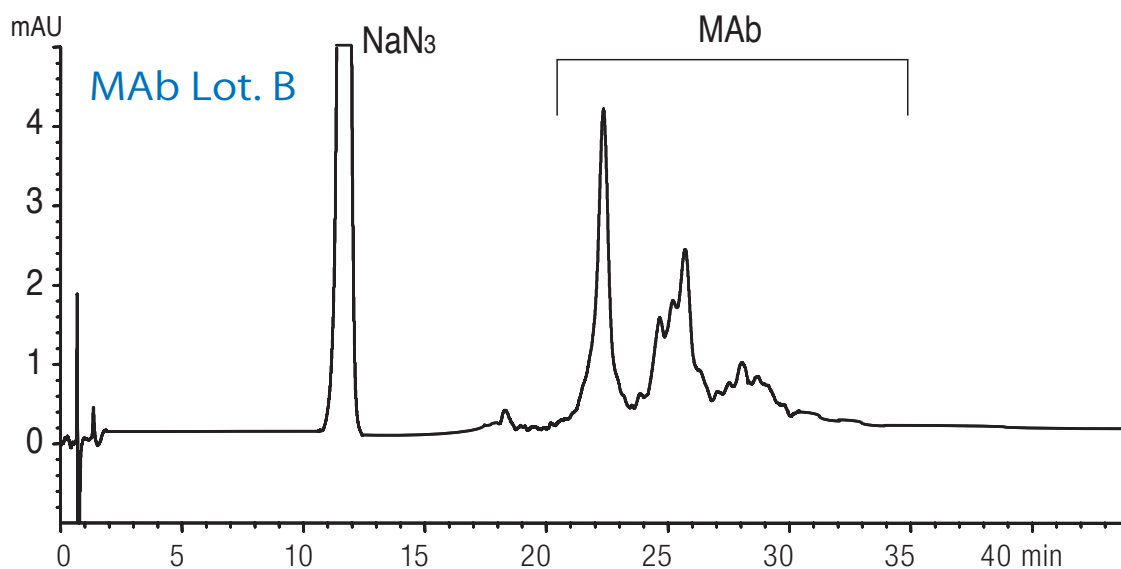
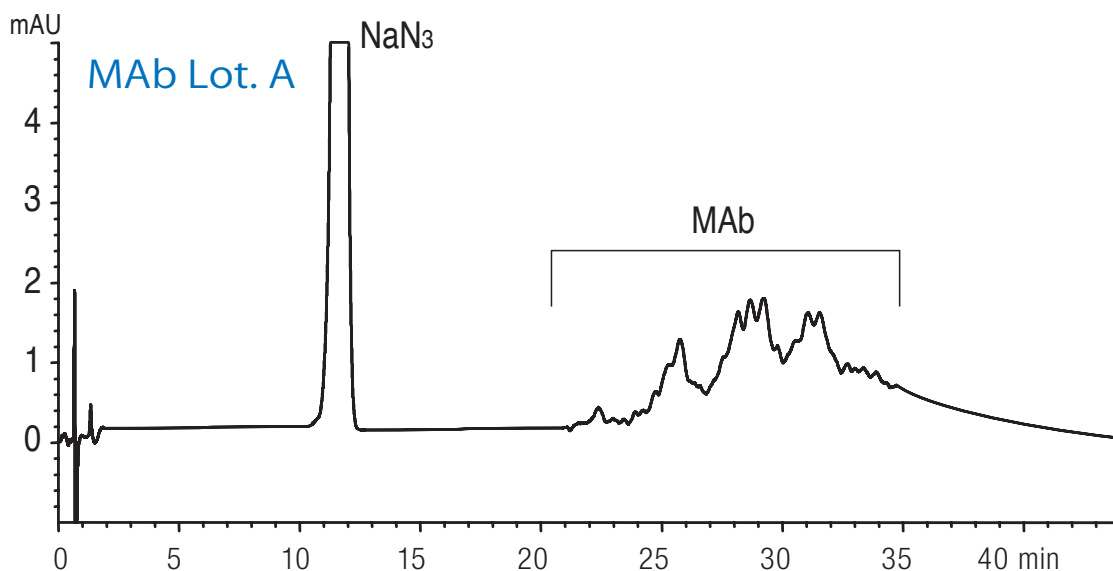


Column: BioPro HIC BF (4 μ m) 100 x 4.6 mm ID
 Part No.: BHB00S04-1046WT
 Eluent: A) 20 mM NaH_2PO_4 - Na_2HPO_4 (pH 7.4) containing 1.5 M $(\text{NH}_4)_2\text{SO}_4$
 B) 20 mM NaH_2PO_4 - Na_2HPO_4 (pH 7.4) / 2-propanol (85/15)
 Gradient: 0–100 % B (0–20 min)
 Flow rate: 1.0 mL/min
 Temperature: 20 °C
 Detection: fluorescence: ex 280 nm, em 360 nm
 Injection: 3 μ L (2 mg/mL)

by courtesy of University of Geneva, School of Pharmaceutical Sciences,
Department of Analytical Pharmaceutical Chemistry

Different production batches of IgG1

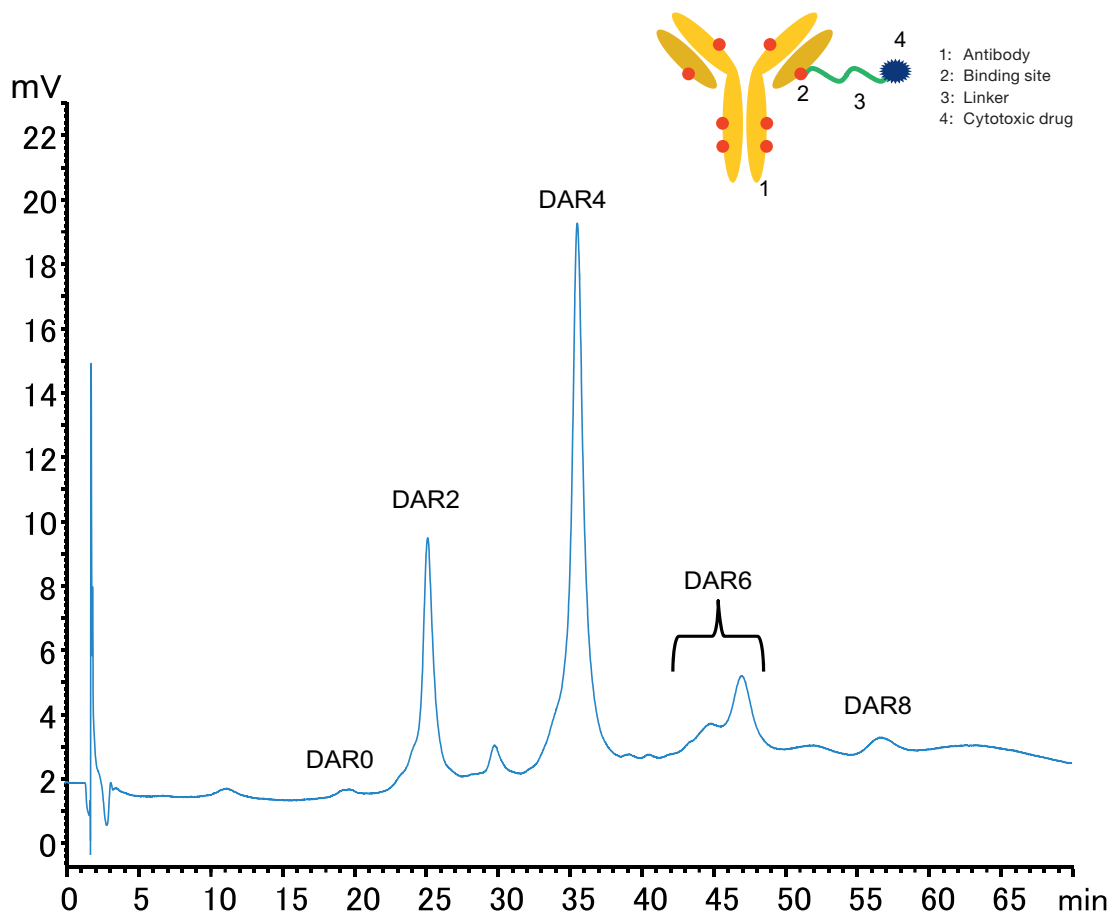
IEX



Column: BioPro IEX QF (5 μm) 100 x 4.6 mm ID
 Part No.: QF00S05-1046WP
 Eluent: A) 20 mM Tris-HCl (pH 8.1)
 B) 20 mM Tris-HCl (pH 8.1) containing 0.5 M NaCl
 Gradient: 10-25%B (0-60 min)
 Flow rate: 1.0 mL/min (360 cm/hr)
 Temperature: 25 °C
 Detection: UV at 220 nm
 Injection: 14 μL (0.1 mg/mL)
 Sample: Mouse monoclonal IgG1 anti-human IgG4
 (Purified by DEAE chromatography, containing NaN_3)

BioLC applications – Antibody-Drug-Conjugates

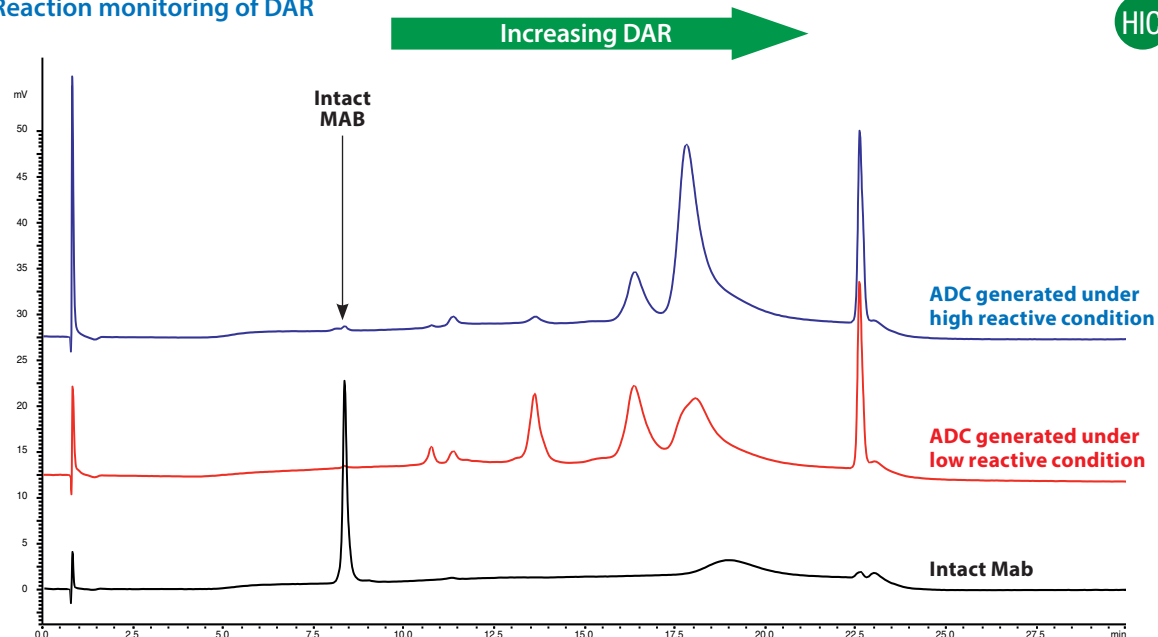
Drug to Antibody Ratio



Column: BioPro HIC BF (4 μ m) 100 x 4.6 mm ID
 Part No.: BHB00S04-1046WT
 Eluent: A) 50 mM NaH_2PO_4 - Na_2HPO_4 (pH 6.8) containing 1.5 M $(\text{NH}_4)_2\text{SO}_4$ /IPA (95/5)
 B) 50 mM NaH_2PO_4 - Na_2HPO_4 (pH 6.8)/2-propanol (80/20)
 Gradient: 30%B (0-5 min)
 30-80%B (5-45 min)
 80%B (45-70 min)
 Flow rate: 0.4 mL/min
 Temperature: 25 $^\circ\text{C}$
 Detection: UV at 280 nm
 Injection: 20 μL
 Sample: Cystein-conjugated ADC mimic (1.25 mg/mL)

BioLC applications – Antibody-Drug-Conjugates

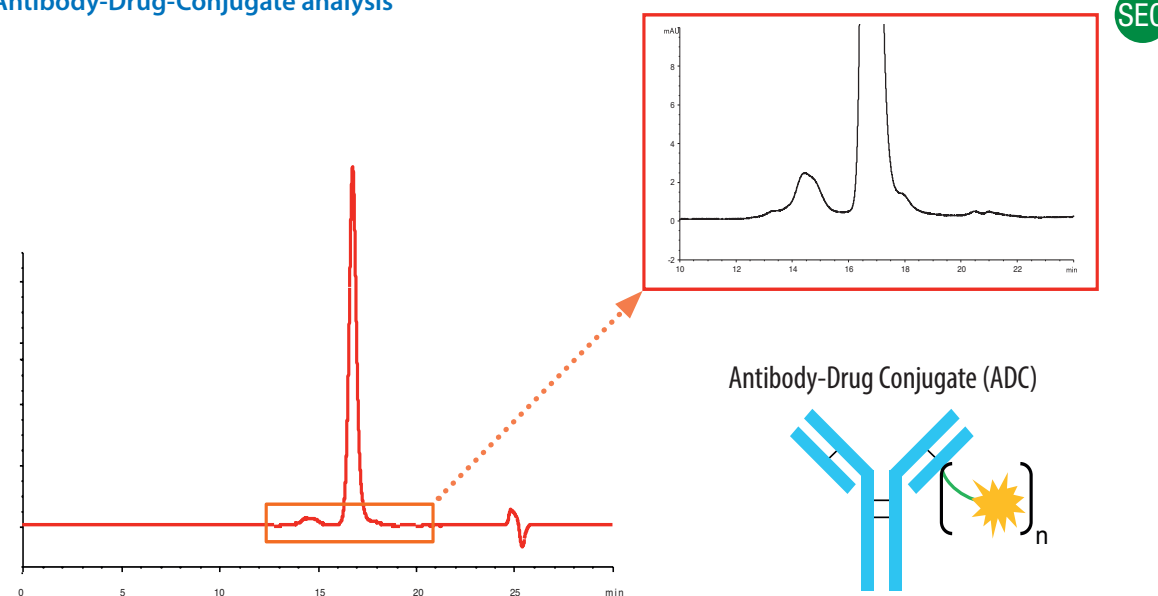
Reaction monitoring of DAR



Column: BioPro HIC BF (4 μ m) 100 x 4.6 mm ID
 Part No.: BHB00S04-1046WT
 Eluent: A) 50 mM NaH_2PO_4 - Na_2HPO_4 (pH 7.0) containing 1.5 M $(\text{NH}_4)_2\text{SO}_4$ /2-propanol (95/5)
 B) 50 mM NaH_2PO_4 - Na_2HPO_4 (pH 7.0)/2-propanol (80/20)
 Gradient: 0%B (0-1 min), 0-100%B (1-15 min), 100%B (15-20 min), 0%B (20-30 min)
 Flow rate: 1.0 mL/min
 Temperature: 25 $^\circ\text{C}$
 Detection: UV at 280 nm
 Sample: Antibody Drug Conjugate*

*Courtesy of RIKEN

Antibody-Drug-Conjugate analysis

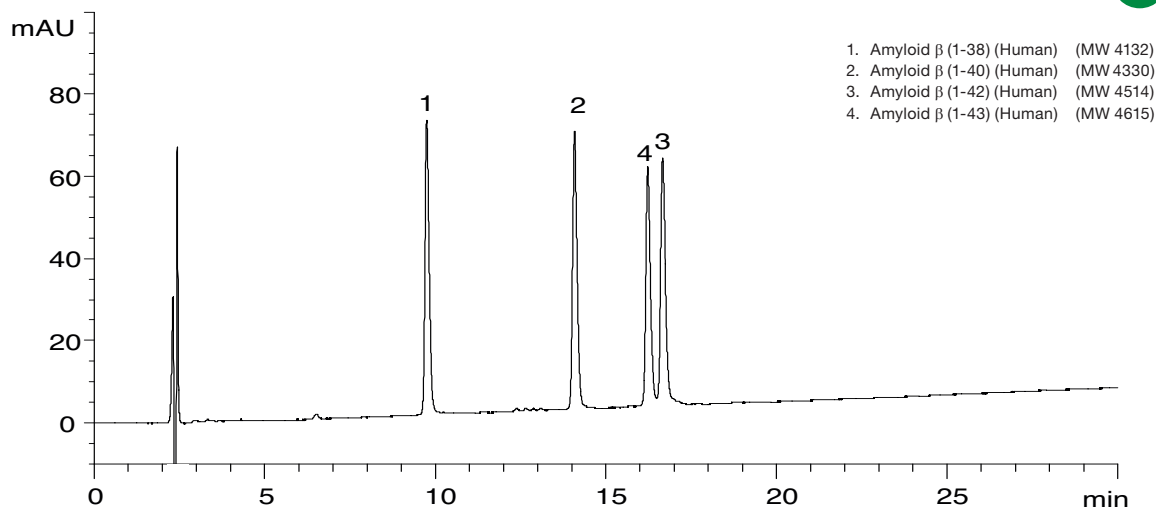


Column: YMC-SEC MAB (3 μ m, 25 nm) 300 x 4.6 mm ID
 Part No.: DLM25S03-3046WT
 Eluent: 0.1 M phosphate buffer (pH = 7) cont. 0.2 M NaCl / 2-propanol (85 / 15)
 Flow rate: 0.165 mL/min
 Temperature: 25 $^\circ\text{C}$
 Detection: UV at 280 nm
 Injection: 4 μ L (2.5 mg/mL)
 Sample: SigmaMAb Antibody Drug Conjugate Mimic

BioLC applications – Proteins

Amyloid β -proteins

RP



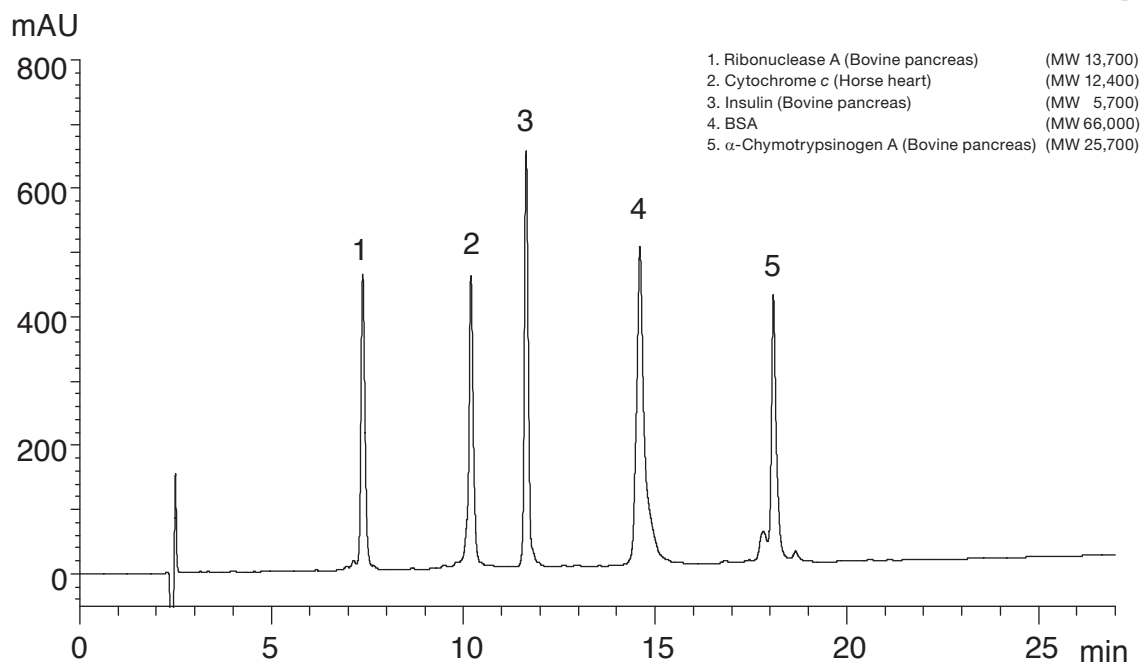
Amyloid β (1-43) : Asp-Ala-Glu-Phe-Arg-His-Asp-Ser-Gly-Tyr-Glu-Val-His-His-Gln-Lys-Leu-Val-Phe-Phe-Ala-Glu-Asp-Val-Gly-Ser-Asn-Lys-Gly-Ala-Ile-Ile-Gly-Leu-Met-Val-Gly-Gly-Val-Val-Ile-Ala-Thr

Column: YMC-Triart Bio C4 (3 μ m, 30 nm) 150 x 3.0 mm ID
 Part No.: TB30S03-1503PTH
 Eluent: A) water/TFA (100/0.1)
 B) acetonitrile/TFA (100/0.1)
 Gradient: 25-40%B (0-30 min), 90%B (30-40 min)

Flow rate: 0.4 mL/min
 Temperature: 70 °C
 Detection: UV at 220 nm
 Injection: 4 μ L (each 0.1 mg/mL)

Proteins (MW 5,700 ~ 66,000)

RP

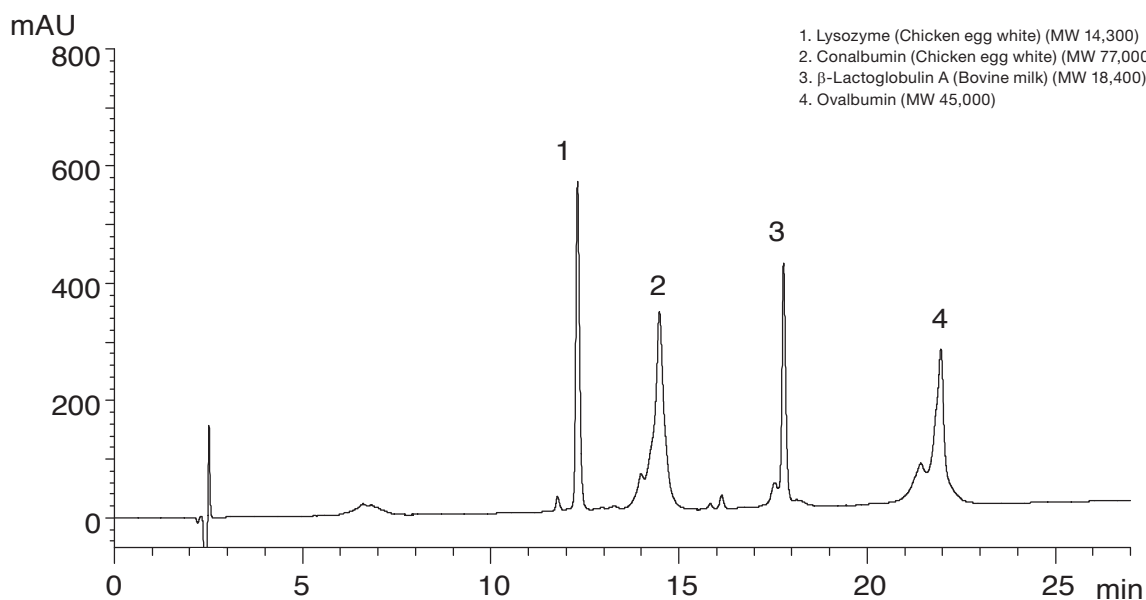


Column: YMC-Triart Bio C4 (5 μ m, 30 nm) 150 x 3.0 mm ID
 Part No.: TB30S03-1503PTH
 Eluent: A) water/TFA (100/0.1)
 B) acetonitrile/TFA (100/0.1)
 Gradient: 20-60%B (0-27 min), 90%B (27-35 min)

Flow rate: 0.4 mL/min
 Temperature: 70 °C
 Detection: UV at 220 nm
 Injection: 10 μ L (0.25 ~ 0.50 mg/mL)

Proteins (MW 14,300 ~ 77,000)

RP

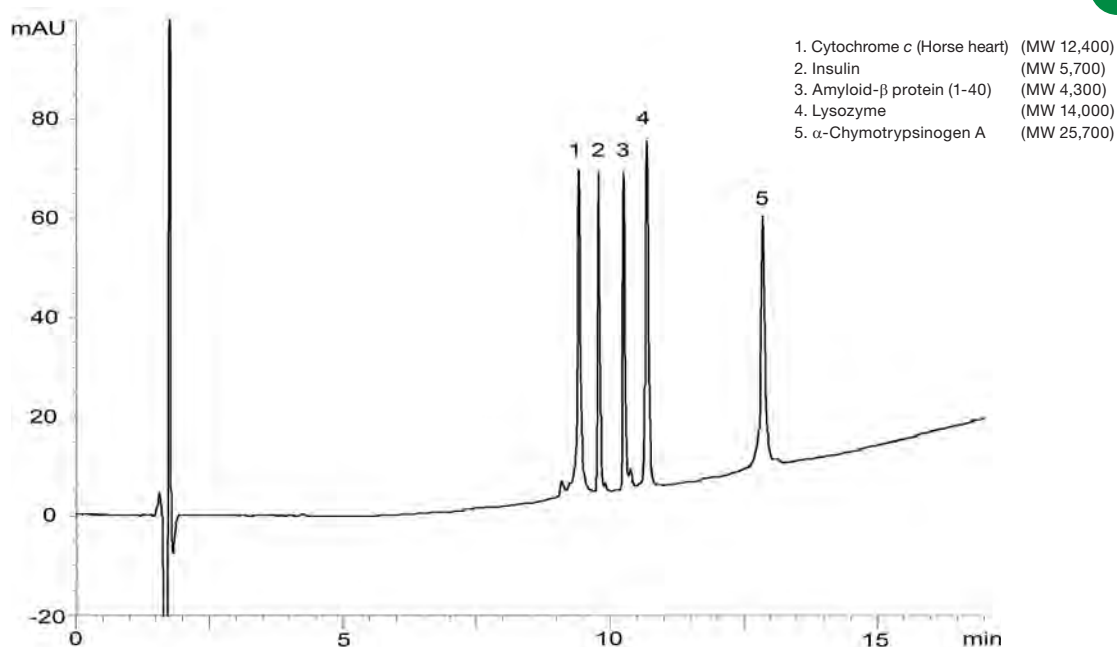


Column: YMC-Triart Bio C4 (5 μ m, 30 nm) 150 x 3.0 mm ID
 Part No.: TB30S03-1503PTH
 Eluent: A) water/TFA (100/0.1)
 B) acetonitrile/TFA (100/0.1)
 Gradient: 20-60%B (0-27 min), 90%B (27-35 min)

Flow rate: 0.4 mL/min
 Temperature: 70 °C
 Detection: UV at 220 nm
 Injection: 10 μ L (0.25 ~ 0.50 mg/mL)

Peptides and proteins

RP

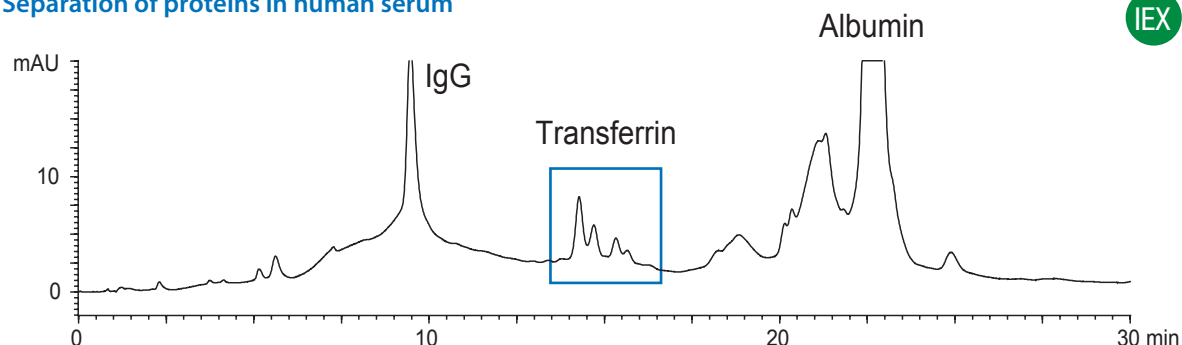


Column: Meteoric Core C18 BIO (2.7 μ m, 16 nm) 150 x 2.1 mm ID
 Part No.: CAW16SQ7-15Q1PT
 Eluent: A) water/TFA (100/0.1)
 B) acetonitrile/TFA (100/0.1)
 Gradient: 20-70% B (0-15 min), 70% B (15-17 min)

Flow rate: 0.2 mL/min
 Temperature: 40 °C
 Detection: UV at 220 nm
 Injection: 2 μ L (0.05-0.2 mg/mL)
 Pressure: 12.8-16.1 MPa (1860-2330 psi)

BioLC applications – Proteins

Separation of proteins in human serum

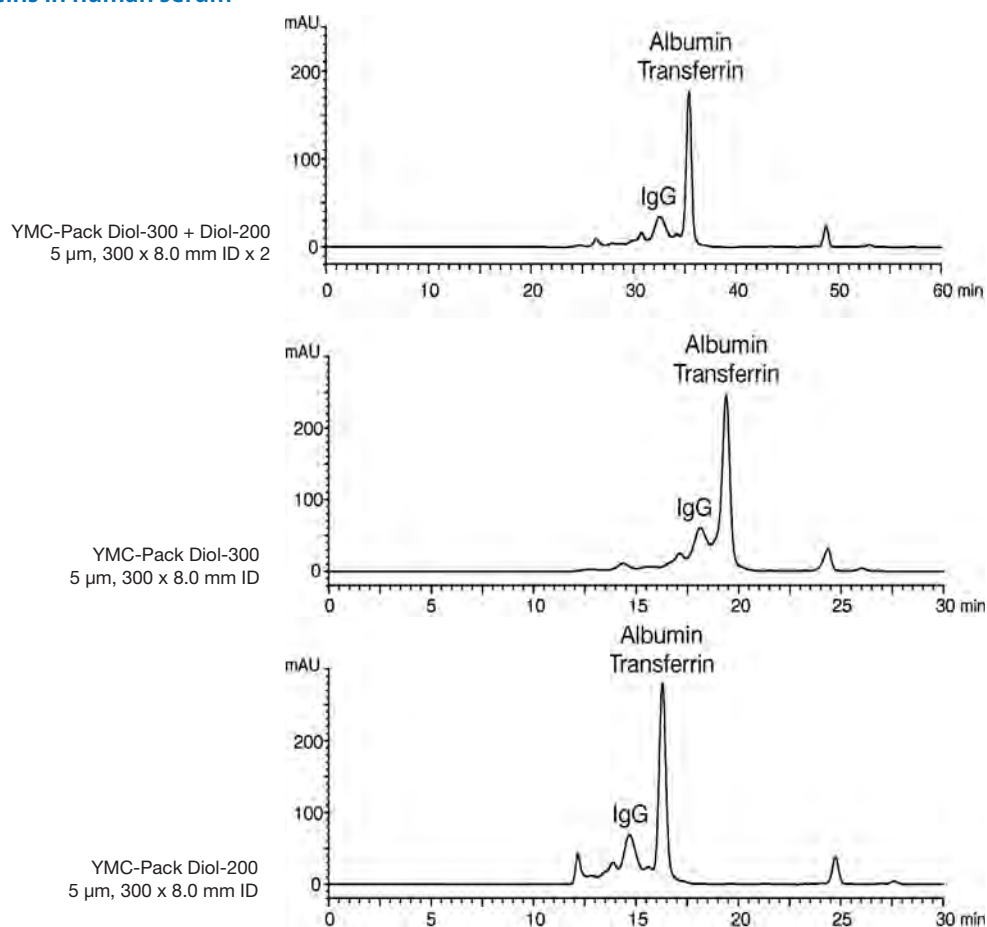


IEX

Column: BioPro IEX QA (5 μ m) 50 x 4.6 mm ID
 Part No.: QAA0S05-0546WP
 Eluent: A) 20 mM Tris-HCl (pH 8.6)
 B) 20 mM Tris-HCl (pH 8.6) containing 0.5 M NaCl
 Gradient: 0-30%B (0-15 min), 30-100%B (15-30 min)
 Flow rate: 0.5 mL/min

Temperature: 25 $^{\circ}$ C
 Detection: UV at 280 nm
 Injection: 20 μ L
 Sample: Human serum (100 μ L/mL)

Proteins in human serum



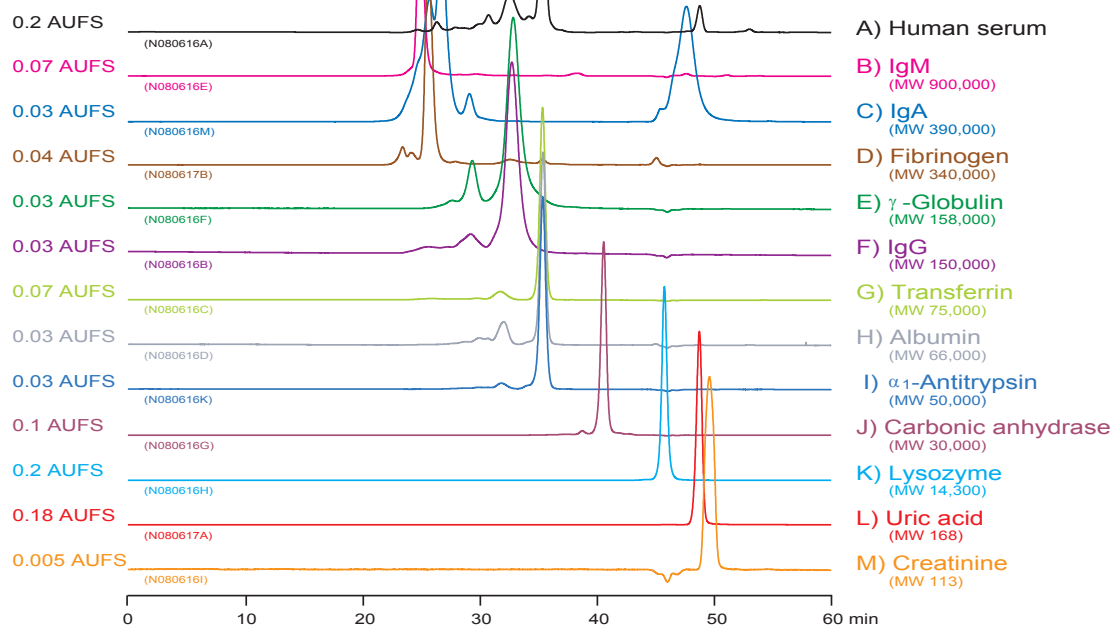
SEC

Eluent: 0.1 M KH_2PO_4 - K_2HPO_4 (pH 7.0) containing 0.2 M NaCl
 Part Nos.: DL30S05-3008WT + DL20S05-3008WT
 Flow rate: 0.5 mL/min
 Temperature: ambient (25 $^{\circ}$ C)
 Detection: UV at 280 nm
 Injection: 20 μ L
 Sample: Human serum (100 μ L/mL)

Plasma constituents

SEC

blood analysis

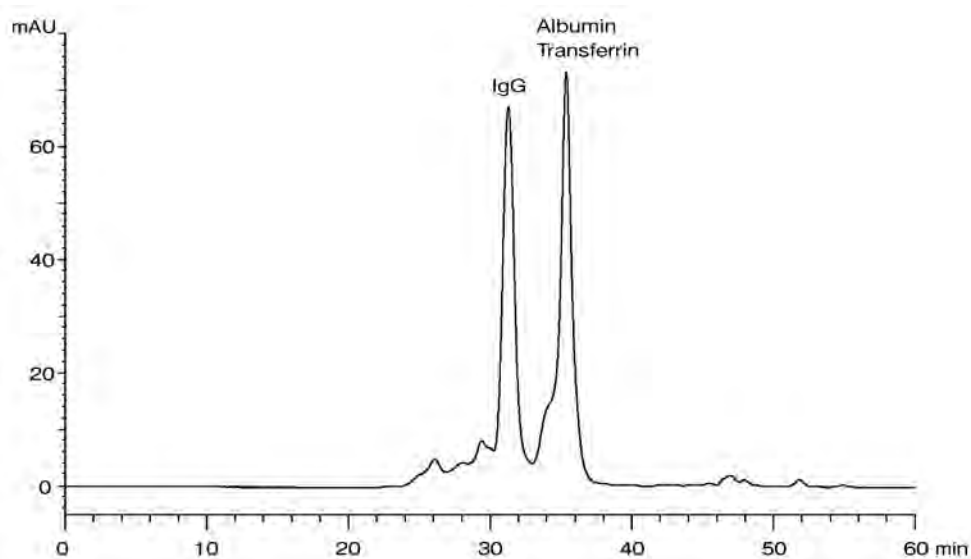


Columns: YMC-Pack Diol-300 + Diol-200 (5 μ m) 300 x 8.0 mm ID x 2
Part Nos.: DL30S05-3008WT + DL20S05-3008WT
Eluent: 0.1 M KH_2PO_4 - K_2HPO_4 (pH 7.0) containing 0.2 M NaCl
Flow rate: 0.5 mL/min

Temperature: ambient (25 $^{\circ}\text{C}$)
Detection: UV at 280 nm
Injection: 20 μ L (L: 1 μ L)
Sample: A) 100 μ L/mL; B-M) 1.0 mg/mL

Proteins in mouse ascites fluid

SEC



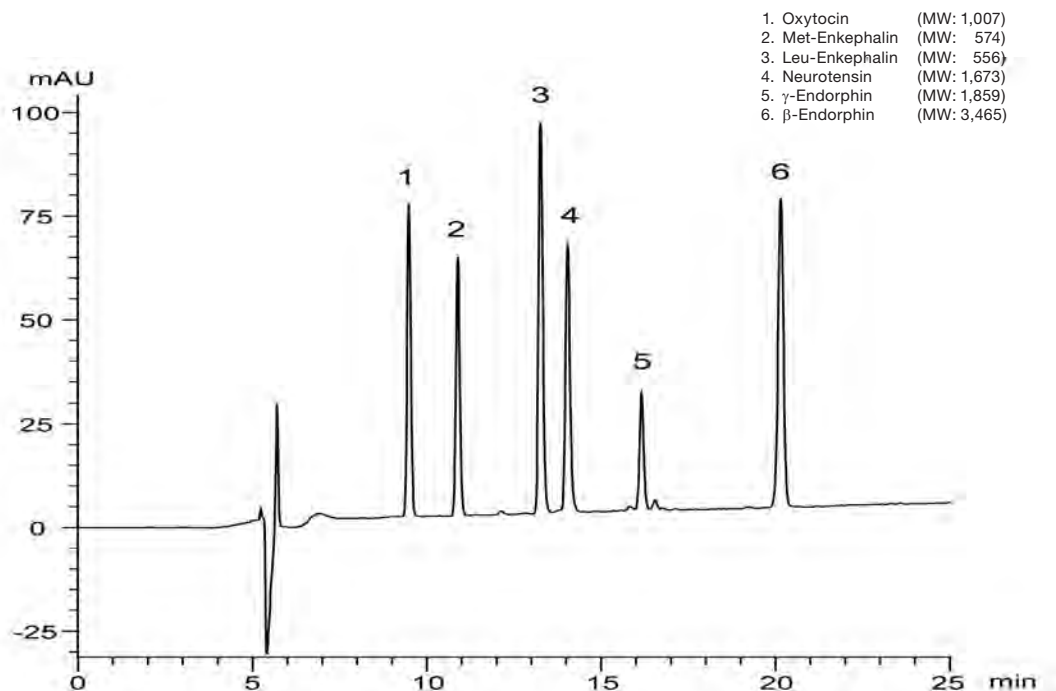
Columns: YMC-Pack Diol-300 + Diol-200 (5 μ m) 300 x 4.6 mm ID
Part Nos.: DL30S05-3046WT + DL20S05-3046WT
Eluent: 0.1 M KH_2PO_4 - K_2HPO_4 (pH 7.0)
Flow rate: 0.17 mL/min

Temperature: ambient (25 $^{\circ}\text{C}$)
Detection: UV at 220 nm
Injection: 10 μ L (60 times dilution with water)

BioLC applications – Peptides

Peptides

RP

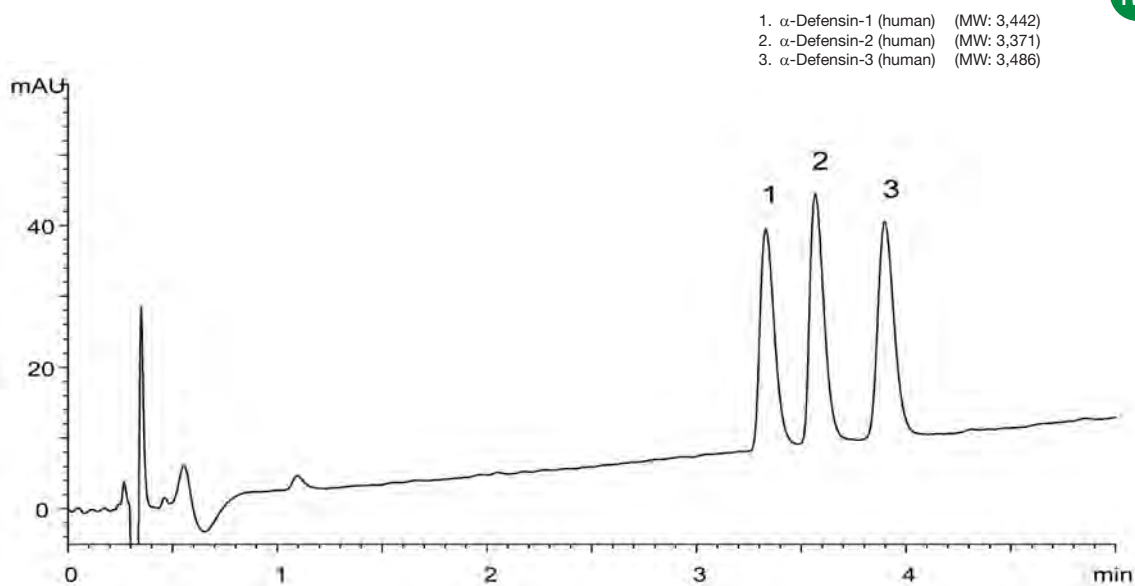


Column: YMC-Triart C18 (5 μ m, 12 nm) 150 x 2.0 mm ID
 Part No.: TA12S05-1502WT
 Eluent: A) water / TFA (100/0.1)
 B) acetonitrile / TFA (100/0.1)
 Gradient: 20%-45% B (0-25 min)

Flow rate: 0.2 mL/min
 Temperature: 37 °C
 Detection: UV at 220 nm
 Injection: 2 μ L (0.075 $\approx$ 0.25 mg/mL)

Antimicrobial peptides

RP

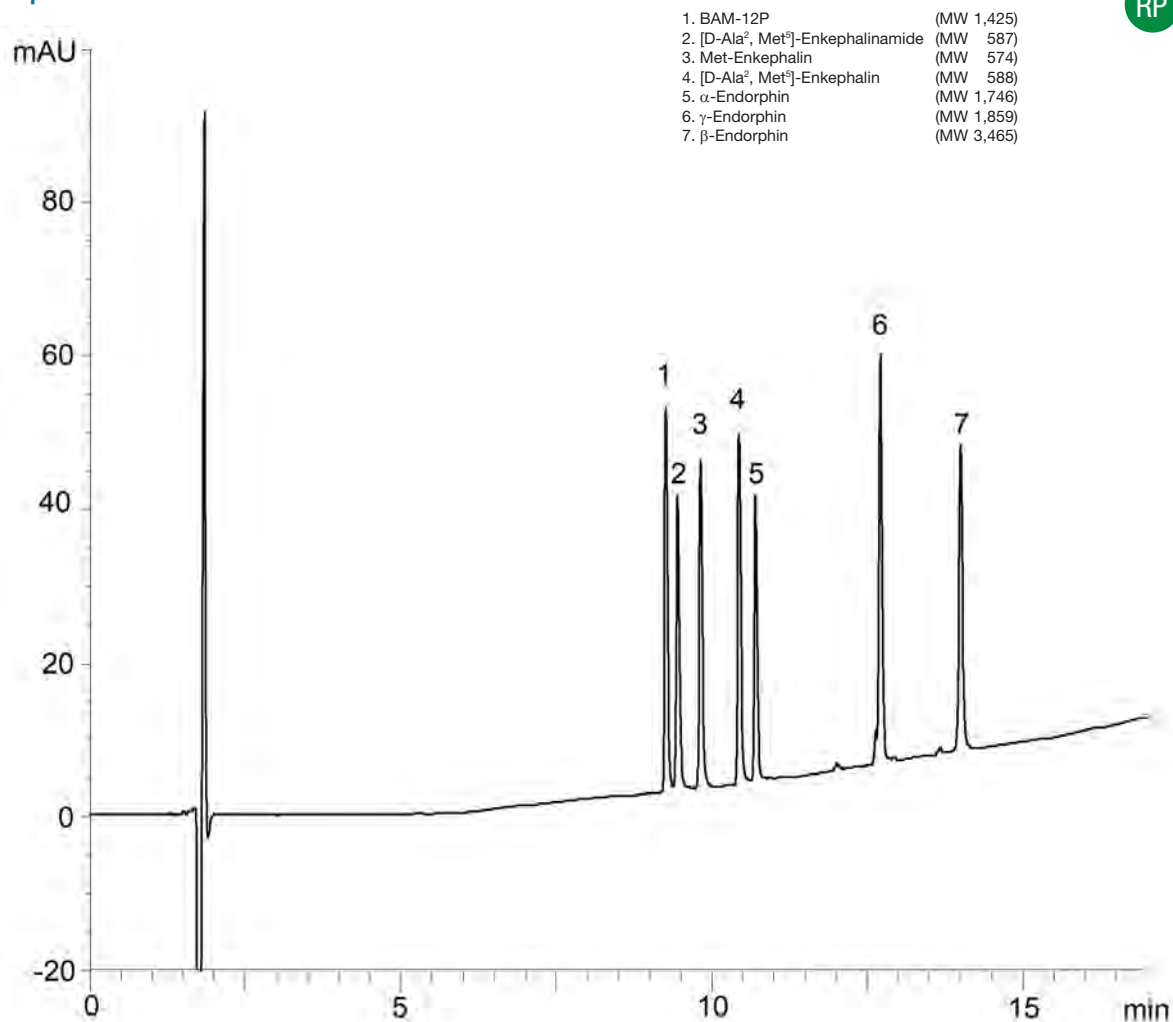


Column: YMC-Triart C18 (1.9 μ m, 12 nm) 50 x 2.0 mm ID
 Part No.: TA12SP9-0502PT
 Eluent: A) water / formic acid (100/0.1)
 B) 2-propanol / acetonitrile / formic acid (50/50/0.08)
 Gradient: 10%-25% B (0-10 min)

Flow rate: 0.4 mL/min
 Temperature: 70 °C
 Detection: UV at 220 nm
 Injection: 1 μ L (50 μ g/mL)

Peptides

RP

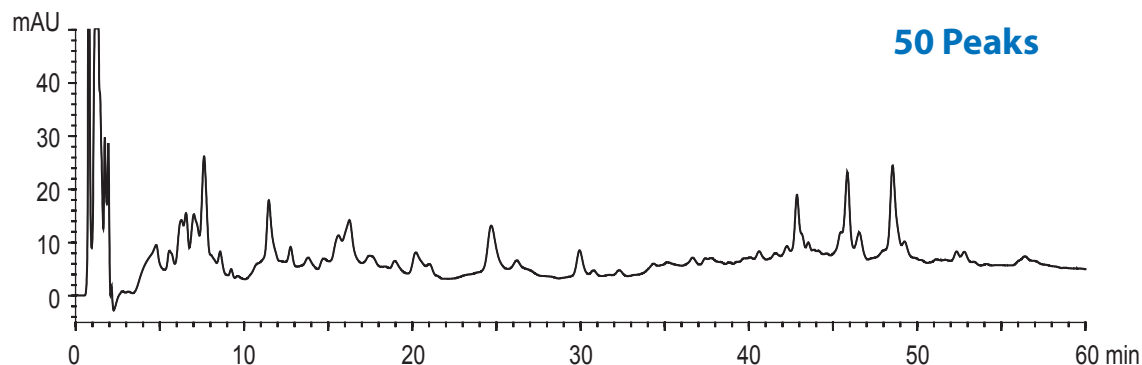


Column: Meteoric Core C18 BIO (2.7 μm, 16 nm) 150 × 2.1 mm ID
 Part No.: CAW16SQ7-15Q1PT
 Eluent: A) water/TFA (100/0.1)
 B) acetonitrile/TFA (100/0.1)
 15-55% B (0-15 min), 55% B (15-17 min)
 Flow rate: 0.2 mL/min
 Temperature: 40 °C
 Detection: UV at 220 nm
 Injection: 2 μL (0.02-0.5 mg/mL)
 Pressure: 14.9-16.1 MPa (2160-2330 psi)

BioLC applications – Peptide Mapping

Peptide mapping of tryptic digests of BSA with highest sensitivity

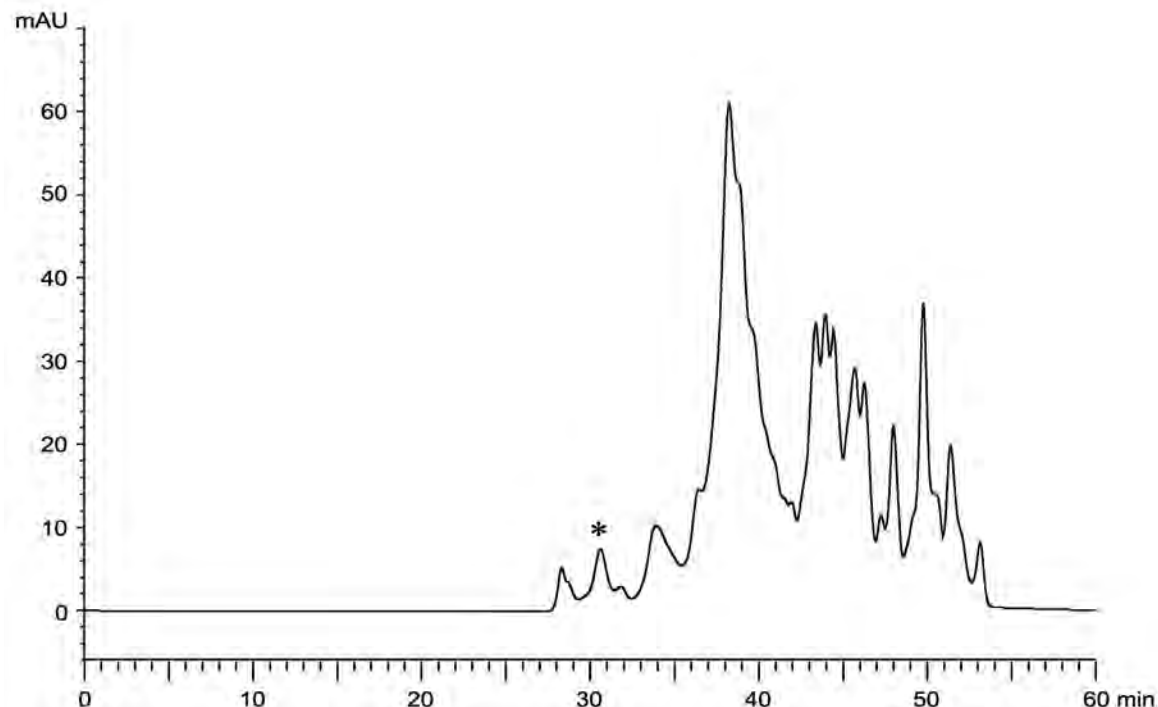
IEX



Column: BioPro IEX QA (5 μ m) 50 x 4.6 mm ID
Part No.: QAA0S05-0546WP
Eluent: A) 20 mM Tris-HCl (pH 8.6)
B) 20 mM Tris-HCl (pH 8.6) containing 0.5 M NaCl
Gradient: 0-15%B (0-30 min), 15-60%B (30-60 min)
Flow rate: 0.5 mL/min
Temperature: 25 °C
Detection: UV at 220 nm
Injection: 20 μ L
Sample: Tryptic digest of BSA

Tryptic digest of BSA

SEC

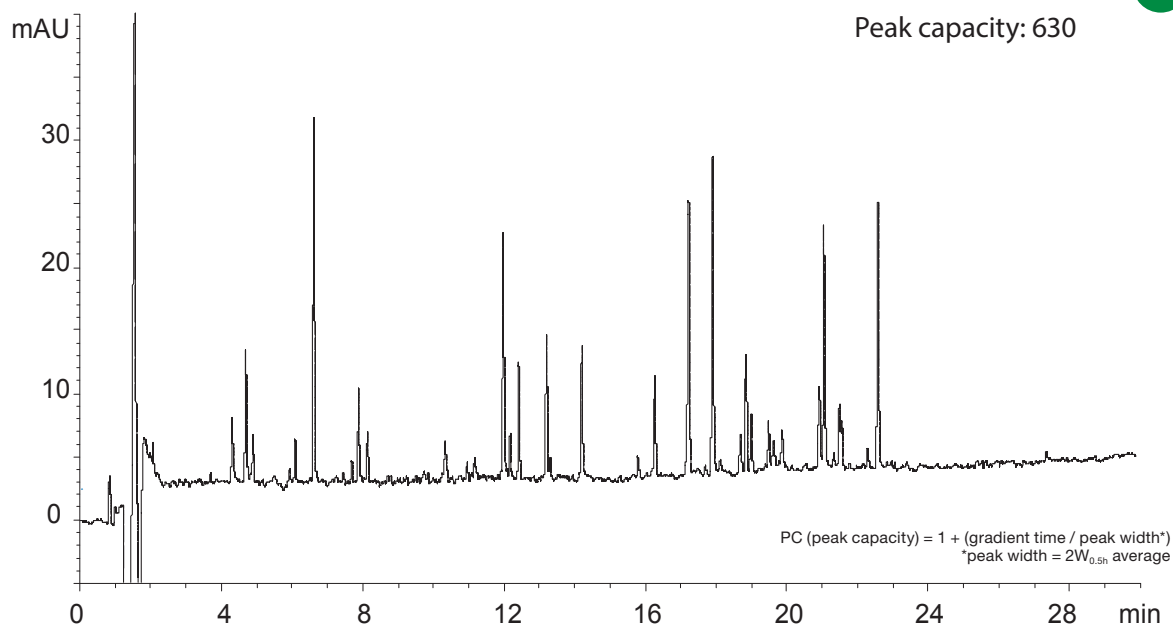


Columns: YMC-Pack Diol-120 + Diol-60 (5 μ m) 500 x 8.0 mm ID
Part Nos.: DL12S05-5008WT + DL06S05-5008WT
Eluent: 0.1 M KH_2PO_4 - K_2HPO_4 (pH 7.0)
containing 0.2 M NaCl/acetonitrile (70/30)
Flow rate: 0.7 mL/min
Temperature: ambient (25 °C)
Detection: UV at 220 nm
Injection: 5 μ L
Sample: Tryptic digest of BSA

*undigested BSA

Tryptic digest of Hemoglobin

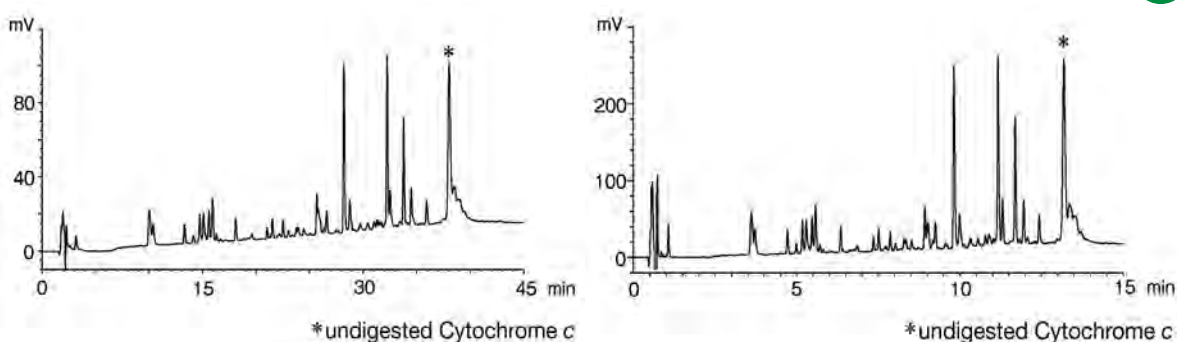
RP



Column: YMC-Triart C18 (1.9 μm , 12 nm) 200 x 2.0 mm ID (Two coupled 100 x 2.0 mm ID)
 Part No.: TA12SP9-1002PT (2x)
 Eluent: A) water/TFA (100/0.1)
 B) acetonitrile/TFA (100/0.08)
 Gradient: 5–40%B (0–30 min)
 Flow rate: 0.4 mL/min
 Temperature: 70 °C
 Detection: UV at 220 nm
 Injection: 20 μL
 Sample: Tryptic digest of Bovine Hemoglobin (2.5 nmol/mL)
 Pressure: 58.1–61.6 MPa (8,430–8,930 psi)

Peptide mapping – excellent reproducibility between 5 μm and 2 μm

RP



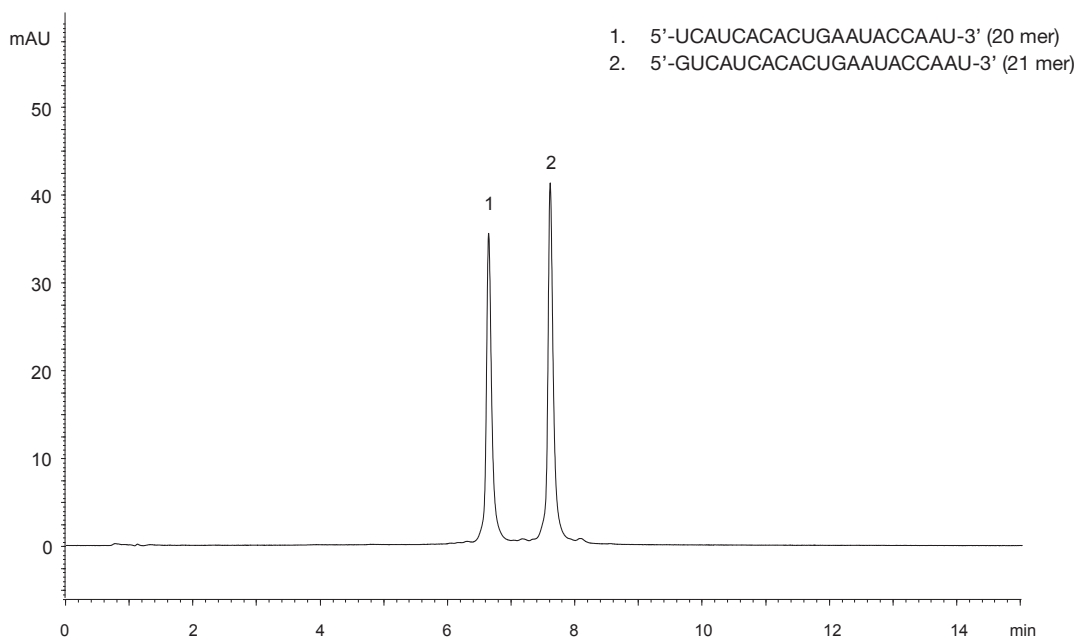
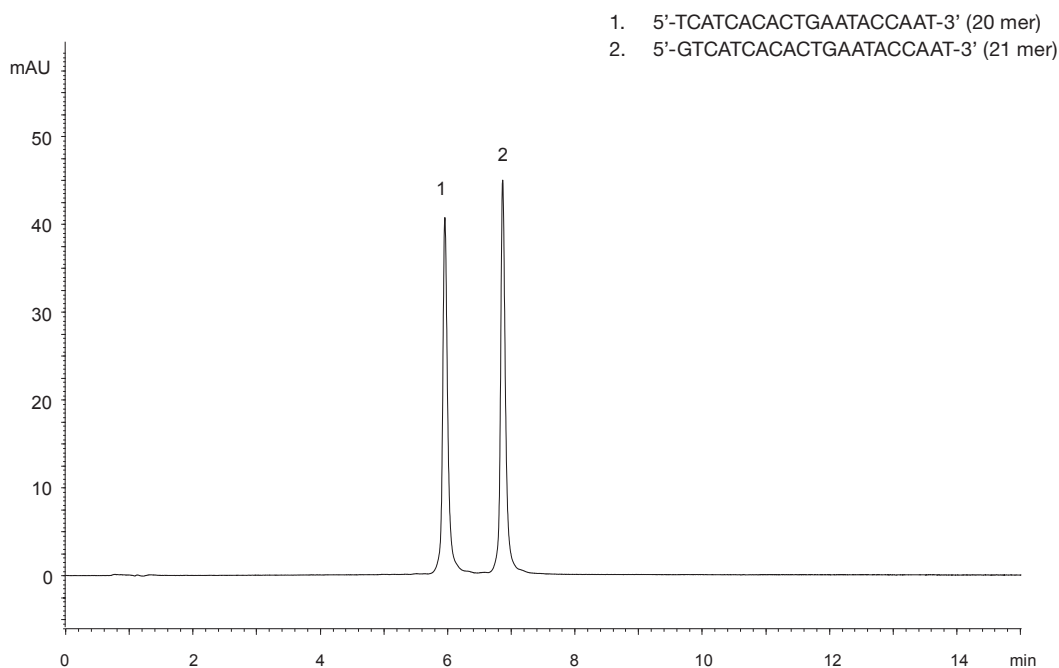
Column: YMC-Pack Pro C18 (5 μm , 12 nm) 150 x 2.0 mm ID
 Part No.: AS12S05-1502WT
 Eluent: A) acetonitrile/water/trifluoroacetic acid (10/90/0.1)
 B) acetonitrile/water/trifluoroacetic acid (35/65/0.1)
 Gradient: Time A (in %) B (in %)
 0 100 0
 5 100 0
 40 0 100
 45 0 100
 Flow rate: 0.2 mL/min
 Temperature: 37 °C
 Detection: UV at 220 nm
 Injection: 1 μL
 Sample: Tryptic digest of Cytochrome c

Column: YMC-UltraHT Pro C18 (2 μm , 12 nm) 50 x 2.0 mm ID
 Part No.: AS12S02-0502WT
 Eluent: A) acetonitrile/water/trifluoroacetic acid (10/90/0.1)
 B) acetonitrile/water/trifluoroacetic acid (35/65/0.1)
 Gradient: Time A (in %) B (in %)
 0 100 0
 1.65 100 0
 13.35 0 100
 15.00 0 100
 Flow rate: 0.2 mL/min
 Temperature: 37 °C
 Detection: UV at 220 nm
 Injection: 1 μL
 Sample: Tryptic digest of Cytochrome c

BioLC applications – Oligonucleotides

Separation of synthetic oligonucleotides (single-strand DNA)

IEX

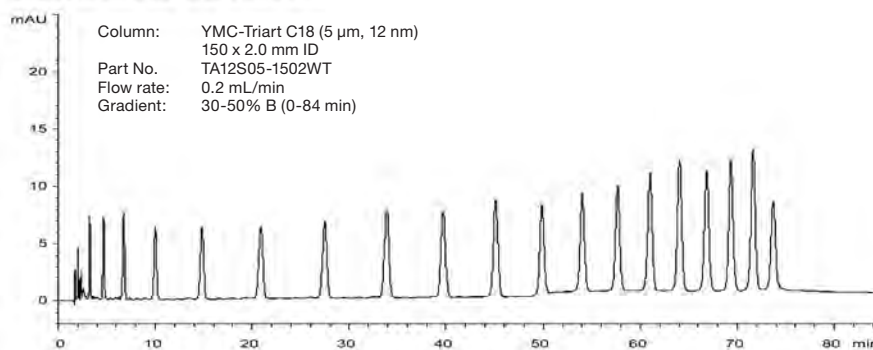


Column: BioPro IEX QF (5 μ m) 100 x 4.6 mm ID
 Part no.: QF00S05-1046WP
 Eluent: A) 10 mM NaOH
 B) 10 mM NaOH containing 1.0 M NaClO₄
 Gradient: 25-55%B (0-15 min), 100%B (15-20 min)
 Flow rate: 1.0 mL/min
 Temperature: 25 °C
 Detection: UV at 260 nm
 Injection: 4 μ L (5 nmol/L)

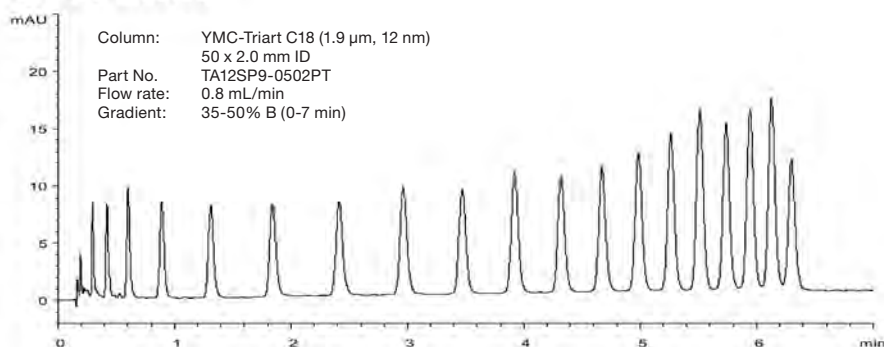
Oligonucleotides d(T)2-20 method transfer from HPLC to UHPLC

RP

Conventional LC method



UHPLC method



80 min

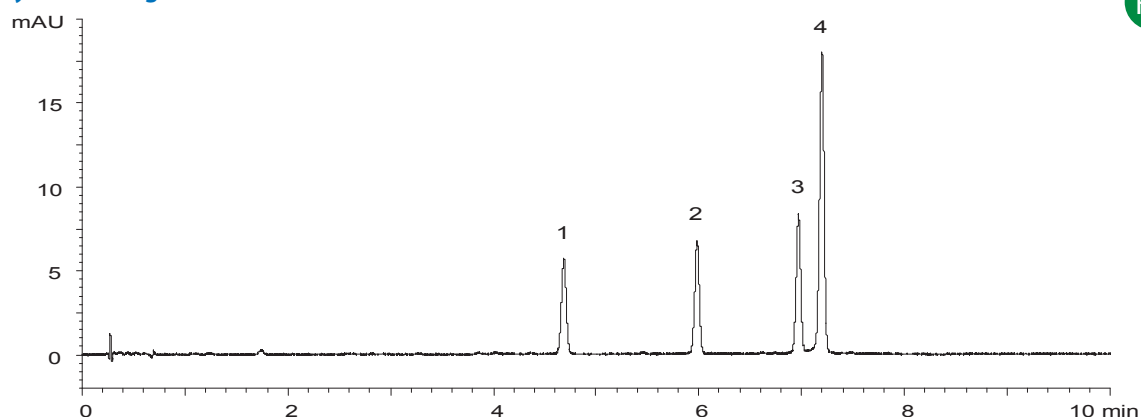
11x
faster

7 min

Eluent: A) 10 mM di-n-butylamine-acetic acid (pH 6.0)
B) methanol
Detection: UV at 269 nm
Injection: 1 µL (5 nmol/mL)
Temperature: 37 °C

Synthetic oligonucleotides

RP



1. 5'-CAC UGA AUA CCA AU-3' (14mer)

2. 5'-UCA CAC UGA AUA CCA AU-3' (17mer)

3. 5'-UCA UCA CAC UGA AUA CCA AU-3' (20mer)

4. 5'-GUC AUC ACA CUG AAU ACC AAU-3' (21mer)

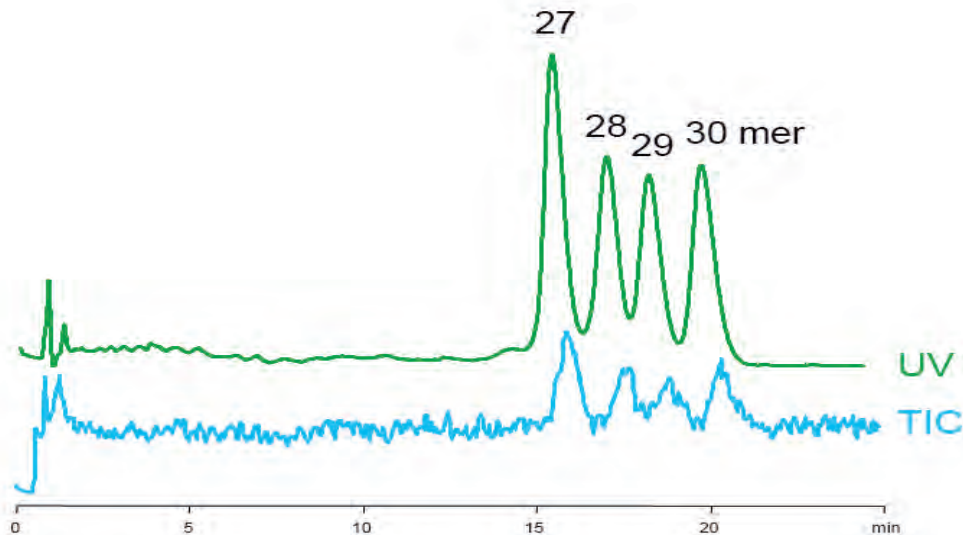
Column: YMC-Triart C18 (1.9 µm, 12 nm) 50 x 2.1 mm ID
Part No.: TA12SP9-05Q1PT
Eluent: A) 200 mM HFIP*-8 mM triethylamine
B) methanol
Gradient: 10-20%B (0-10 min)

*hexafluoroisopropanol
Flow rate: 0.42 mL/min
Temperature: 65 °C
Detection: UV at 260 nm
Injection: 1 µL (2-4 nmol/mL)

BioLC applications – Oligonucleotides

LC-MS analysis of synthetic 27-30 mer oligonucleotides

RP



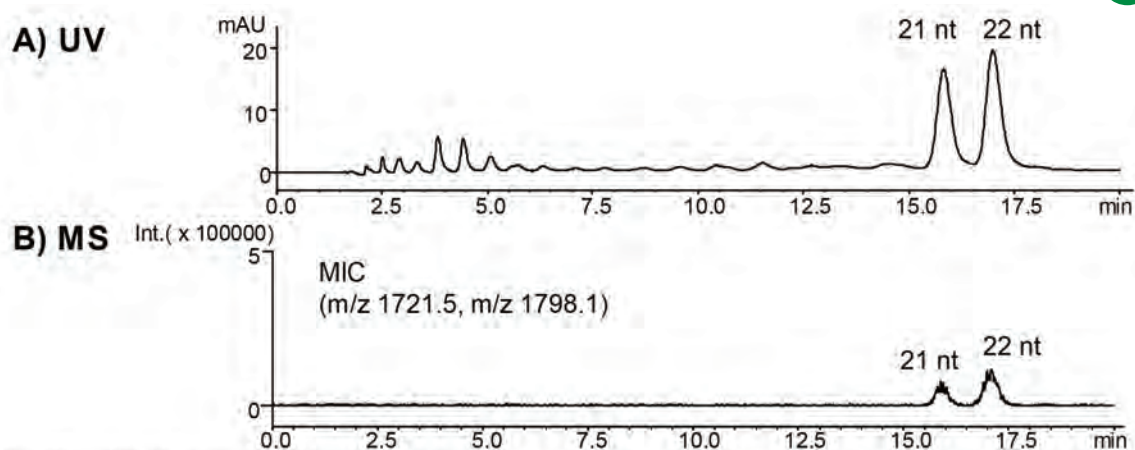
Sample: Primer of DNA sequencing

5'-CCGCTCGAGCTAAAAAAGCCTGTGTACC-3' (30 mer)

Column: Hydrosphere C18 (3 μ m) 50 x 2.0 mm ID
 Part No.: HS12S03-0502WT
 Eluent: A) 10 mM DBAA (pH 6.0)
 B) Mobile phase A / acetonitrile (50/50)
 Gradient: 58%-62% B (0-20 min), 62% B (20-25 min)
 Flow rate: 0.2 mL/min
 Temperature: 35 $^{\circ}$ C
 Detection: UV at 269 nm and ESI negative-mode
 Injection: 1 μ L (10 pmol/component)

LC/MS analysis of miRNA

RP

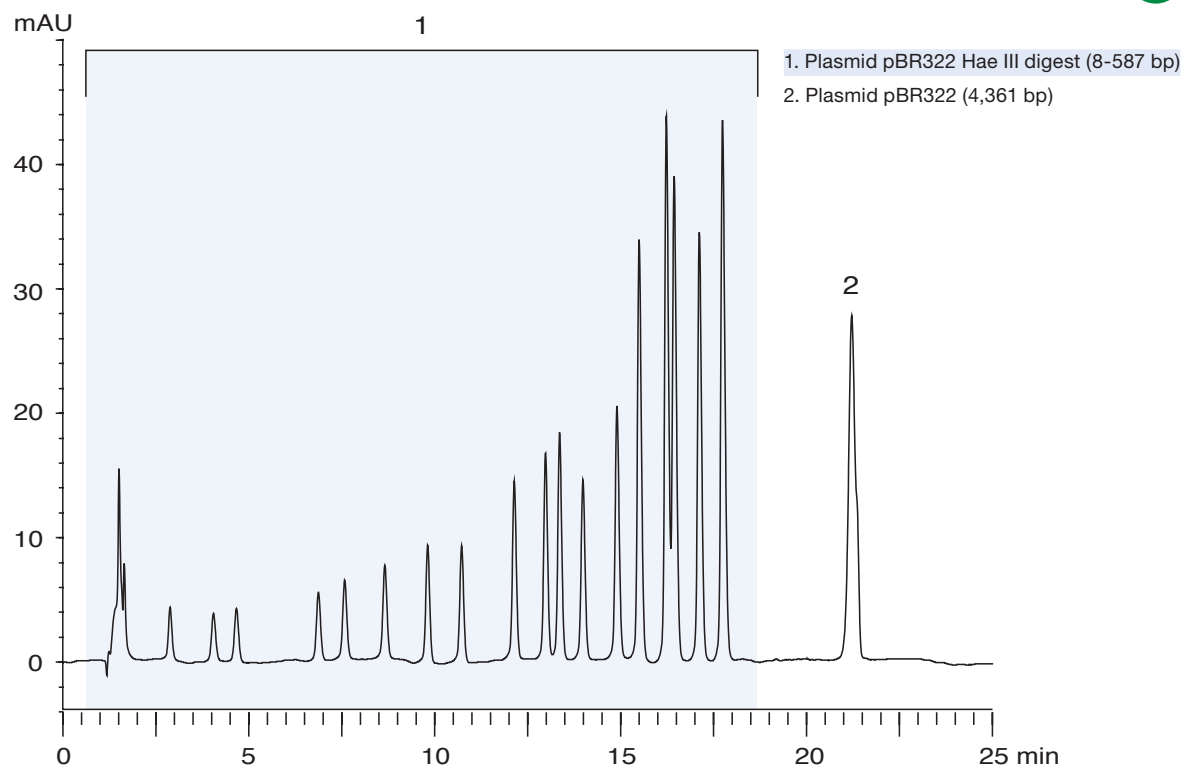


Courtesy of M. Yamada, SHIMADZU CORPORATION

5'-pUGG AGU GUG ACA AUG GUG UUG-3' (21 nt, MW 6890.1)
 5'-pUGG AGU GUG ACA AUG GUG UUG U-3' (22 nt, MW 7196.3)

Column: YMC-Triart C18 (3 μ m, 12 nm) 150 x 2.0 mm ID
 Part No.: TA12S03-1502WT
 Eluent: A) 10 mM di-n-butylamine-acetic acid (pH 7.5)
 B) 10 mM di-n-butylamine-acetic acid (pH 7.5)/acetonitrile (50/50)
 Gradient: 62-72%B (0-20 min)
 Flow rate: 0.2 mL/min
 Temperature: 30 $^{\circ}$ C
 Detection: A) UV at 260 nm
 B) ESI-negative mode
 Injection: 4 μ L (5 nmol/mL)
 System: LC) Shimadzu Prominence
 MS) Shimadzu LCMS2020

High resolution analysis using non-porous BioPro IEX QF for fragment identification



Column: BioPro IEX QF (5 μ m) 100 x 4.6 mm ID
Part No.: QF00S05-1046WP
Eluent: A) 20 mM Tris-HCl (pH 8.1)
B) 20 mM Tris-HCl (pH 8.1) containing 1.0 M NaCl
Gradient: 70-85%B (0-20 min), 85%B (20-25 min)
Flow rate: 0.5 mL/min

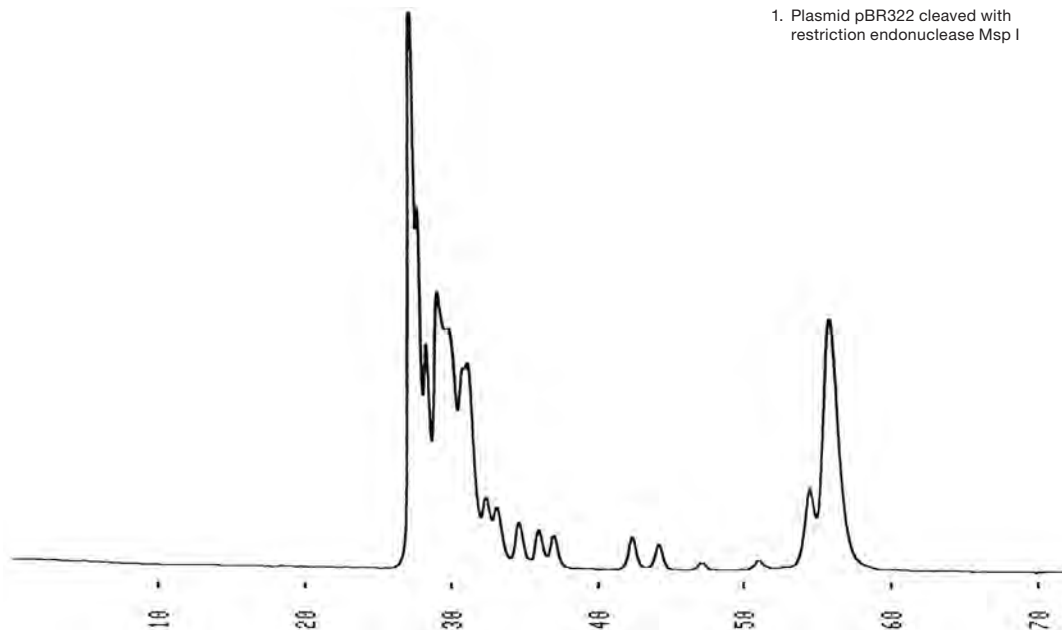
Temperature: 35 °C
Detection: UV at 260 nm
Injection: 10 μ L
Sample: Plasmid pBR322 Hae III digest (0.13 mg/mL)
Plasmid pBR322 (0.03 mg/mL)

BioLC applications – Plasmids

Plasmid pBR322 restriction fragment

SEC

1. Plasmid pBR322 cleaved with restriction endonuclease Msp I



Columns: YMC-Pack Diol-300 + Diol-200 (5 µm) 500 x 8.0 mm ID
Part Nos.: DL30S05-5008WT + DL20S05-5008WT
Eluent: 0.1M KH_2PO_4 - K_2HPO_4 (pH 7.0) containing 0.2M NaCl
Flow rate: 0.7 mL/min

Temperature: ambient (26 °C)
Detection: UV at 260 nm, 0.01 AUFS
Injection: 3 µL (0.49 mg/mL)
Sample: Plasmid pBR322 cleaved with restriction endonuclease Msp I

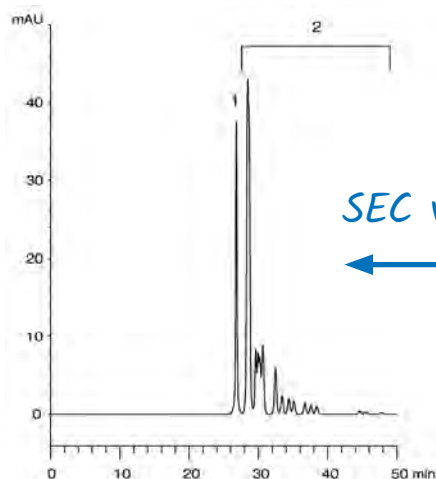
Plasmid pBR322 restriction and pBR322 Hae III restriction fragment

SEC

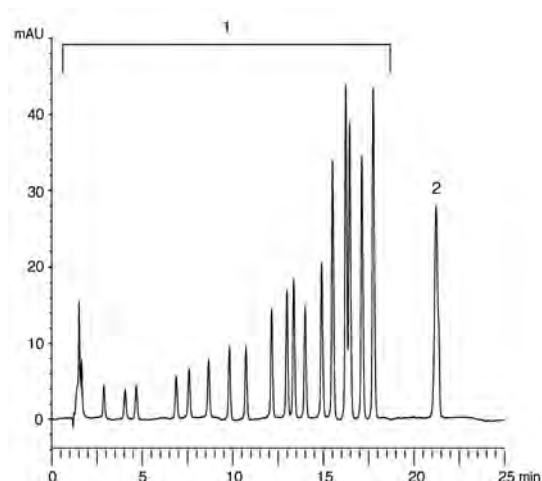
IEX

1. Plasmid pBR322 (4,361 bp)
2. Plasmid pBR322 Hae III digest (8-587 bp)

1. Plasmid pBR322 Hae III digest (8-587 bp)
2. Plasmid pBR322 (4,361 bp)

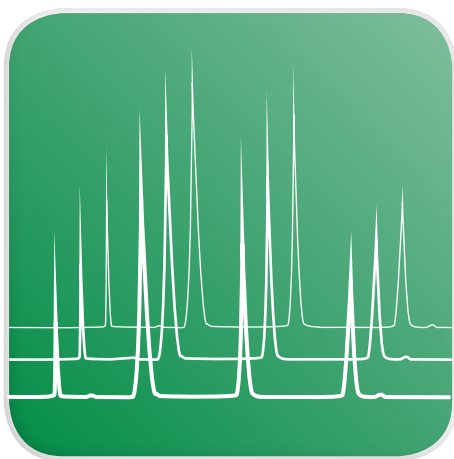


SEC vs. IEX!

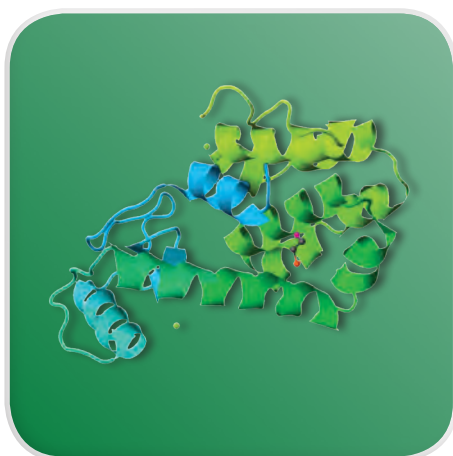


Columns: YMC-Pack Diol-300 + Diol-200 (5 µm) 500 x 8.0 mm ID
Part Nos.: DL30S05-5008WT + DL20S05-5008WT
Eluent: 0.1 M KH_2PO_4 - K_2HPO_4 (pH 7.0) containing 0.2 M NaCl
Flow rate: 0.7 mL/min
Temperature: ambient (25 °C)
Detection: UV at 260 nm
Injection: 10 µL

Column: BioPro IEX QF (5 µm) 100 x 4.6 mm ID
Part No.: QF00S05-1046WP
Eluent: A) 20 mM Tris-HCl (pH 8.1)
B) 20 mM Tris-HCl (pH 8.1) containing 0.1 M NaCl
Gradient: 70-85% B (0-20 min), 85% B (20-25 min)
Flow rate: 0.5 mL/min
Temperature: 35 °C
Detection: UV at 260 nm
Injection: 10 µL



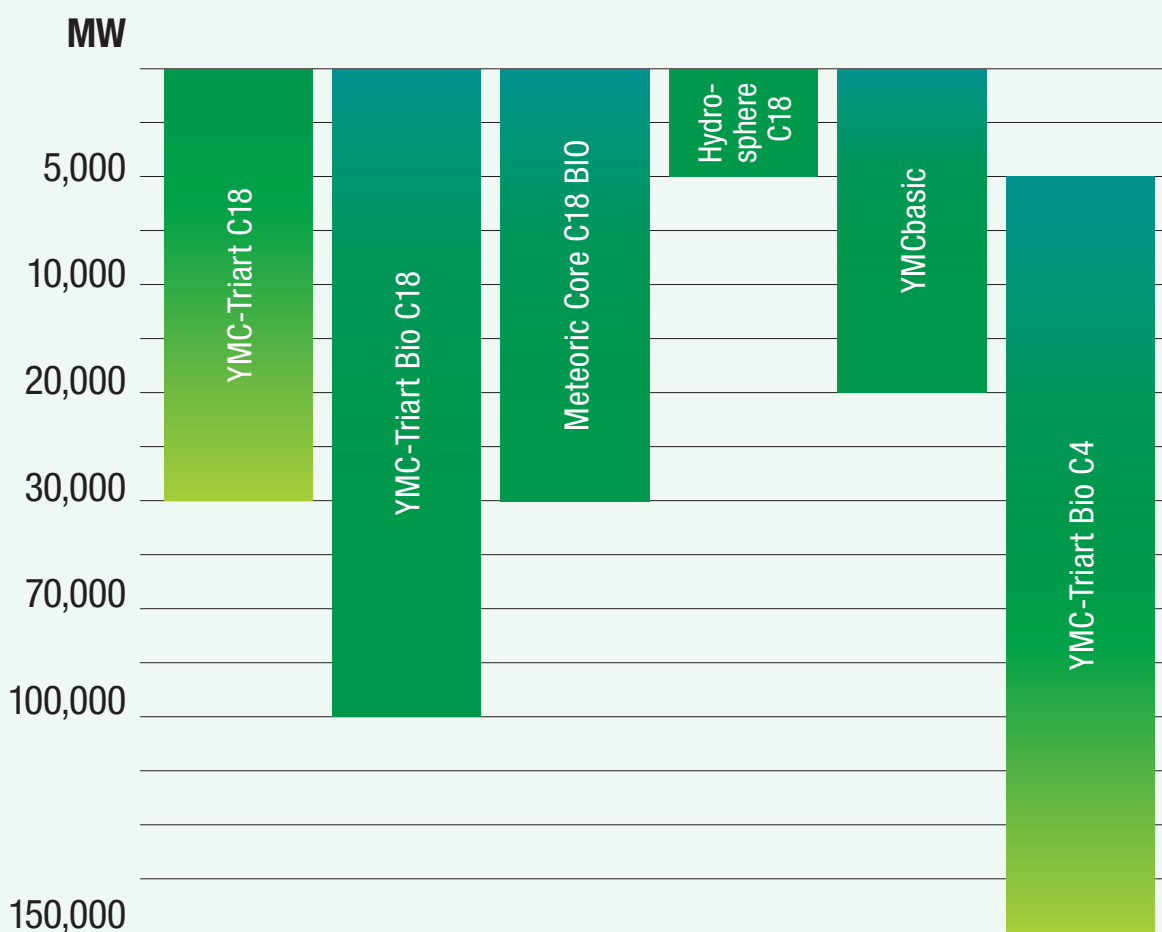
RP



RP – Bioseparation Columns

- Applicable to proteins, antibodies, peptides and oligonucleotides
- Selection of C18, C8 and C4 columns
- For UHPLC and HPLC
- pH- and temperature stable phases
- Superior reproducibility

Column Selection Tool according to molecular weight



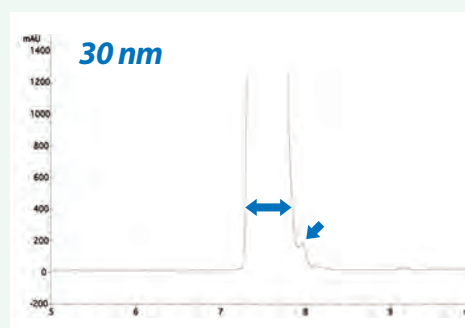
- most appropriate MW range
- extended MW range by elevated temperature
- appropriate MW range

Influence of pore size

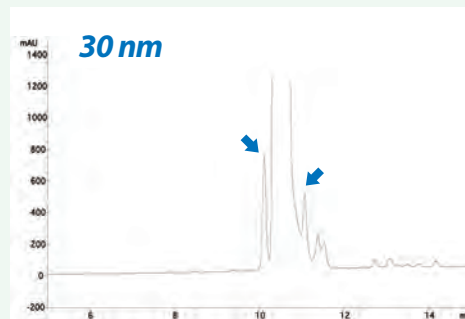
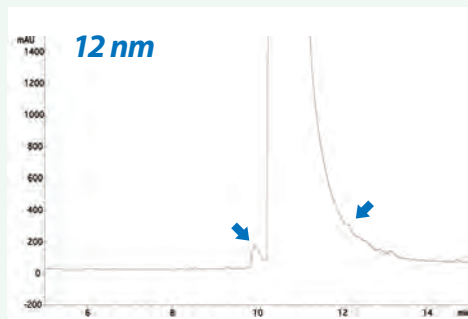
As shown in the table (on the left), the C18 column with 12 nm pore size is suitable for small peptides up to a MW of 5,000 Da. The highest efficiency for large peptides or small proteins can be obtained by using a wide pore C8 phase with 20 nm porosity. Most proteins can be eluted efficiently with a wide pore C4 column with 30 nm porosity.

However, the separation may also be influenced by the hydrophobicity of the peptide/protein and the nature of the column's bonded phase. Therefore, for initial method development, it can be useful, in the first instance, to follow the arrow shown in the **Column Selection Tool** for method optimisation.

Angiotensin II
(MW 1,046)



BSA
(MW 67,000)



For smaller peptides a small pore size is more successful. Larger molecules are separated much better with larger pore sizes!

RP – UHPLC / HPLC Selectivities

C4 and C8 selectivities for proteins (and peptides) and antibodies

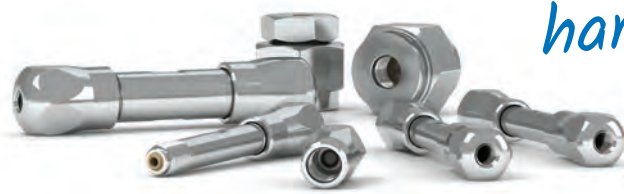
	YMC-Triart Bio C4	YMC basic
Modification	C4 (USP L26)	C8 (USP L7)
Particle Size / μm	1.9, 3, 5	3, 5
Pore Size / nm	30	20
pH range	1.0 – 10.0	2.0 – 7.5
Temperature range	pH < 7: 90 °C pH > 7: 50 °C	50 °C

C18 selectivities for peptides (and proteins)

	YMC-Triart C18	YMC-Triart Bio C18	Meteoric Core Bio C18
Modification	C18 (USP L1)	C18 (USP L1)	C18 (USP L1)
Particle Size / μm	1.9, 3, 5	1.9, 3, 5	2.7
Pore Size / nm	12	30	8 (16)
pH range	1.0 – 12.0	1.0 – 12.0	1.5 – 10.0
Temperature range	pH < 7: 90 °C pH > 7: 50 °C	pH < 7: 90 °C pH > 7: 50 °C	pH < 7: 70 °C pH > 7: 50 °C

C18 and C8 selectivities for oligonucleotides

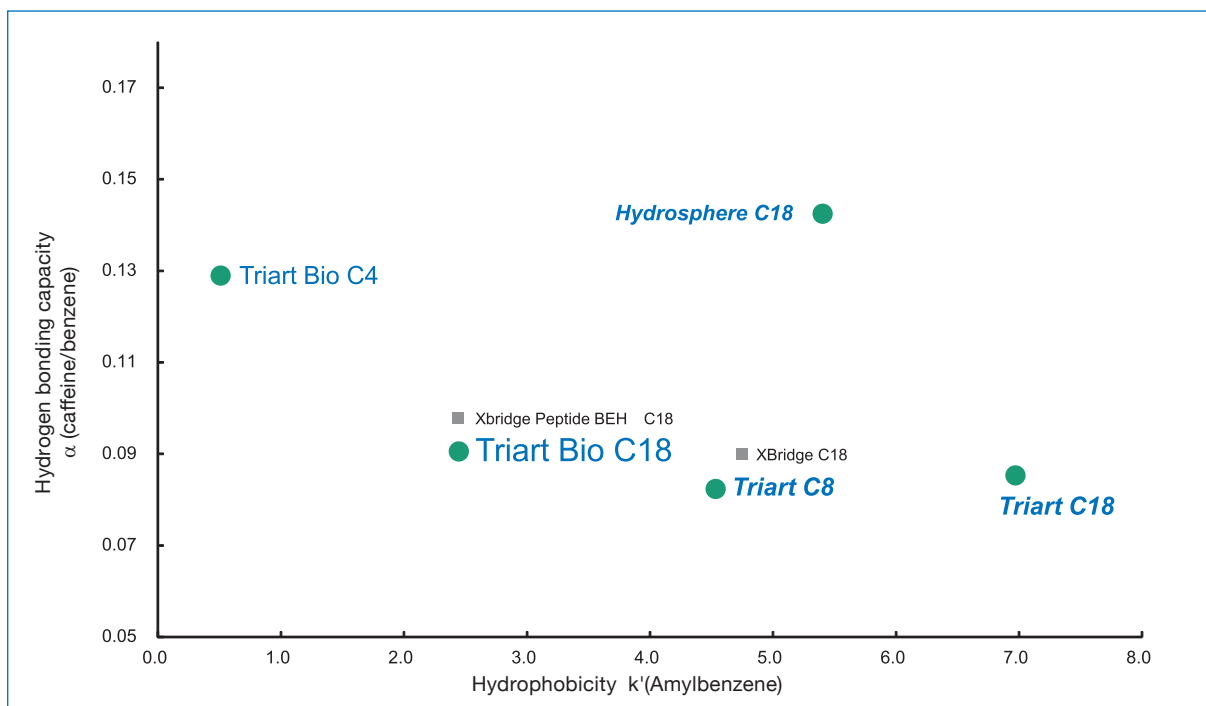
	YMC-Triart C18	YMC-Triart Bio C18	Hydrosphere C18	YMC-Triart C8
Modification	C18 (USP L1)	C18 (USP L1)	C18 (USP L1)	C8 (USP L7)
Particle Size / μm	1.9, 3, 5	1.9, 3, 5	2, 3, 5	1.9, 3, 5
Pore Size / nm	12	30	12	12
pH range	1.0 – 12.0	1.0 – 12.0	2.0 – 8.0	1.0 – 12.0
Temperature range	pH < 7: 90 °C pH > 7: 50 °C	pH < 7: 90 °C pH > 7: 50 °C	50 °C	pH < 7: 90 °C pH > 7: 50 °C



*Biocompatible
hardware available!*

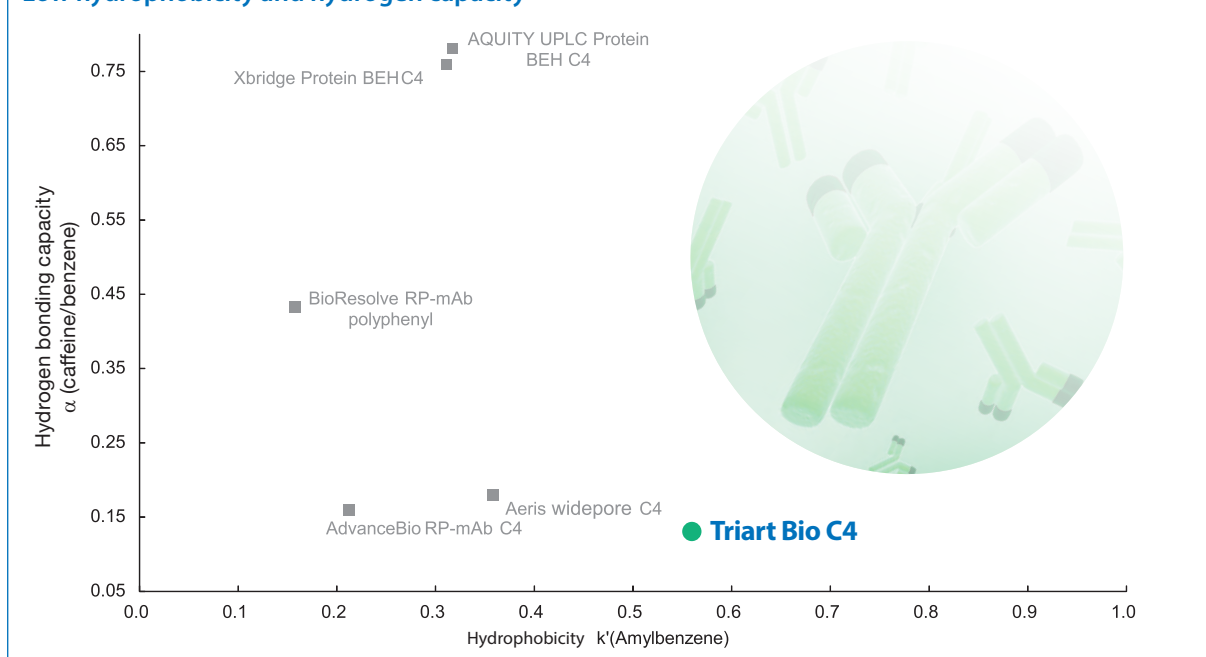
YMC-Triart metal-free columns are available for improved sensitivity and peak shape of coordinating compounds such as nucleotides or oligonucleotides, see page 47.

Ideal Selectivity for biomolecules



YMC-Triart Bio C18 shows a selectivity between that of YMC-Triart Bio C4 and regular YMC-TriartC18/C8 with 12 nm pore. This feature perfectly supplements the existing products.

Low hydrophobicity and hydrogen capacity



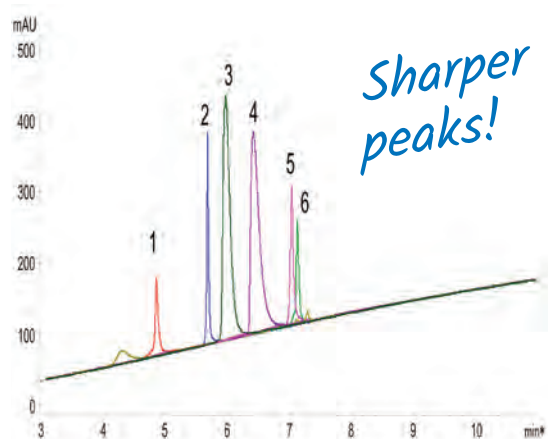
YMC-Triart Bio C4 is based on the robust, well-established organic/hybrid particle. It provides low hydrophobicity and lower hydrogen bonding capacity compared to other BioLC columns.

RP – YMC-Triart Bio C4: Sharper peaks

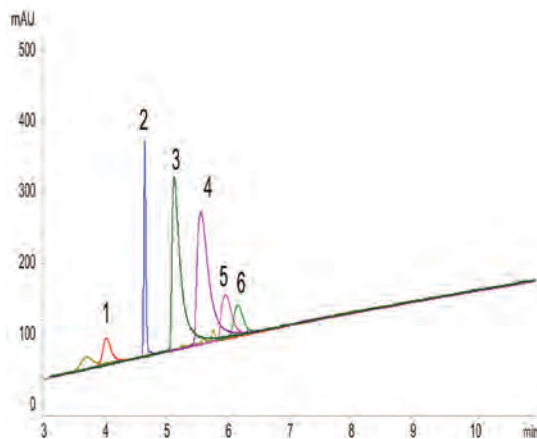
Better performance using YMC-Triart Bio C4

High sensitivity and sharp peaks under LC/MS compatible conditions

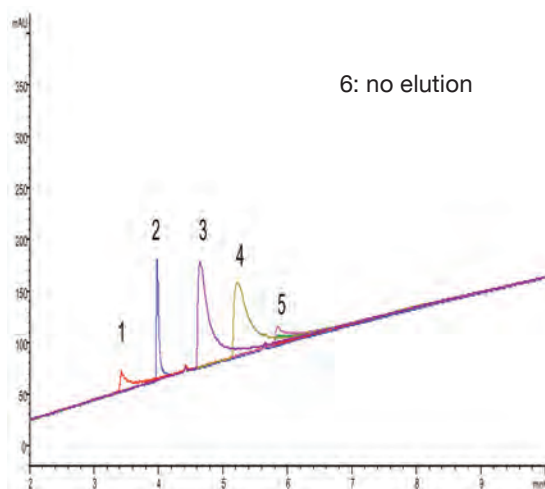
YMC-Triart Bio C4 (3 μ m, 30 nm)



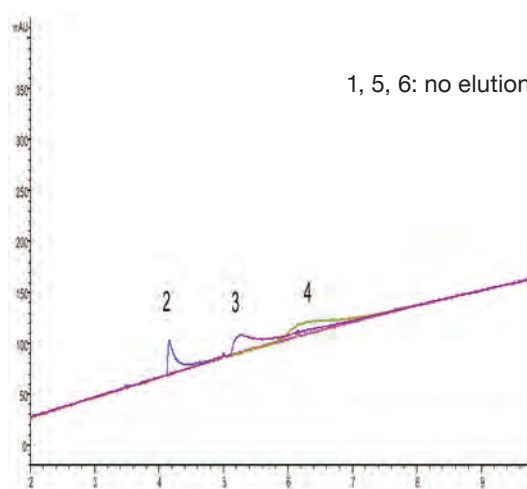
XBridge Protein BEH C4 (3.5 μ m, 30 nm)



AdvanceBio RP-mAb C4 (3.5 μ m, 45 nm)



Aeris widepore C4 (3.6 μ m, 20 nm)



Column: 150 x 3.0 mm ID
 Eluent: A) water/formic acid (100/0.1)
 B) acetonitrile/formic acid (100/0.1)
 Gradient: 10-95%B (0-15 min)
 Temperature: 40 °C
 Detection: UV at 220 nm

Sample:
 1. Cytochrome c (Horse heart)
 2. Insulin (Bovine pancreas)
 3. Transferrin (Human)
 4. BSA
 5. β -Lactoglobulin (Bovine)
 6. α -Chymotrypsinogen A (Bovine pancreas)

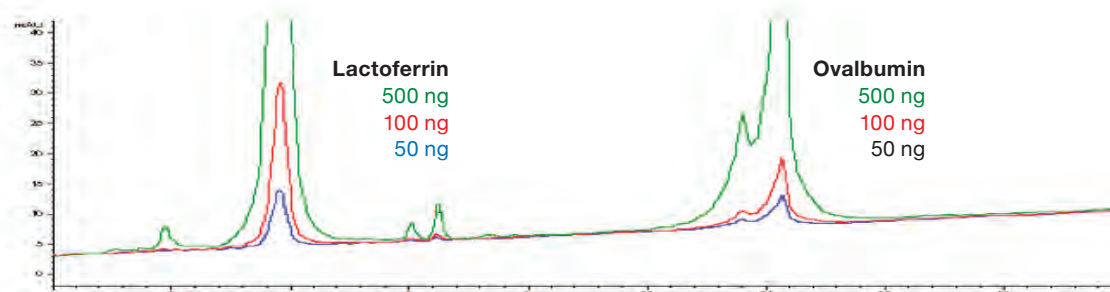
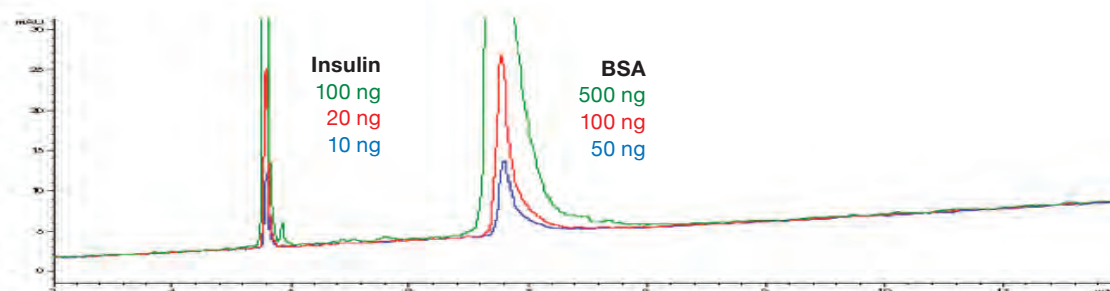
YMC-Triart Bio C4 shows better peak shape and recovery with a mobile phase containing formic acid, which is commonly used for LC/MS analysis. Therefore, YMC-Triart Bio C4 is ideal for high sensitivity analysis of proteins.

RP – YMC-Triart Bio C4: No column adsorption

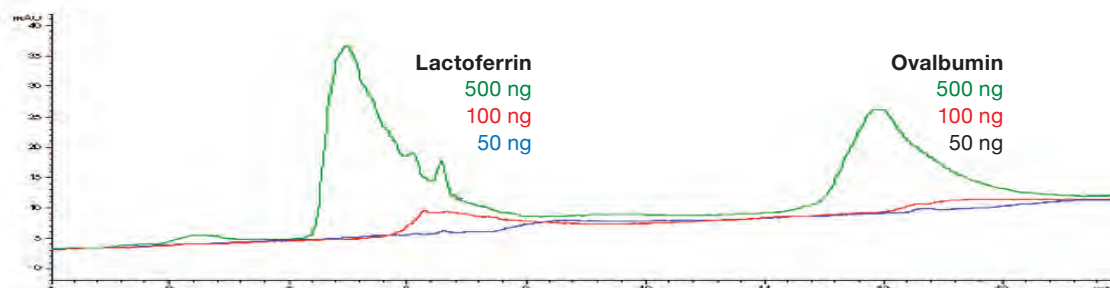
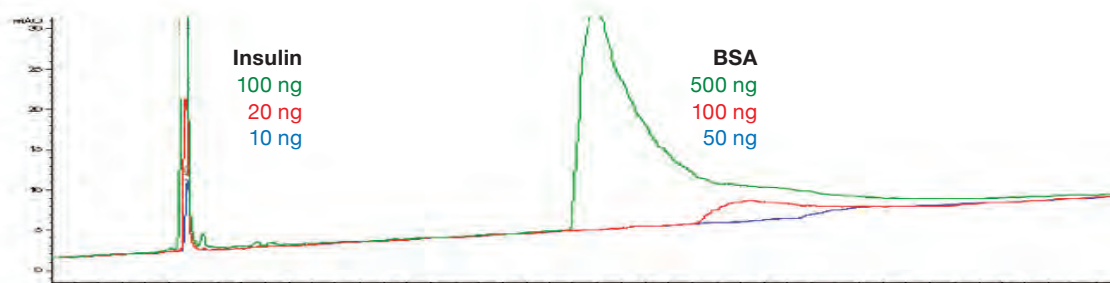
No sample adsorption by YMC-Triart Bio C4 columns

Ideal for Microanalysis

YMC-Triart Bio C4 (1.9 μ m, 30 nm)



Aeris widepore C4 (3.6 μ m, 20 nm)



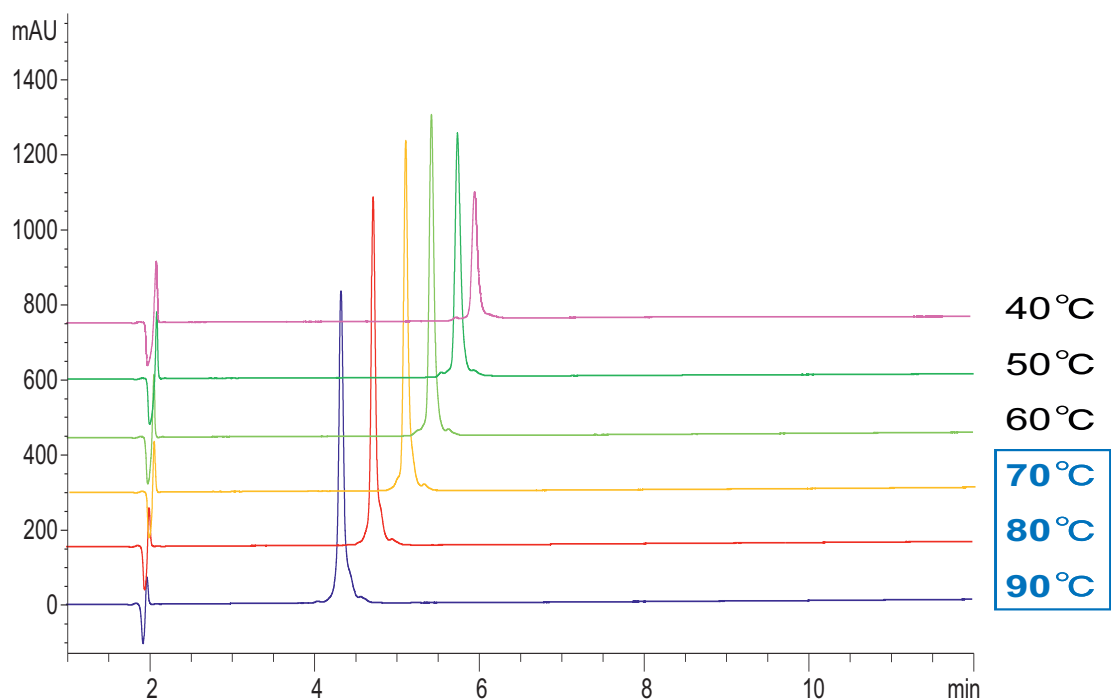
Column: 150 x 2.1 mm ID
Eluent: A) water/TFA (100/0.05)
B) acetonitrile/TFA (100/0.05)
Gradient: 25-60%B (0-15 min), 90%B (15-20 min), 25%B (20-35 min)
Detection: UV at 220 nm
Temperature: 40 °C

No sample adsorption was observed on YMC-Triart Bio C4 even at a low sample loading. This makes YMC-Triart Bio C4 ideal for microanalysis of proteins.

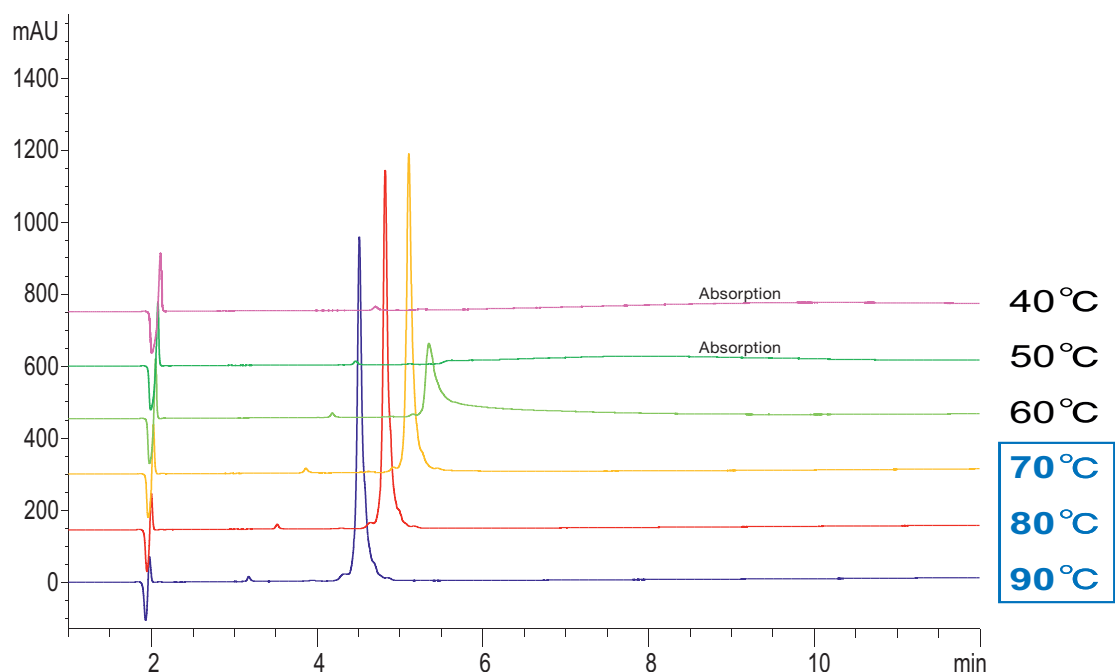
RP – YMC-Triart BioC4: Antibody analysis

High temperature tolerance allows antibody analysis

Adalimumab (Humira®, MW: ca. 148 kDa)



Bevacizumab (Avastin®, MW: ca. 148 kDa)

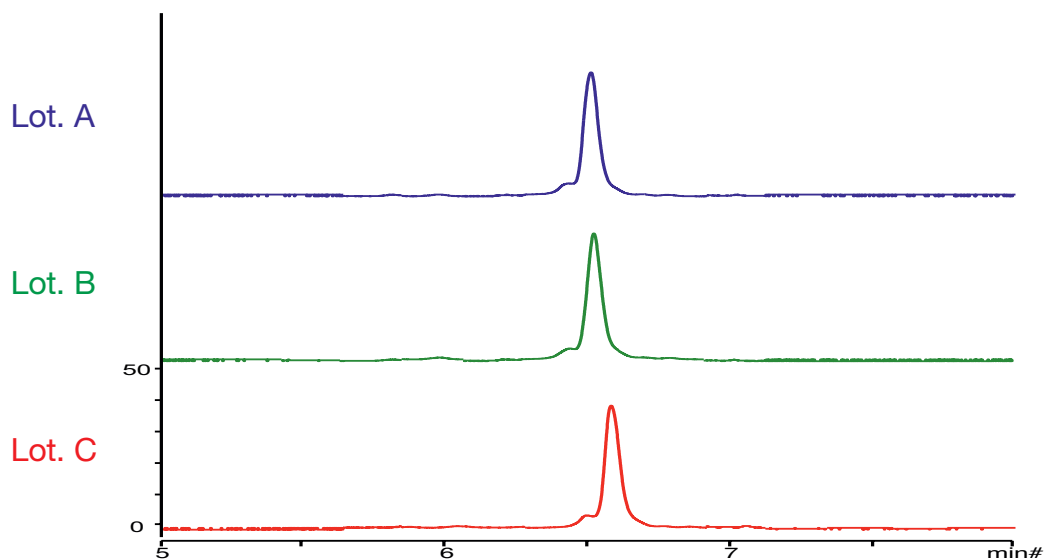


Column: YMC-Triart Bio C4 (3 μ m, 30 nm) 150 x 3.0 mm ID
 Part No: TB30S03-1503PTH
 Eluent: A) water/TFA (100/0.1).
 B) acetonitrile/TFA (100/0.1)
 Gradient: 30-60%B (0-15 min), 90%B (15-30min)
 Detection: UV at 220 nm
 Flow rate: 0.4 mL/min
 Sample: Adalimumab (0.5 mg/mL) or Bevacizumab (0.5 mg/mL)
 Injection: 4 μ L

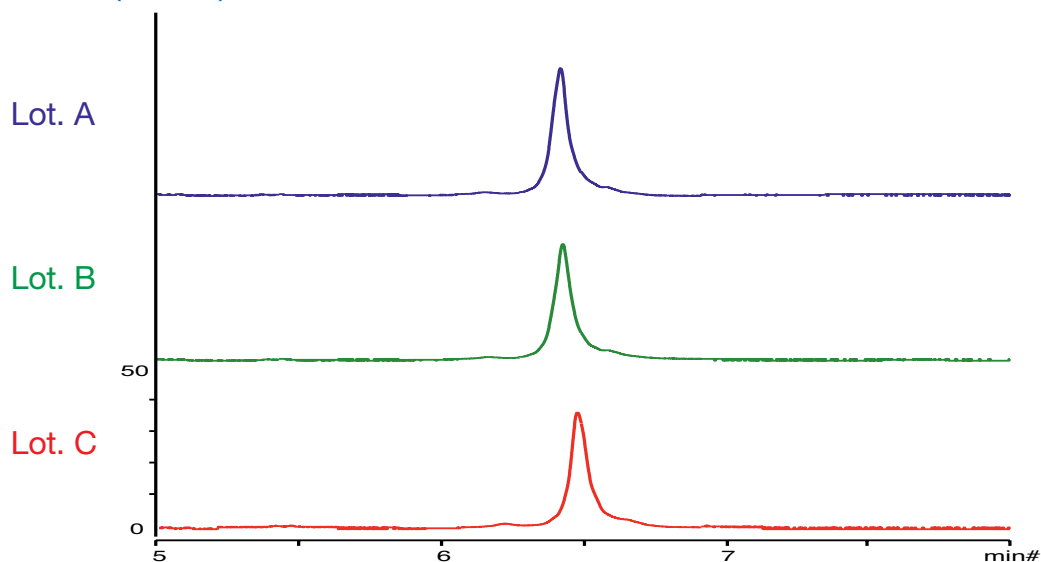
RP – YMC-Triart Bio C4: Reproducibility

Excellent Batch-to-batch reproducibility for antibody analysis

NIST mAb, 8671



Bevacizumab (Avastin®)



Column: YMC-Triart Bio C4 (1.9 μ m, 30 nm) 50 x 2.1 mm ID
Part No.: TB30SP9-05Q1PT
Eluent: A) water/TFA (100/0.1), B) acetonitrile/TFA (100/0.1)
Gradient: 25-45%B (0-10 min)
Detection: UV at 280 nm
Flow rate: 0.4 mL/min
Temperature: 80 °C
Injection: 2 μ L (0.5 mg/mL)

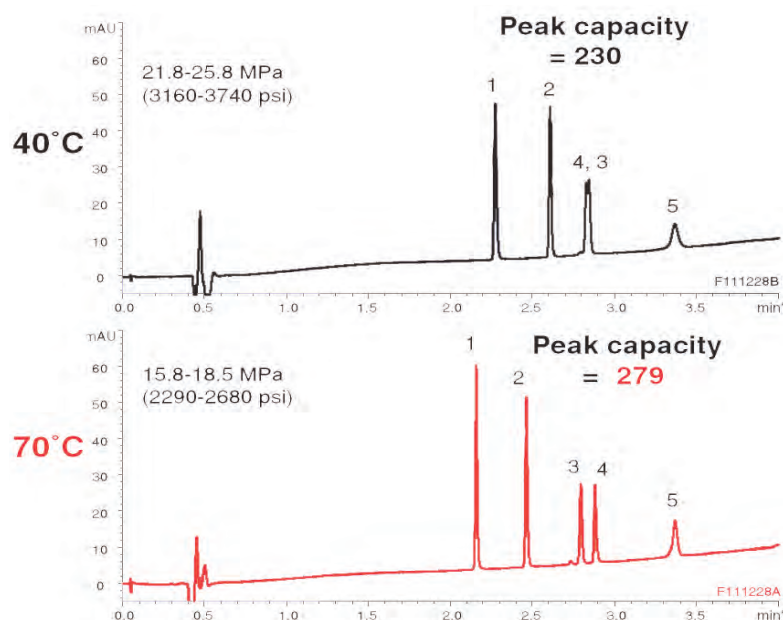
YMC-Triart Bio C4 shows excellent lot-to-lot reproducibility for antibodies. Not only is retention time highly reproducible, but also the resolution of minor impurity peaks. This makes YMC-Triart Bio C4 ideal for quality control of biopharmaceuticals.

RP – YMC-Triart C18: Temperature stability

More temperature flexibility using YMC-Triart

Highly efficient RP-HPLC separation of proteins

Mixture A (MW 500–18,400)



Analytes	MW	Peak width ½h (min)	
		40 °C	70 °C

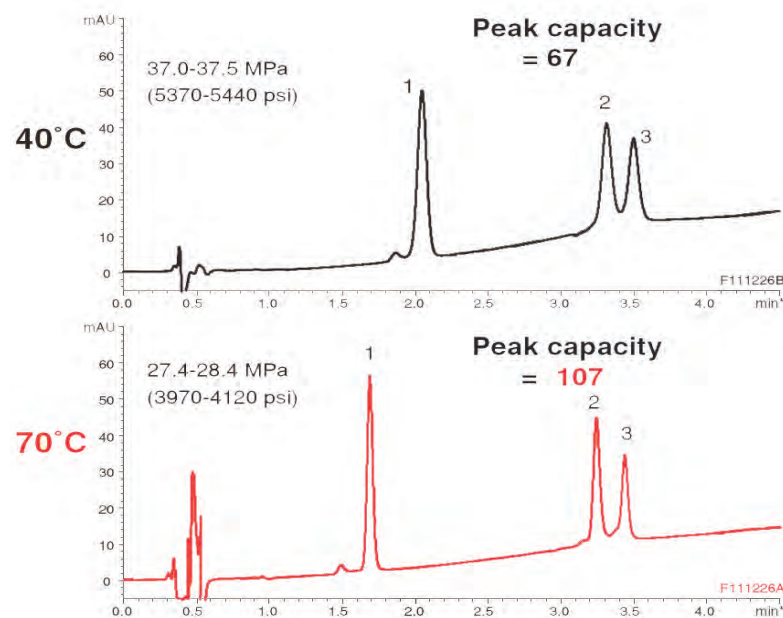
Mixture A

1. Oxytocin	1,007	0.017	0.014
2. Leu-Enkephalin	556	0.015	0.015
3. β-Endorphin	3,465	—	0.016
4. Insulin	5,733	—	0.015
5. β-Lactoglobulin A	18,400	0.043	0.030

Mixture B

1. Lysozyme	14,300	0.069	0.044
2. α-Chymotrypsinogen	25,700	0.080	0.049
3. β-Lactoglobulin A	18,400	0.080	0.048

Mixture B (MW 14,300–25,700)



High temperatures only possible with YMC-Triart

Column: YMC-Triart C18 (1.9 μm, 12 nm) 50 x 2.0 mm ID
Part-No.: TA12SP9-0502WT
Eluent: A) water / TFA (100/0.1)
B) acetonitrile / TFA (100/0.1) - mixture A
B) acetonitrile / 2-propanol / TFA (50/50/0.1) - mixture B
Gradient: 10–80% B (0–5 min) - mixture A
30–60% B (0–5 min) - mixture B

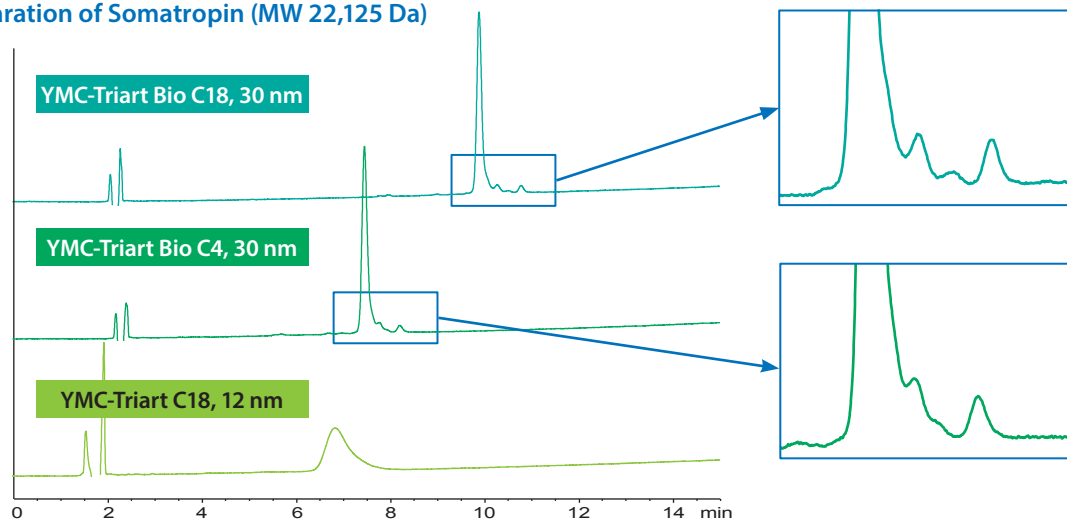
Flow rate: 0.4 mL/min
Detection: UV at 220 nm
Injection: 1 μL (50 μg/mL) - condition A
1 μL (250 μg/mL) - condition B
System: Agilent 1200SL

PC (peak capacity) = 1 + (gradient time / peak width*)
*peak width = 2W_{0.5h} average

RP – YMC-Triart Bio C18: Great peak shapes

Ideal solutions for any kind of biomolecule

Separation of Somatropin (MW 22,125 Da)



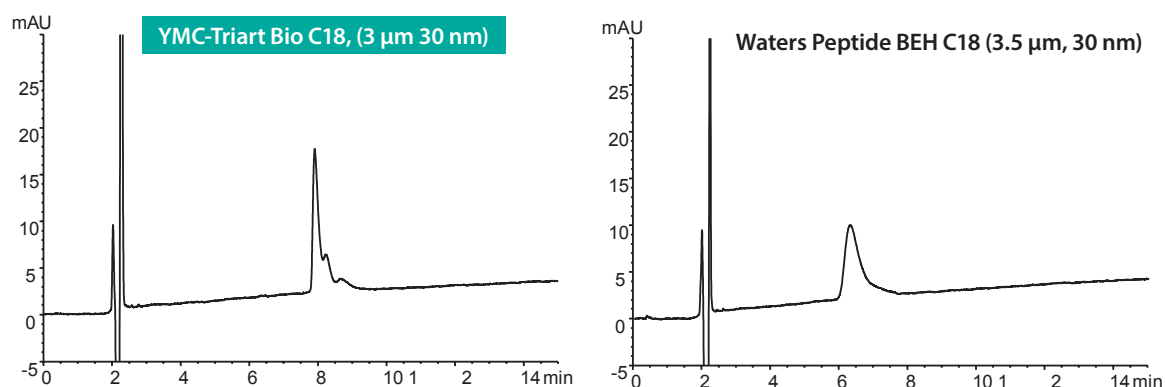
Columns: 150 x 3.0 mm ID (3 μ m)
Part Nos.: TA30S03-1503PTH
TB30S03-1503PTH
TA12S03-1503PTH
Eluent: A) water/TFA (100/0.1)
B) acetonitrile/TFA (100/0.08)

Gradient: 50-70%B (0-15 min)
Flow rate: 0.425 mL/min
Temperature: 40
Detection: UV at 220 nm
Injection: 4 μ L
Sample: Somatropin (0.1 mg/mL)

In this example of somatropin, a peptide of 22,125 Da, good peak shape can be obtained with the widepore columns YMC-Triart Bio C18 and YMC-Triart Bio C4. Excellent separation was achieved using YMC-Triart Bio C18 with longer alkyl chains in its bonded phase.

Ideal for MS conditions

Good peak shape with mobile phase containing formic acid



Column: 150 x 3.0 mm ID; 150 x 4.6 mm ID
Part No.: TA30S03-1503PTH
Eluent: A) water/formic acid (100/0.1)
B) acetonitrile/formic acid (100/0.08)
Gradient: 45-65%B (0-15 min)

Flow rate: 0.425 mL/min for 3.0 mm ID; 1.0 mL/min for 4.6 mm ID
Temperature: 40 °C
Detection: UV at 220 nm
Sample: Somatropin (0.1 mg/mL)

YMC-Triart Bio C18 is suitable for highly sensitive analysis and structural analysis of proteins using LC/MS since good peak shapes in mobile phase containing formic acid can be achieved.

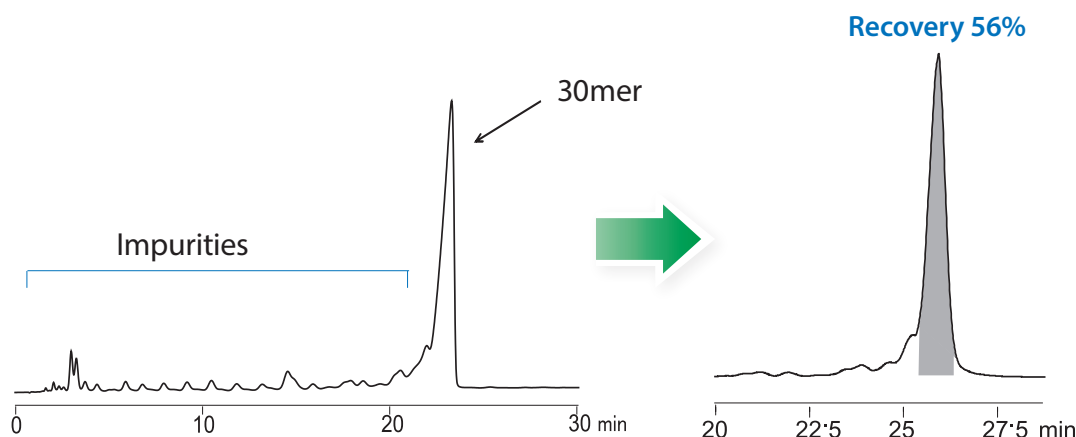
RP – Hydrosphere C18: Oligonucleotide purification

Easy purification of Oligonucleotides with YMC-Actus semi prep columns

Purification of synthetic 30mer oligonucleotide

Analysis 1.0 mL/min, 5 μ L injection
Hydrosphere C18
 50 x 4.6 mm ID, 5 μ m

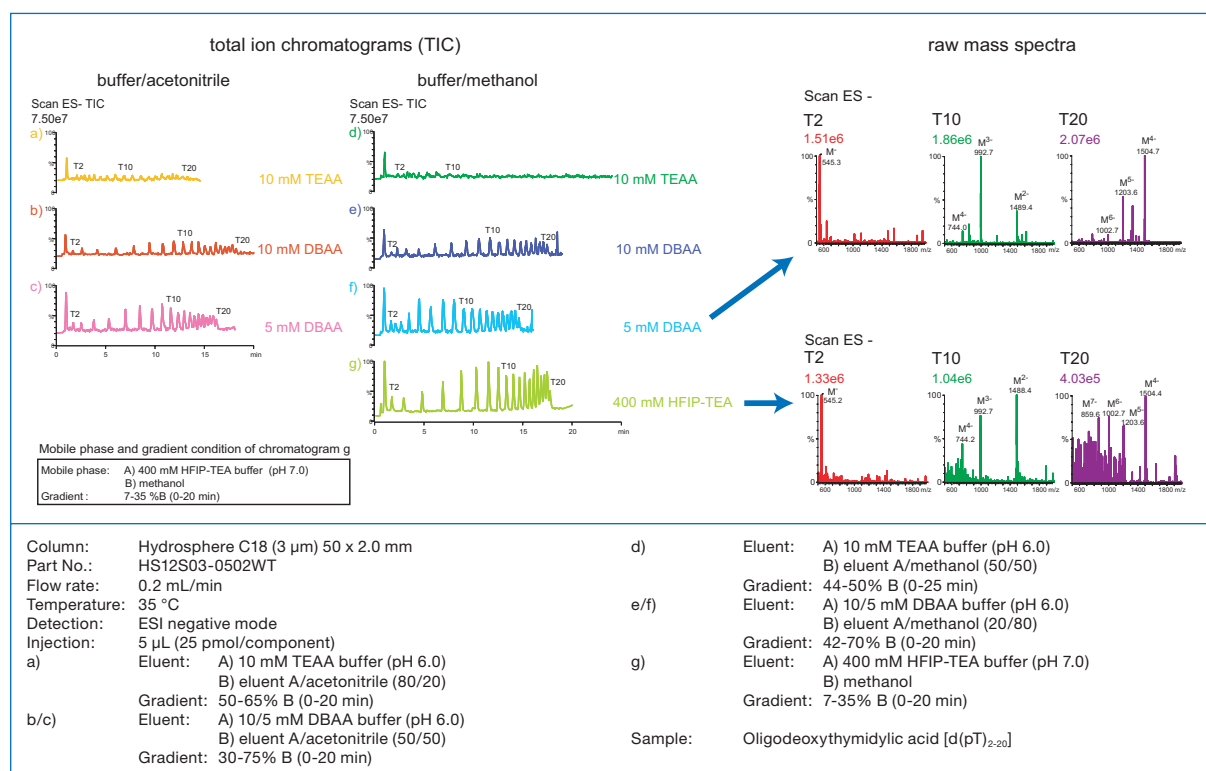
Purification 19 mL/min, 100 μ L injection
YMC-Actus Hydrosphere C18
 50 x 20 mm ID, 5 μ m



Part Nos.: HS12S05-0546WT
 HS12S05-0520WX
 Eluent: A) 10 mM DBA-acetic acid (pH 6.0) / methanol (60/40)
 B) 10 mM DBA-acetic acid (pH 6.0) / methanol (20/80)
 Gradient: 10% -35% B (0-30 min.)
 Temperature: ambient
 Detection: UV at 269 nm
 Sample: synthetic oligonucleotide (100 μ M)

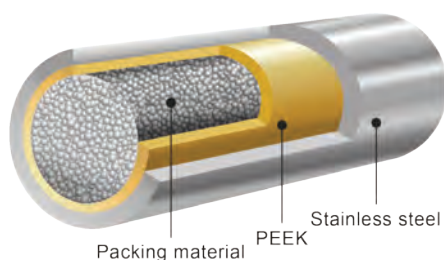
purity >99%

Influences of mobile phase conditions on intensity of ESI-MS



RP – YMC-Triart: Biocompatible hardware

Metal-free column hardware ideal for oligonucleotide analysis



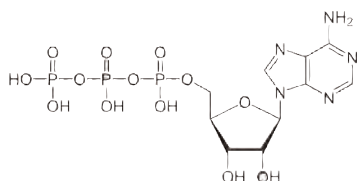
Specifications

YMC-Triart Phases	C18, Bio C18, C8, Bio C4
Particle Size	1.9, 3, 5 μm
Inner layer	PEEK
Outer layer	Stainless steel
Frit	PEEK
Pressure limit	1.9 μm : 100 MPa (15,000 psi) 3/5 μm : 45 MPa (6,525 psi)

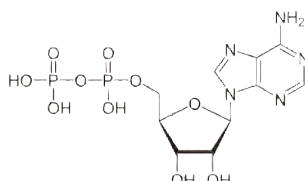
Special column connectors required.
See ordering information recommendations.

Improved sensitivity for coordination compounds

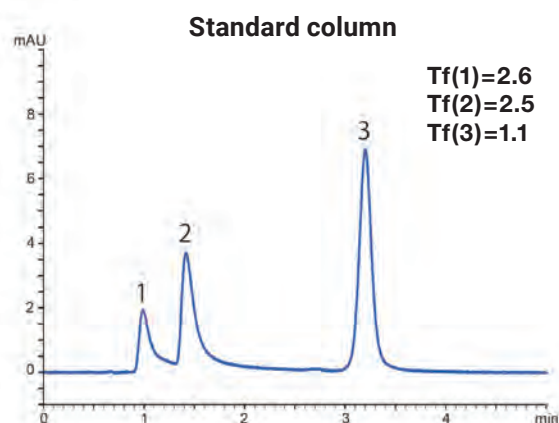
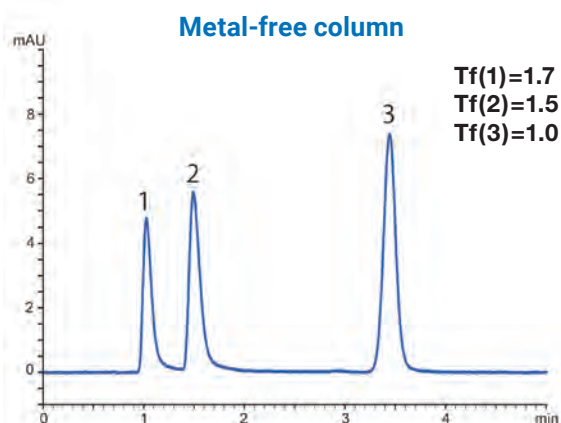
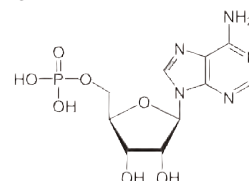
1. ATP



2. ADP



3. AMP



Column: YMC-Triart C18 (3 μm) 50 x 2.1 mm ID
Part Nos.: TA12S03-05Q1PTP (metal-free) or
TA12S03-05Q1PTH (regular hardware)
Eluent: 5 mM HCOONH_4
Flow rate: 0.21 mL/min
Temperature: 25 $^{\circ}\text{C}$
Detection: UV at 265 nm
Injection: 1 mL (10 mg/mL)
System: bioinert/"non-metal" HPLC system

Metal coordinating compounds, which have a phosphate group in their structure, tend to show poor peak shape due to interactions with metals, such as the stainless steel in column bodies and frits. By using the metal-free column hardware, better peak shapes can be expected. Nucleotides with phosphate groups show better peak shapes when compared to the regular column hardware. The metal-free column hardware is very suitable for highly sensitive analyses using LC/MS.

RP – Expert Tips: Proteins/Peptides

Optimisation of peptide and protein separation by reversed-phase

Factors to be considered for method optimisation of peptides and proteins

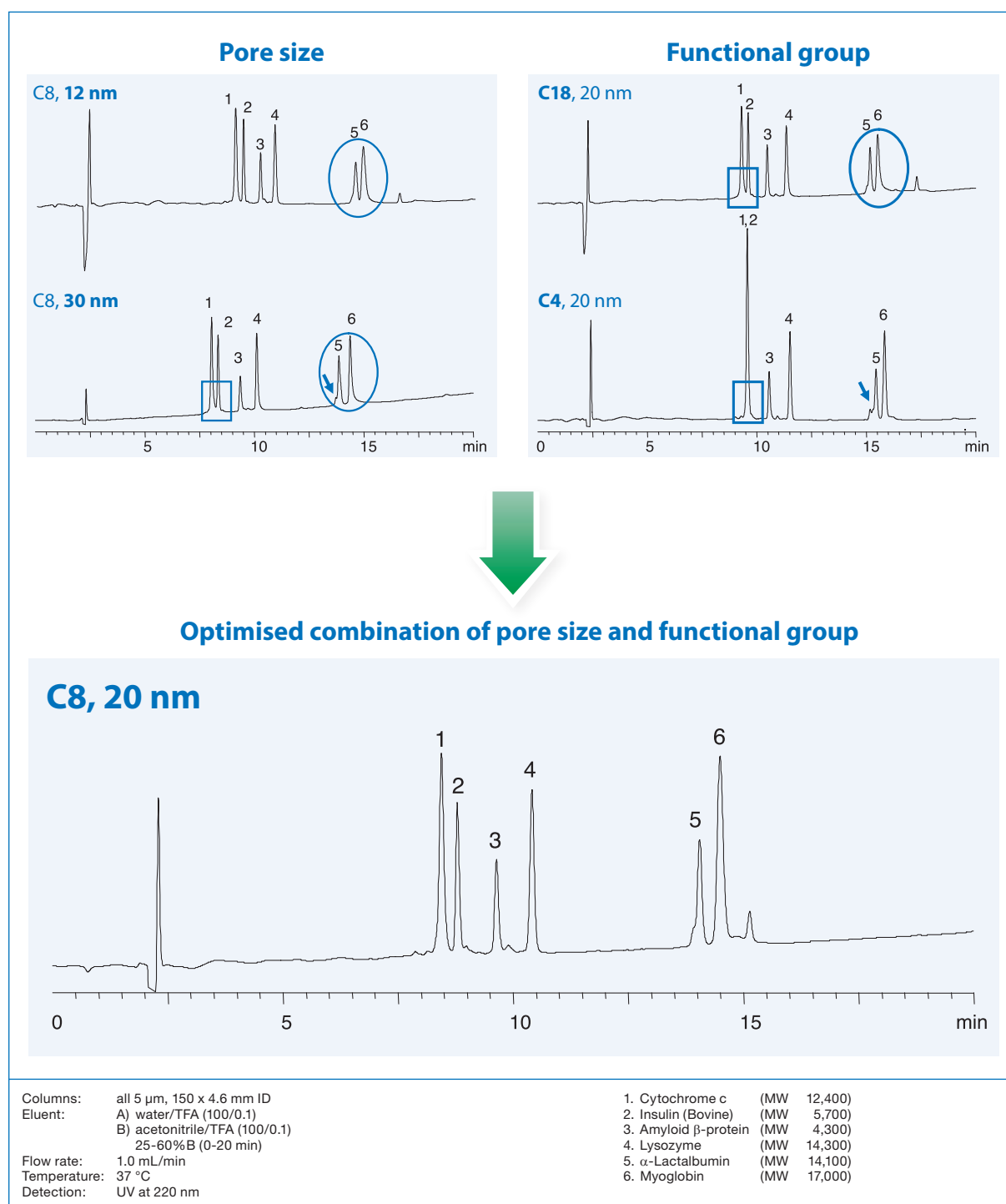
Column	Combination of functional group and pore diameter ► Choose an optimal combination for the molecular weight and hydrophobicity of peptides and proteins. Generally, a column with large pore size and low surface hydrophobicity is suitable for large molecules
Mobile phase	Gradient elution with 0.1% TFA/acetonitrile system as first choice ► Change 1) concentration of TFA, 2) acid species and/or pH if the sample is a mixture of compounds with various ionic characteristics. ► Adjust the gradient conditions, 2-propanol might be effective for separation of large proteins
Temperature	Effective for changing separation selectivity or improving peak shape. However, usable temperature range is limited by column stability. (strongly acidic conditions and heating will accelerate the loss of functional groups resulting in shorter retention times and/or increase in unfavourable secondary interactions between the packing material and sample)

Comparison of separation on columns with different pore sizes and functional groups

The separation characteristics of proteins and peptides with molecular weights of 4,300–17,000 are compared using columns with different pore sizes and functional groups. The figure below shows that the most suitable column for compounds with a molecular weight within this range is C8, 20 nm.

If either the pore size or functional group of the packing material is not optimised, peak broadening and poor resolution are observed.

By using the most suitable column (C8, 20 nm) for the target compounds, sharp peak shapes and excellent separation are achieved.



RP – Expert Tips: Proteins/Peptides

Effect of column temperature and mobile phase composition (Optimisation of antimicrobial peptides separation)

Structure (antimicrobial peptides)

- | | | |
|-------------------------|----------|---------------------------------------|
| 1. α-Defensin-1: | A | CYCRIPACIAGERRYGTCTIYQGRLWAFCC |
| MW 3,442 | | |
| 2. α-Defensin-2: | | CYCRIPACIAGERRYGTCTIYQGRLWAFCC |
| MW 3,371 | | |
| 3. α-Defensin-3: | D | CYCRIPACIAGERRYGTCTIYQGRLWAFCC |
| MW 3,486 | | |

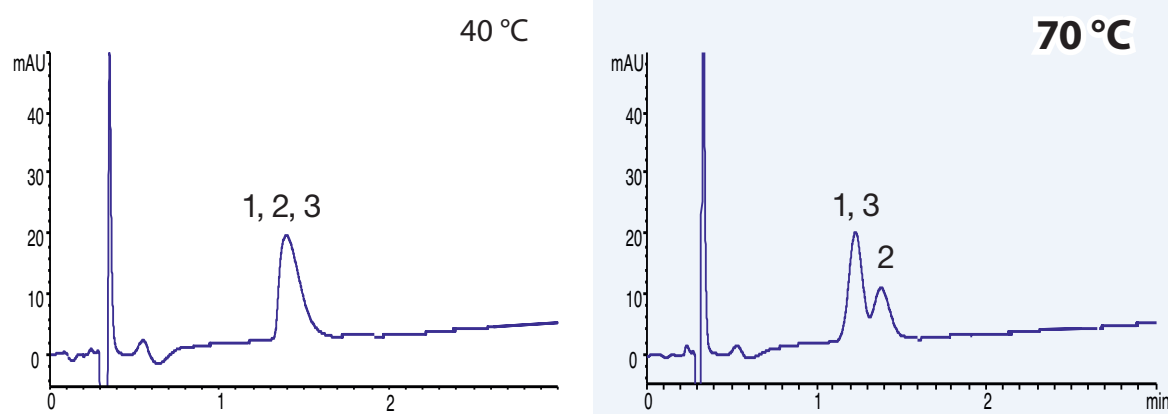
* Difference in amino acid residue on N-terminal

Common HPLC conditions

Column: **YMC-Triart C18** (1.9 µm, 12 nm) 50 x 2.0 mm ID
 Part No.: TA12SP9-0502PT
 Flow rate: 0.4 mL/min
 Detection: 220 nm

1 Effect of column temperature

Changing the column temperature will provide selectivity changes as well as peak shape improvement. The high stability column, YMC-Triart, can offer a wider usable temperature range. Temperature can be used as a tool for method optimisation.

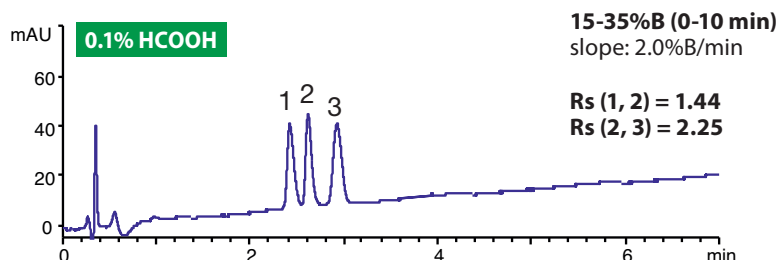
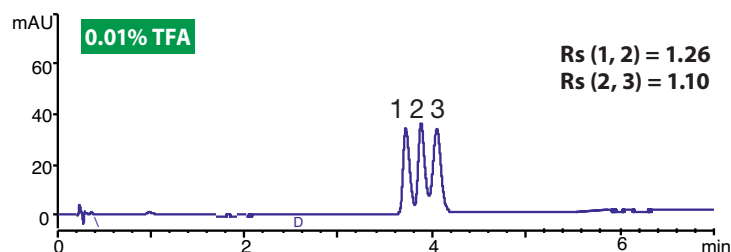
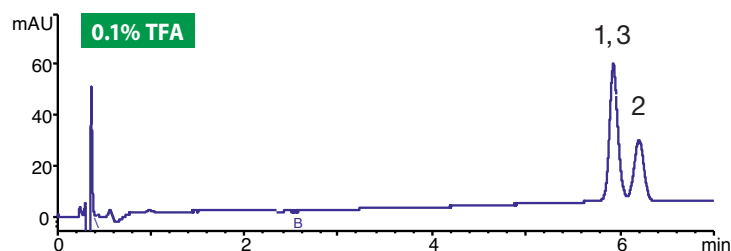


Eluent: A) water/TFA (100/0.1)
 B) acetonitrile/TFA (100/0.1)
 Gradient: 25–45%B (0-5 min)

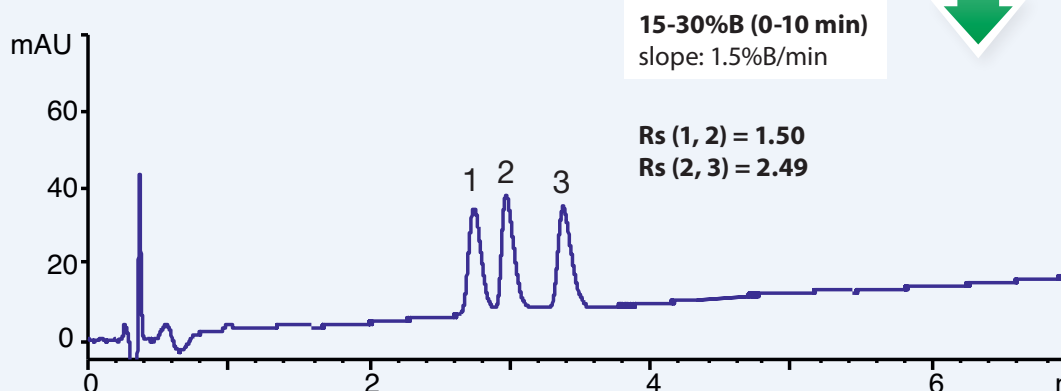
2 Effect of acid type, acid concentration and gradient conditions (at 70°C)

Selectivity changes can be expected when changing acid type and/or concentration. It is useful when there is a large difference in ionic characteristics of compounds.

Acid type/Acid concentration



Gradient slope



Eluent: A) acid-containing aqueous solution
B) acid-containing acetonitrile solution
(0.1% HCOOH in B solution is 0.08%)

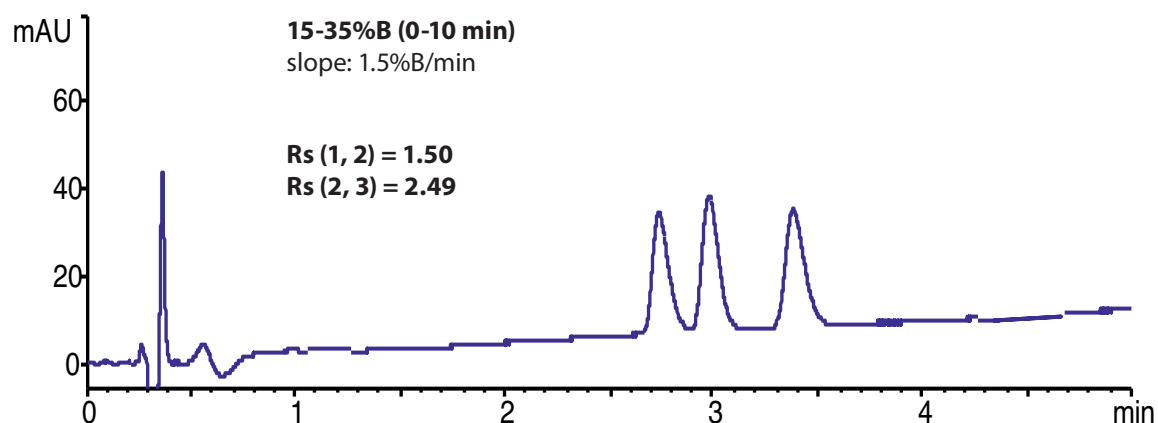
Temperature: 70 °C

RP – Expert Tips: Proteins/Peptides

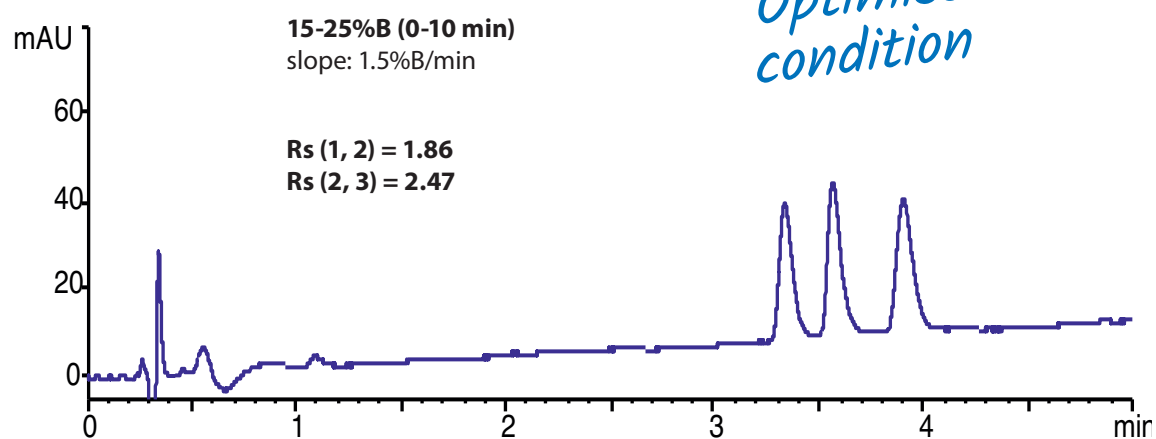
3 Addition of 2-propanol in a mobile phase

When 2-propanol is added in the mobile phase and the gradient slope is optimised resolution between peaks 1 and 2 is improved while the analysis time remains the same.

acetonitrile



2-propanol/acetonitrile (50/50)



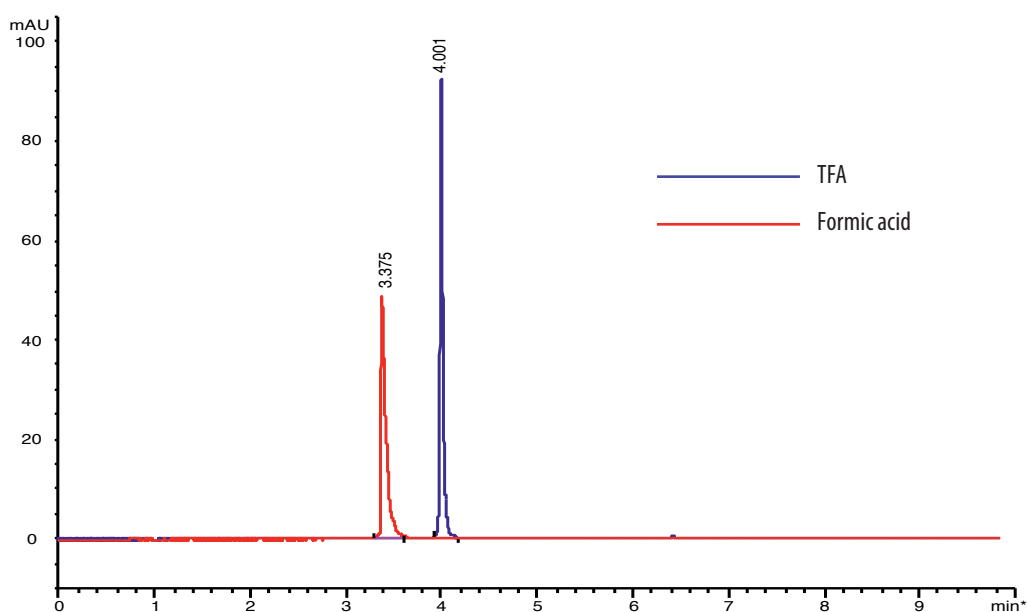
Eluent: A) 0.1% formic acid in water
B) 0.08% formic acid in organic solvent

Temperature: 70 °C

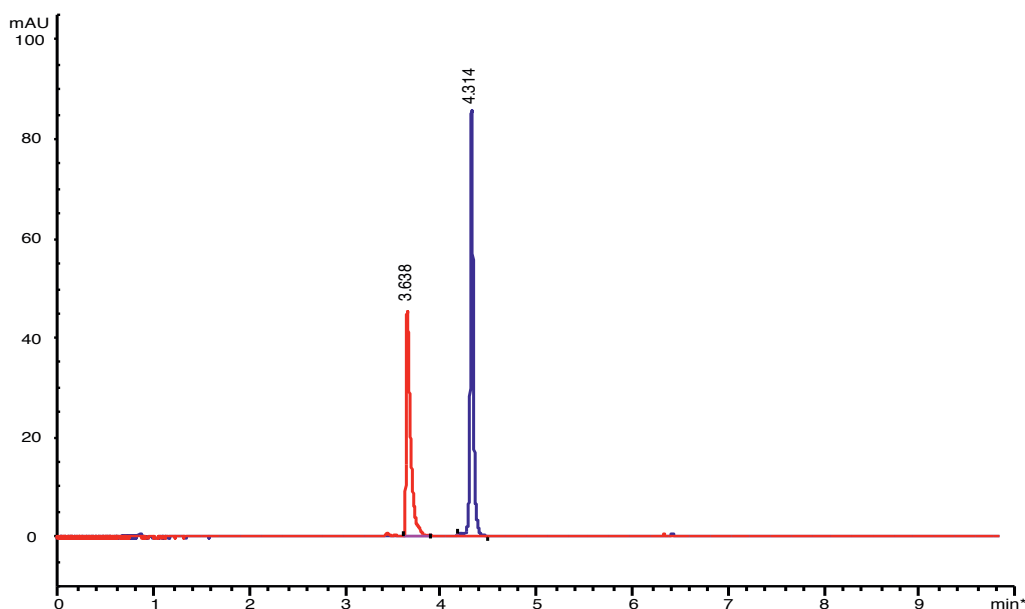
Use of MS compatible conditions for antibody analysis by RP

Although the best peak shapes and greater sensitivity can be provided by mobile phases containing TFA, suitable peaks can also be obtained with mobile phases containing formic acid – especially in combination with YMC-Triart Bio C4.

Adalimumab (Humira®)



NIST Mab

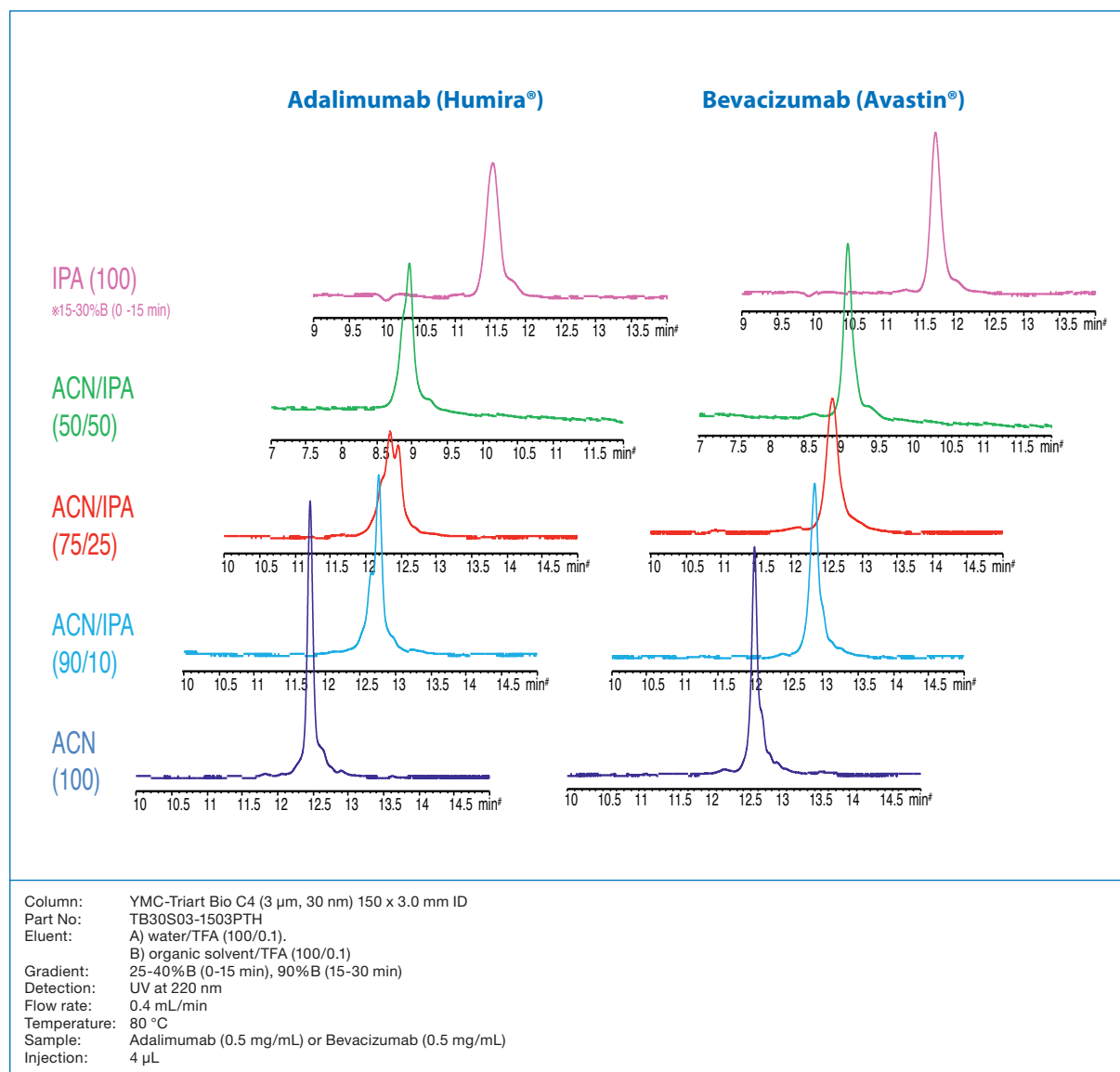


Column: YMC-Triart Bio C4 (1.9 μ m, 30 nm) 150 x 2.1 mm ID
 Part No: TB30SP9-15Q1PT
 Eluent: A) water/TFA or Formic acid (100/0.1)
 B) acetonitrile/TFA or Formic acid (100/0.1)
 Gradient: 10-95%B (0-10 min)
 Detection: UV at 280 nm (0.13s, 40Hz)
 Flow rate: 0.4 mL/min
 Temperature: 80 °C
 Sample conc.: 0.5 mg/mL
 Injection: 2 μ L

RP – Expert Tips: Antibodies

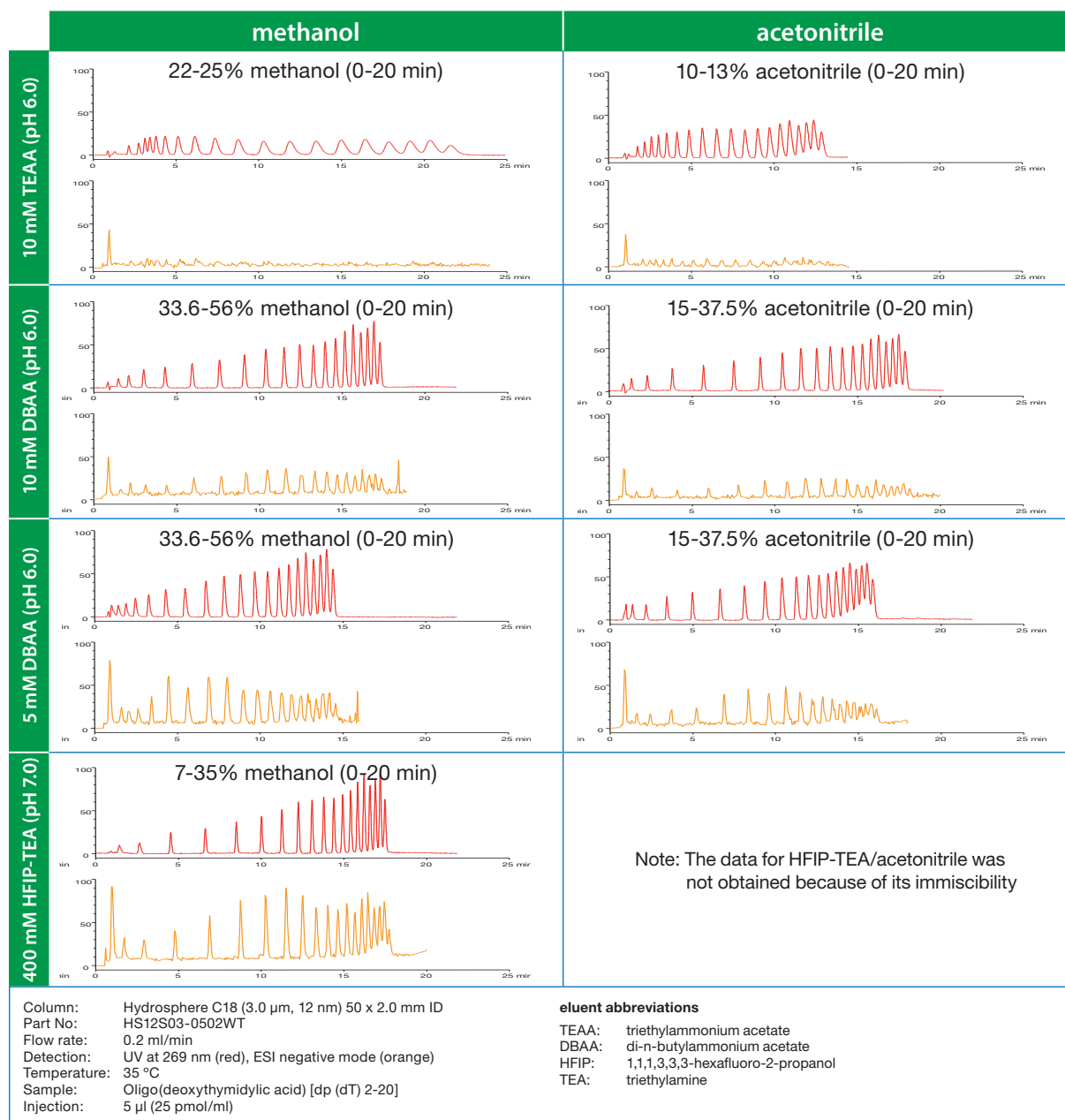
Influence of organic solvents on antibody analysis

The separation behaviour can be influenced by a large amount by an organic solvent in the mobile phase. Using the optimum organic solvent is beneficial for MAb characterisation.



Effect of composition and salt concentration of ion-pairing mobile phase on the separation and signal intensity

Comparison of separation and ESI-MS signal intensity using different ion-pairing buffers and organic solvents



The mobile phase composition has different effects on the separation and signal intensity in electrospray ionisation mass spectrometry (ESI-MS) of oligonucleotides. Using different gradient conditions, acceptable retention and resolution can be achieved (upper UV chromatograms; red trace) for each separation by optimising the gradient slope of the organic solvent regardless of the type of mobile phase. The ESI-MS intensity is significantly influenced

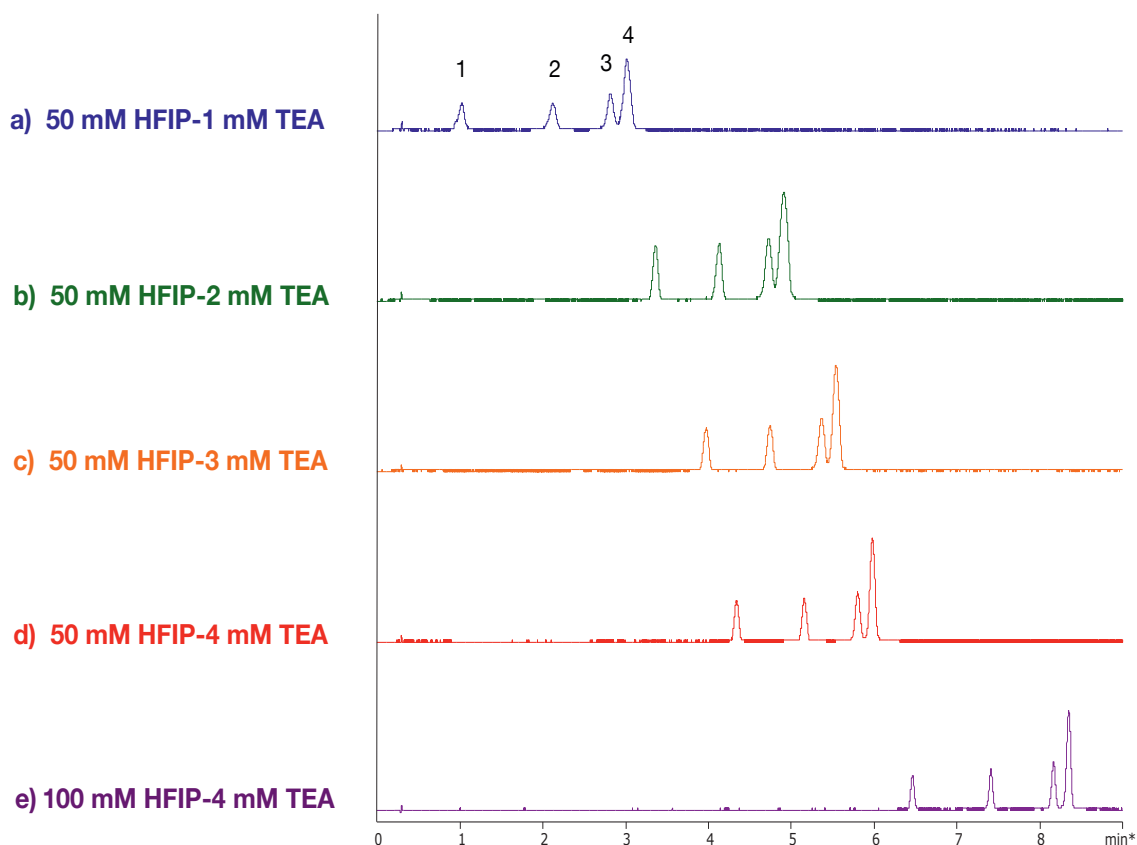
by the type and concentration of ion-pairing buffer as shown in the lower MS chromatograms (orange trace). HFIP-TEA buffer/methanol systems provide the maximum MS intensity. Enhanced retention and MS intensity are obtained using 10 mM DBAA buffer compared to 10 mM TEAA buffer, and the lower DBAA concentration results in approximately 1.5–3 times increase in the intensity without any change in the concentration of organic solvent.

RP – Expert Tips: Oligonucleotides

Salt concentration in HFIP-TEA buffer

RNA Sample

rCrArCrUrGrArArUrArCrCrArArU (14mer)
 rUrCrArCrArCrUrGrArArUrArCrCrArArU (17mer)
 rUrCrArUrCrArCrArCrUrGrArArUrArCrCrArArU (20mer)
 rGrUrCrArUrCrArCrArCrUrGrArArUrArCrCrArArU (21mer)



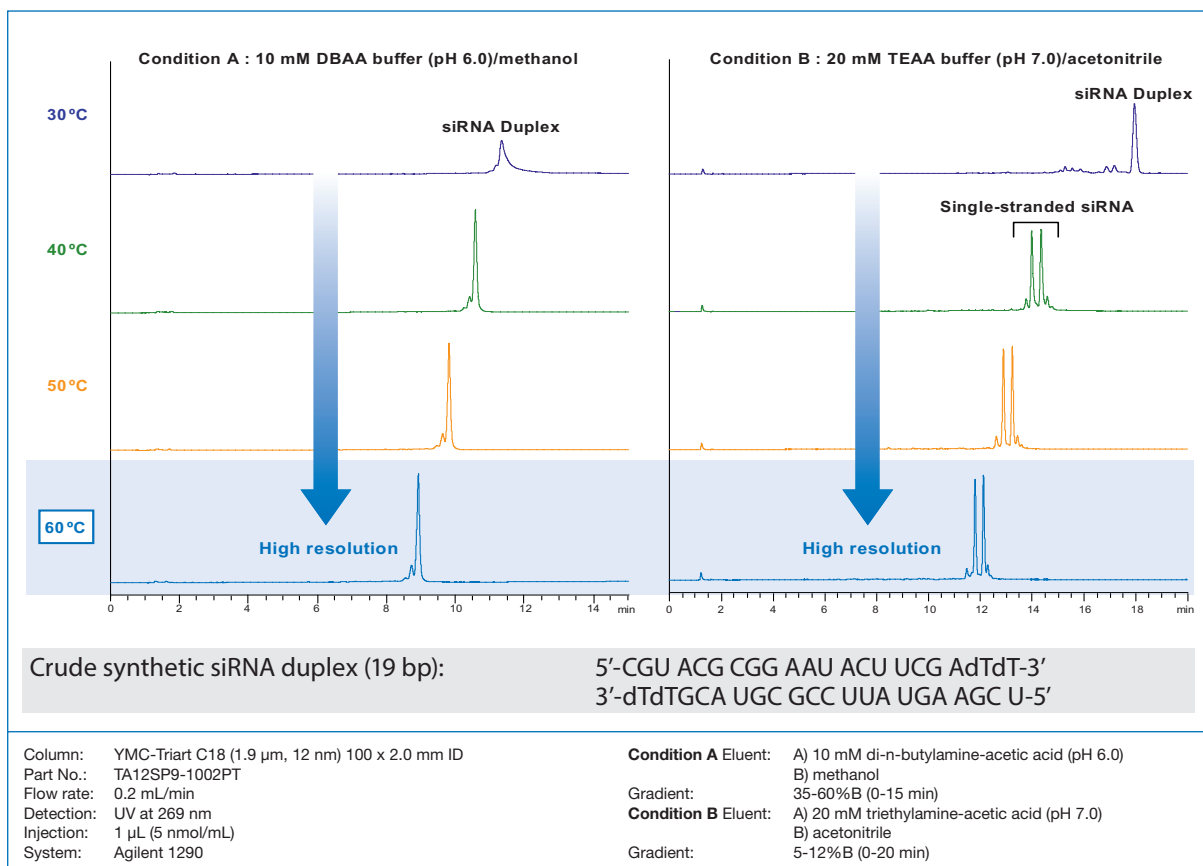
Column: YMC-Triart C18 (1.9 μ m, 12 nm) 50 x 2.1 mm ID
 Part No: TA12SP9-05Q1PT
 Eluent: A) shown in figure
 B) methanol
 Gradient: 5-15%B (0-10 min)
 Flow rate: 0.42 ml/min
 Detection: UV at 260 nm
 Temperature: 65 $^{\circ}$ C
 Injection: 1.0 μ l (2-4 nmol/ml)

Oligonucleotide separations are influenced by the use of mobile phases consisting of different ratios of HFIP and TEA. The retention time is approximately doubled when the TEA concentration is increased from 1 to 4 mM at a constant 50 mM HFIP concentration (chromatograms a – d), and increased approximately 1.5 times when

the HFIP concentration is increased from 50 mM to 100 mM at a constant 4 mM TEA concentration (chromatograms d and e).

These results indicate that small changes in concentration of basic ion-pairing reagents such as TEA and DBA provide a great impact on oligonucleotide separations.

Effect of mobile phase and column temperature on the separation of siRNA duplex



Using different mobile phase conditions, peak shape and resolution between intermediate peaks is improved by increasing the column temperature. As a result of enhanced dispersion and distribution velocity when increasing column temperature, biomolecules such as RNA and DNA generally exhibit sharper peak shapes and improved resolution.

Under condition B at 40°C or above, two peaks of single-stranded RNA, generated by the denaturation of the siRNA duplex, are observed. This HPLC technique using high

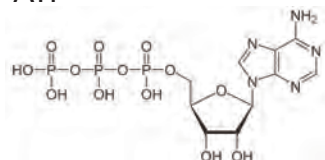
temperature to generate single-stranded RNA is called “Denaturing HPLC”, and is widely used in the field of gene mutation analysis.

Denaturation of duplex DNA or RNA is also influenced by ionic strength (type and concentration), pH and polarity. It is recommended that these analysis conditions (temperature and mobile phase) should be optimised depending on characteristics of the target analyte and the purpose of the analysis.

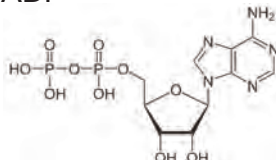
RP – Expert Tips: (Oligo)nucleotides

Influence of system and column hardware on the analysis of nucleotides

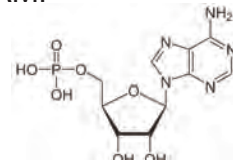
1 ATP



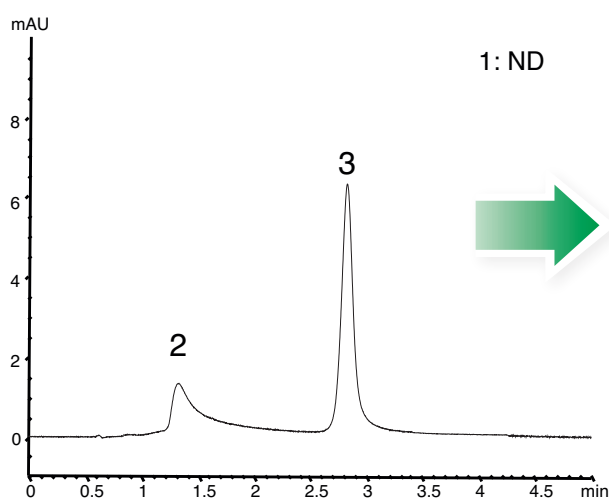
2 ADP



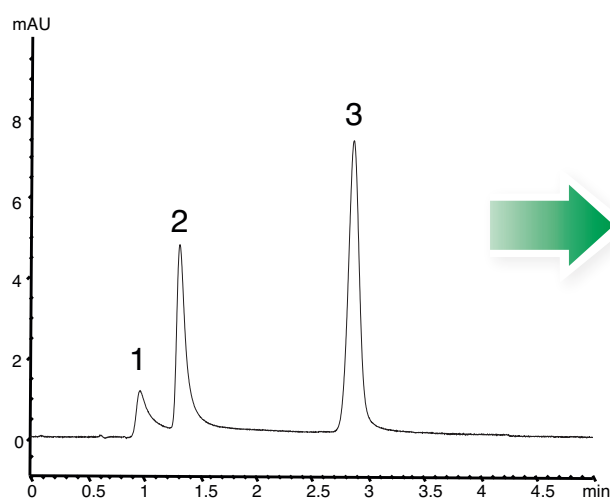
3 AMP



Ordinary HPLC system
with standard column

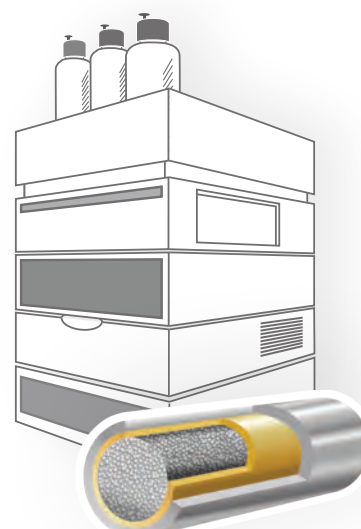
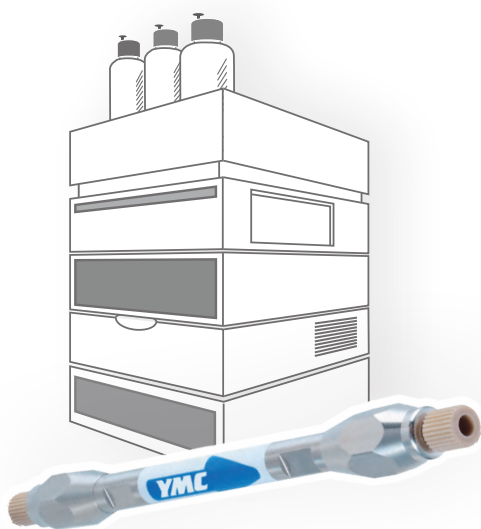


Ordinary HPLC system
with metal-free column



Column: YMC-Triart C18 (3 μ m, 12 nm) 50 x 2.1 mm ID
 Part Nos: TA12S03-05Q1PT (standard hardware)
 TA12S03-05Q1PTP (metal-free hardware)
 Eluent: 5 mM HCOONH₄
 Flow rate: 0.21 mL/min
 Temperature: 25 °C
 Detection: UV at 265 nm
 Injection: 1 μ L (10 μ g/mL)

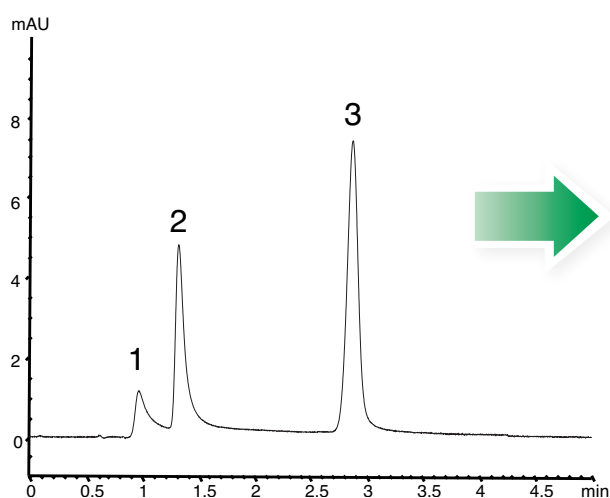
"Non-metal" HPLC system: PEEK sample loop, PEEK injector port, and PEEK tubing are used.



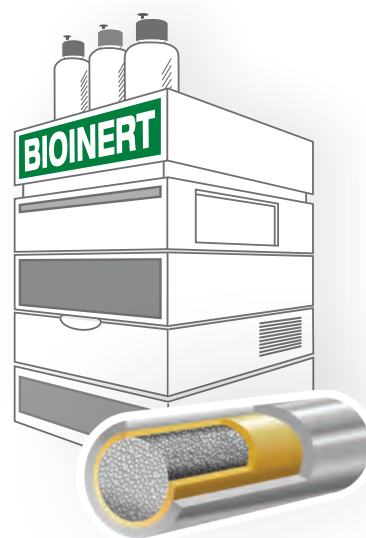
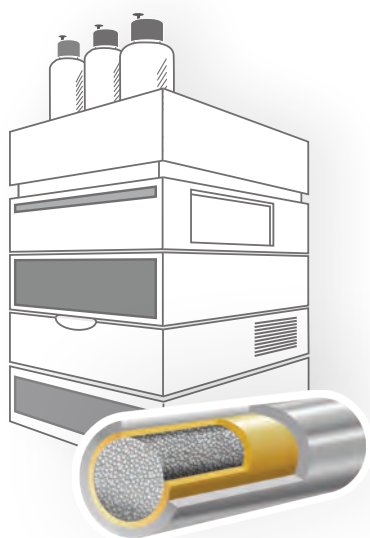
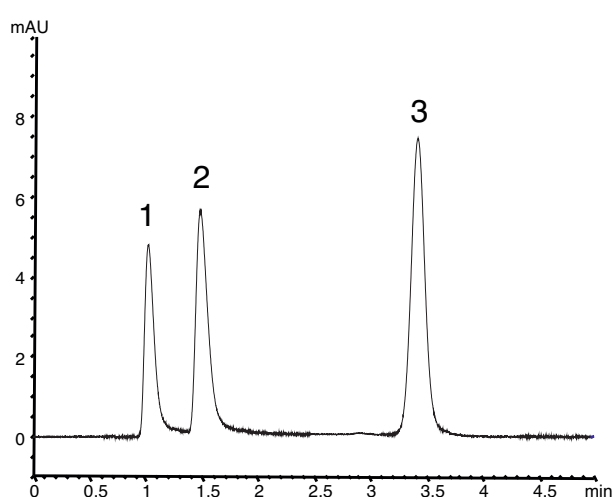
ATP peak is detected, and peak shape of ADP is improved as a result of using the metal-free column.

RP – Expert Tips: (Oligo)nucleotides

Ordinary HPLC system
with metal-free column



“Non-metal” HPLC system*
with metal-free column



Peak shape is greatly improved as a result of using “non-metal” HPLC system

RP – Ordering information

1.9 µm UHPLC columns

Phase	Column ID [mm]	Column length [mm]					Guard cartridges* with 5 mm length
		30	50	75	100	150	(pack of 3)
YMC-Triart C18	2.0	TA12SP9-0302PT	TA12SP9-0502PT	TA12SP9-L502PT	TA12SP9-1002PT	TA12SP9-1502PT	TA12SP9-E5Q1CC**
	2.1	TA12SP9-03Q1PT	TA12SP9-05Q1PT	TA12SP9-L5Q1PT	TA12SP9-10Q1PT	TA12SP9-15Q1PT	TA12SP9-E5Q1CC**
	3.0	—	TA12SP9-0503PT	TA12SP9-L503PT	TA12SP9-1003PT	TA12SP9-1503PT	TA12SP9-E503CC
YMC-Triart Bio C18	2.0	TA30SP9-0302PT	TA30SP9-0502PT	TA30SP9-L502PT	TA30SP9-1002PT	TA30SP9-1502PT	TA30SP9-E5Q1CC**
	2.1	TA30SP9-03Q1PT	TA30SP9-05Q1PT	TA30SP9-L5Q1PT	TA30SP9-10Q1PT	TA30SP9-15Q1PT	TA30SP9-E5Q1CC**
	3.0	—	TA30SP9-0503PT	TA30SP9-L503PT	TA30SP9-1003PT	TA30SP9-1503PT	TA30SP9-E503CC
YMC-Triart C8	2.0	T012SP9-0302PT	T012SP9-0502PT	T012SP9-L502PT	T012SP9-1002PT	T012SP9-1502PT	T012SP9-E5Q1CC**
	2.1	T012SP9-03Q1PT	T012SP9-05Q1PT	T012SP9-L5Q1PT	T012SP9-10Q1PT	T012SP9-15Q1PT	T012SP9-E5Q1CC**
	3.0	—	T012SP9-0503PT	T012SP9-L503PT	T012SP9-1003PT	T012SP9-1503PT	T012SP9-E503CC
YMC-Triart Bio C4	2.0	TB30SP9-0302PT	TB30SP9-0502PT	TB30SP9-L502PT	TB30SP9-1002PT	TB30SP9-1502PT	TB30SP9-E5Q1CC**
	2.1	TB30SP9-03Q1PT	TB30SP9-05Q1PT	TB30SP9-L5Q1PT	TB30SP9-10Q1PT	TB30SP9-15Q1PT	TB30SP9-E5Q1CC**
	3.0	—	TB30SP9-0503PT	TB30SP9-L503PT	TB30SP9-1003PT	TB30SP9-1503PT	TB30SP9-E503CC

*Guard cartridge holder required, part no. XPCUHP

**Guard cartridge: 2.1 mm ID

1.9 µm metal-free UHPLC columns

Phase	Column ID [mm]	Column length [mm]		
		50	100	150
YMC-Triart C18	2.1	TA12SP9-05Q1PTP	TA12SP9-10Q1PTP	TA12SP9-15Q1PTP
YMC-Triart Bio C18	2.1	TA30SP9-05Q1PTP	TA30SP9-10Q1PTP	TA30SP9-15Q1PTP
YMC-Triart C8	2.1	T012SP9-05Q1PTP	T012SP9-10Q1PTP	T012SP9-15Q1PTP
YMC-Triart Bio C4	2.1	TB30SP9-05Q1PTP	TB30SP9-10Q1PTP	TB30SP9-15Q1PTP

Special column connectors required.

Column connectors

Recommendation	✓ ✓		✓	
Ferrule	no		replaceable	
Product	MarvelX™	MarvelXACT™	Handy connector 2	Hand-tight EXP® fitting
Manufacturer	IDEX Health & Science LLC	IDEX Health & Science LLC	YMC Co., Ltd.	Optimize Technologies, Inc.
				
Pressure rating	131 MPa / 1,310 bar	131 MPa / 1,310 bar	42 MPa / 420 bar	137 MPa / 1,370 bar
Product code	e.g. UPFP-6050250	e.g. UPFP-YM7050250	XRP0204	XRHTF-01

MarvelX (ACT) is a registered trademark of IDEX Health & Science LLC · EXP® is a registered trademark of Optimize Technologies, Inc.

RP – Ordering information

3 µm HPLC columns

Phase	Column ID [mm]	Column length [mm]						Guard cartridges* with 10 mm length
		30/33	50	75	100	150	250	(pack of 5)
YMC-Triart C18	2.1	TA12S03-H3Q1PTH	TA12S03-05Q1PTH	TA12S03-L5Q1PTH	TA12S03-10Q1PTH	TA12S03-15Q1PTH	–	TA12S03-01Q1GC
	3.0	–	TA12S03-05Q3PTH	TA12S03-L5Q3PTH	TA12S03-10Q3PTH	TA12S03-15Q3PTH	–	TA12S03-01Q3GC
	4.6	TA12S03-H346PTH	TA12S03-0546PTH	TA12S03-L546PTH	TA12S03-1046PTH	TA12S03-1546PTH	TA12S03-2546PTH	TA12S03-01Q4GC
YMC-Triart Bio C18	2.1	TA30S03-H3Q1PTH	TA30S03-05Q1PTH	TA30S03-L5Q1PTH	TA30S03-10Q1PTH	TA30S03-15Q1PTH	–	TA30S03-01Q1GC
	3.0	–	TA30S03-05Q3PTH	TA30S03-L5Q3PTH	TA30S03-10Q3PTH	TA30S03-15Q3PTH	–	TA30S03-01Q3GC
	4.6	TA30S03-H346PTH	TA30S03-0546PTH	TA30S03-L546PTH	TA30S03-1046PTH	TA30S03-1546PTH	TA30S03-2546PTH	TA30S03-01Q4GC
YMC-Triart C8	2.1	T012S03-H3Q1PTH	T012S03-05Q1PTH	T012S03-L5Q1PTH	T012S03-10Q1PTH	T012S03-15Q1PTH	–	T012S03-01Q1GC
	3.0	–	T012S03-05Q3PTH	T012S03-L5Q3PTH	T012S03-10Q3PTH	T012S03-15Q3PTH	–	T012S03-01Q3GC
	4.6	T012S03-H346PTH	T012S03-0546PTH	T012S03-L546PTH	T012S03-1046PTH	T012S03-1546PTH	T012S03-2546PTH	T012S03-01Q4GC
YMC-Triart Bio C4	2.1	TB30S03-H3Q1PTH	TB30S03-05Q1PTH	TB30S03-L5Q1PTH	TB30S03-10Q1PTH	TB30S03-15Q1PTH	–	TB30S03-01Q1GC
	3.0	–	TB30S03-05Q3PTH	TB30S03-L5Q3PTH	TB30S03-10Q3PTH	TB30S03-15Q3PTH	–	TB30S03-01Q3GC
	4.6	TB30S03-H346PTH	TB30S03-0546PTH	TB30S03-L546PTH	TB30S03-1046PTH	TB30S03-1546PTH	TB30S03-2546PTH	TB30S03-01Q4GC
Hydrosphere C18	2.1	HS12S03-H3Q1QT	HS12S03-05Q1QT	HS12S03-L5Q1QT	HS12S03-10Q1QT	HS12S03-15Q1QT	HS12S03-25Q1QT	HS12S03-01Q1GC
	3.0	HS12S03-H3Q3QT	HS12S03-05Q3QT	HS12S03-L5Q3QT	HS12S03-10Q3QT	HS12S03-15Q3QT	HS12S03-25Q3QT	HS12S03-01Q3GC
	4.6	HS12S03-0346WT	HS12S03-0546WT	HS12S03-L546WT	HS12S03-1046WT	HS12S03-1546WT	HS12S03-2546WT	HS12S03-01Q4GC
YMCbasic (eq. C8)	2.1	BA99S03-H3Q1QT	BA99S03-05Q1QT	BA99S03-L5Q1QT	BA99S03-10Q1QT	BA99S03-15Q1QT	BA99S03-25Q1QT	BA99S03-01Q1GC
	3.0	BA99S03-H3Q3QT	BA99S03-05Q3QT	BA99S03-L5Q3QT	BA99S03-10Q3QT	BA99S03-15Q3QT	BA99S03-25Q3QT	BA99S03-01Q3GC
	4.6	BA99S03-0346WT	BA99S03-0546WT	BA99S03-L546WT	BA99S03-1046WT	BA99S03-1546WT	BA99S03-2546WT	BA99S03-01Q4GC

*Guard cartridge holder required, part no. XPGCH-Q1

3 µm metal-free HPLC columns

Phase	Column ID [mm]	Column length [mm]		
		50	100	150
YMC-Triart C18	2.1	TA12S03-05Q1PTP	TA12S03-10Q1PTP	TA12S03-15Q1PTP
	4.6	TA12S03-0546PTP	TA12S03-1046PTP	TA12S03-1546PTP
YMC-Triart Bio C18	2.1	TA30S03-05Q1PTP	TA30S03-10Q1PTP	TA30S03-15Q1PTP
	4.6	TA30S03-0546PTP	TA30S03-1046PTP	TA30S03-1546PTP
YMC-Triart C8	2.1	T012S03-05Q1PTP	T012S03-10Q1PTP	T012S03-15Q1PTP
	4.6	T012S03-0546PTP	T012S03-1046PTP	T012S03-1546PTP
YMC-Triart Bio C4	2.1	TB30S03-05Q1PTP	TB30S03-10Q1PTP	TB30S03-15Q1PTP
	4.6	TB30S03-0546PTP	TB30S03-1046PTP	TB30S03-1546PTP

Special column connectors required.

RP – Ordering information

2.7 µm Core-Shell columns

Phase	Column ID [mm]	Column length [mm]					Precolumn filter 0.5 µm
		30	50	75	100	150	(pack of 3)
Meteoric Core C18 BIO, 16 nm	2.1	CAW16SQ7-03Q1PT	CAW16SQ7-05Q1PT	CAW16SQ7-L5Q1PT	CAW16SQ7-10Q1PT	CAW16SQ7-15Q1PT	XRPRCS35
	3.0	CAW16SQ7-0303PT	CAW16SQ7-0503PT	CAW16SQ7-L503PT	CAW16SQ7-1003PT	CAW16SQ7-1503PT	
	4.6	CAW16SQ7-0346PT	CAW16SQ7-0546PT	CAW16SQ7-L546PT	CAW16SQ7-1046PT	CAW16SQ7-1546PT	

*Holder required, part no. XRPRCS03

5 µm HPLC columns

Phase	Column ID [mm]	Column length [mm]						Guard cartridges* with 10 mm length
		30/33	50	75	100	150	250	(pack of 5)
YMC-Triart C18	2.1	TA12S05-H3Q1PTH	TA12S05-05Q1PTH	TA12S05-L5Q1PTH	TA12S05-10Q1PTH	TA12S05-15Q1PTH	–	TA12S05-01Q1GC
	3.0	–	TA12S05-0503PTH	TA12S05-L503PTH	TA12S05-1003PTH	TA12S05-1503PTH	–	TA12S05-0103GC
	4.6	TA12S05-H346PTH	TA12S05-0546PTH	TA12S05-L546PTH	TA12S05-1046PTH	TA12S05-1546PTH	TA12S05-2546PTH	TA12S05-0104GC
YMC-Triart Bio C18	2.1	TA30S05-H3Q1PTH	TA30S05-05Q1PTH	TA30S05-L5Q1PTH	TA30S05-10Q1PTH	TA30S05-15Q1PTH	–	TA30S05-01Q1GC
	3.0	–	TA30S05-0503PTH	TA30S05-L503PTH	TA30S05-1003PTH	TA30S05-1503PTH	–	TA30S05-0103GC
	4.6	TA30S05-H346PTH	TA30S05-0546PTH	TA30S05-L546PTH	TA30S05-1046PTH	TA30S05-1546PTH	TA30S05-2546PTH	TA30S05-0104GC
YMC-Triart C8	2.1	T012S05-H3Q1PTH	T012S05-05Q1PTH	T012S05-L5Q1PTH	T012S05-10Q1PTH	T012S05-15Q1PTH	–	T012S05-01Q1GC
	3.0	–	T012S05-0503PTH	T012S05-L503PTH	T012S05-1003PTH	T012S05-1503PTH	–	T012S05-0103GC
	4.6	T012S05-H346PTH	T012S05-0546PTH	T012S05-L546PTH	T012S05-1046PTH	T012S05-1546PTH	T012S05-2546PTH	T012S05-0104GC
YMC-Triart Bio C4	2.1	TB30S05-H3Q1PTH	TB30S05-05Q1PTH	TB30S05-L5Q1PTH	TB30S05-10Q1PTH	TB30S05-15Q1PTH	–	TB30S05-01Q1GC
	3.0	–	TB30S05-0503PTH	TB30S05-L503PTH	TB30S05-1003PTH	TB30S05-1503PTH	–	TB30S05-0103GC
	4.6	TB30S05-H346PTH	TB30S05-0546PTH	TB30S05-L546PTH	TB30S05-1046PTH	TB30S05-1546PTH	TB30S05-2546PTH	TB30S05-0104GC
Hydrosphere C18	2.1	HS12S05-H3Q1QT	HS12S05-05Q1QT	HS12S05-L5Q1QT	HS12S05-10Q1QT	HS12S05-15Q1QT	HS12S05-25Q1QT	HS12S05-01Q1GC
	3.0	HS12S05-H303QT	HS12S05-0503QT	HS12S05-L503QT	HS12S05-1003QT	HS12S05-1503QT	HS12S05-2503QT	HS12S05-0103GC
	4.6	HS12S05-0346WT	HS12S05-0546WT	HS12S05-L546WT	HS12S05-1046WT	HS12S05-1546WT	HS12S05-2546WT	HS12S05-0104GC
YMCbasic (eq. C8)	2.1	BA99S05-H3Q1QT	BA99S05-05Q1QT	BA99S05-L5Q1QT	BA99S05-10Q1QT	BA99S05-15Q1QT	BA99S05-25Q1QT	BA99S05-01Q1GC
	3.0	BA99S05-H303QT	BA99S05-0503QT	BA99S05-L503QT	BA99S05-1003QT	BA99S05-1503QT	BA99S05-2503QT	BA99S05-0103GC
	4.6	BA99S05-0346WT	BA99S05-0546WT	BA99S05-L546WT	BA99S05-1046WT	BA99S05-1546WT	BA99S05-2546WT	BA99S05-0104GC

*Guard cartridge holder required, part no. XPGCH-Q1

RP – Ordering information

5 µm metal-free HPLC columns

Phase	Column ID [mm]	Column length [mm]		
		50	100	150
YMC-Triart C18	2.1	TA12S05-05Q1PTP	TA12S05-10Q1PTP	TA12S05-15Q1PTP
	4.6	TA12S05-0546PTP	TA12S05-1046PTP	TA12S05-1546PTP
YMC-Triart Bio C18	2.1	TA30S05-05Q1PTP	TA30S05-10Q1PTP	TA30S05-15Q1PTP
	4.6	TA30S05-0546PTP	TA30S05-1046PTP	TA30S05-1546PTP
YMC-Triart C8	2.1	T012S05-05Q1PTP	T012S05-10Q1PTP	T012S05-15Q1PTP
	4.6	T012S05-0546PTP	T012S05-1046PTP	T012S05-1546PTP
YMC-Triart Bio C4	2.1	TB30S05-05Q1PTP	TB30S05-10Q1PTP	TB30S05-15Q1PTP
	4.6	TB30S05-0546PTP	TB30S05-1046PTP	TB30S05-1546PTP

Special column connectors required.

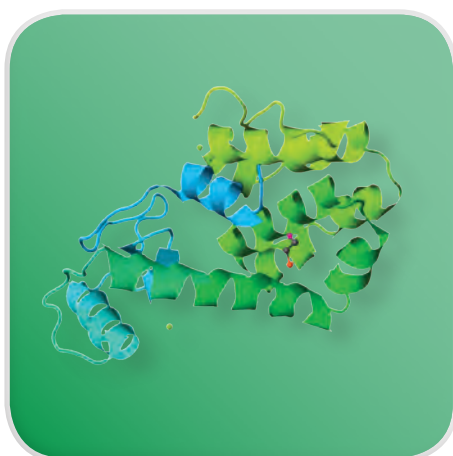
5 µm YMC-Actus high-throughput semipreparative columns

Phase	Column ID [mm]	Column length [mm]					Guard cartridges* with 10 mm length
		50	75	100	150	250	(pack of 5)
YMC-Triart C18	20	TA12S05-0520WX	TA12S05-L520WX	TA12S05-1020WX	TA12S05-1520WX	TA12S05-2520WX	TA12S05-0120CCN
	30	TA12S03-0530WX	TA12S03-L530WX	TA12S03-1030WX	TA12S03-1530WX	TA12S03-2530WX	TA12S05-0130CCN
	50	TA12S05-0553DX	–	TA12S05-1053DX	TA12S05-1553DX	TA12S05-2553DX	TA12S05-0553DXG**
YMC-Triart Bio C18	20	TA30S05-0520WX	TA30S05-L520WX	TA30S05-1020WX	TA30S05-1520WX	TA30S05-2520WX	TA30S05-0120CCN
	30	TA30S03-0530WX	TA30S03-L530WX	TA30S03-1030WX	TA30S03-1530WX	TA30S03-2530WX	TA30S05-0130CCN
	50	TA30S05-0553DX	–	TA30S05-1053DX	TA30S05-1553DX	TA30S05-2553DX	TA30S05-0553DXG**
YMC-Triart C8	20	T012S05-0520WX	T012S05-L520WX	T012S05-1020WX	T012S05-1520WX	T012S05-2520WX	T012S05-0120CCN
	30	T012S03-0530WX	T012S03-L530WX	T012S03-1030WX	T012S03-1530WX	T012S03-2530WX	T012S05-0130CCN
	50	T012S05-0553DX	–	T012S05-1053DX	T012S05-1553DX	T012S05-2553DX	T012S05-0553DXG**
YMC-Triart Bio C4	20	TB30S05-0520WX	TB30S05-L520WX	TB30S05-1020WX	TB30S05-1520WX	TB30S05-2520WX	TB30S05-0120CCN
	30	TB30S03-0530WX	TB30S03-L530WX	TB30S03-1030WX	TB30S03-1530WX	TB30S03-2530WX	TB30S05-0130CCN
	50	TB30S05-0553DX	–	TB30S05-1053DX	TB30S05-1553DX	TB30S05-2553DX	TB30S05-0553DXG**
Hydrosphere C18	20	HS12S05-0520WX	HS12S05-L520WX	HS12S05-1020WX	HS12S05-1520WX	HS12S05-2520WX	HS12S05-0120CCN
	30	HS12S05-0530WX	HS12S05-L530WX	HS12S05-1030WX	HS12S05-1530WX	HS12S05-2530WX	HS12S05-0130CCN
	50	HS12S05-0553DX	–	HS12S05-1053DX	HS12S05-1553DX	HS12S05-2553DX	–
YMbasic (eq. C8)	20	BA99S05-0520WX	BA99S05-L520WX	BA99S05-1020WX	BA99S05-1520WX	BA99S05-2520WX	BA99S05-0120CCN
	30	BA99S05-0530WX	BA99S05-L530WX	BA99S05-1030WX	BA99S05-1530WX	BA99S05-2530WX	BA99S05-0130CCN
	50	BA99S05-0553DX	–	BA99S05-1053DX	BA99S05-1553DX	BA99S05-2553DX	–

*Guard cartridge holder required, part no. XPGHF2P20ID (20 mm ID)
XPGHF2P30ID (30 mm ID)
no holder required for 50 mm



SEC



SEC – UHPLC / HPLC Selectivities

- Applicable to proteins, antibodies, their fragments and peptides
- Also applicable to carbohydrates and nucleic acid components
- Excellent reproducibility with minimal secondary interactions
- 2 µm for UHPLC
- Cost effective

	YMC-Pack Diol-60	YMC-Pack Diol-120	YMC-Pack Diol-200	YMC-Pack Diol-300	YMC-SEC MAB
	For peptides and small proteins	For intermediate proteins	For large proteins	For very large proteins	For antibodies, fragments and aggregates
Base particle	Silica				
Particle Size / µm	3, 5	3,5	2, 3, 5	2, 3, 5	(<2), 3
Pore Size / nm	6	12	20	30	25
Modification	Dihydroxypropyl				
Temperature range	40 °C				
Pressure limit	2 µm: 45 MPa (6,525 psi); 3/5 µm: 20 MPa (3,000 psi)				3 µm: 14 MPa (2,030 psi)

Column Selection Tool

for MW < 10,000

● YMC-Pack Diol-**60**

for MW 5,000 to 100,000

● YMC-Pack Diol-**120**

for MW 5,000 to 300,000

● YMC-Pack Diol-**200**

for MW 10,000 to 700,000

● YMC-**SEC MAB**

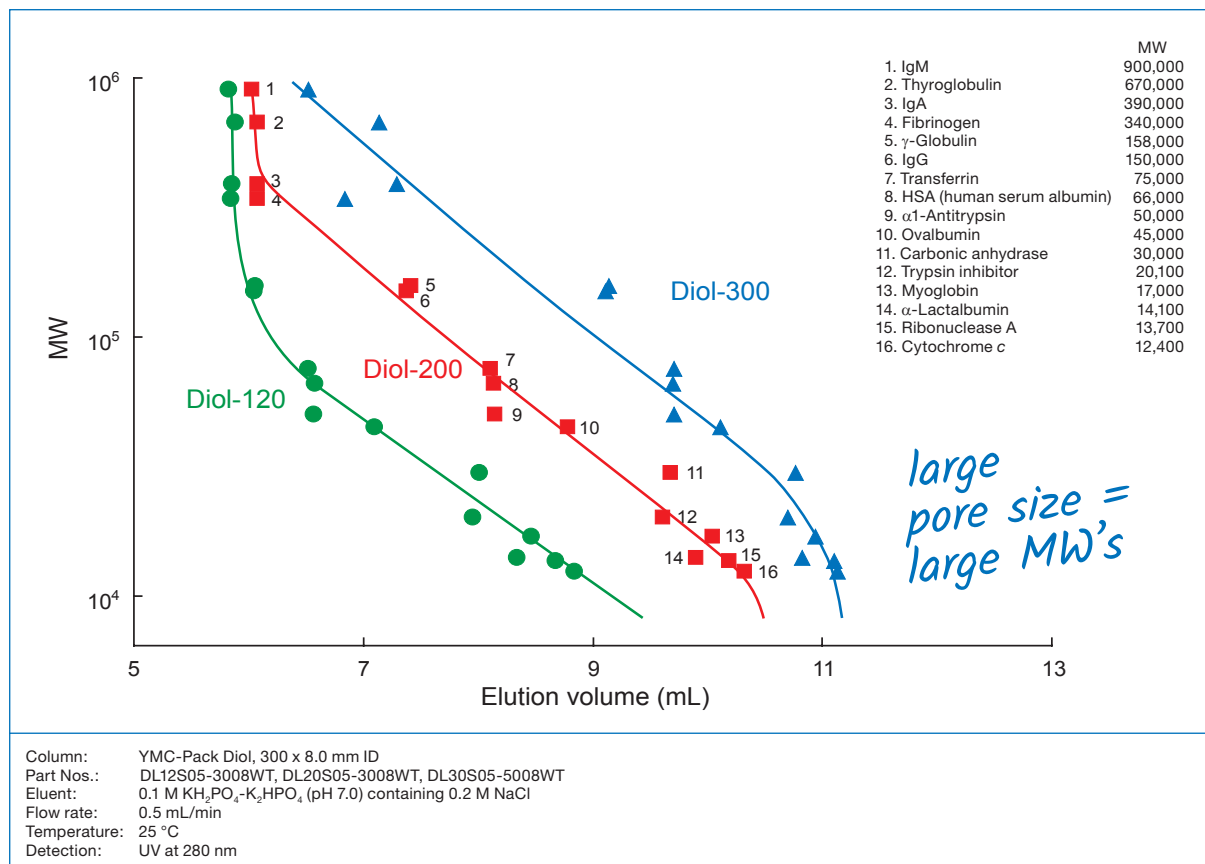
for MW 20,000 to 1,000,000

● YMC-Pack Diol-**300**

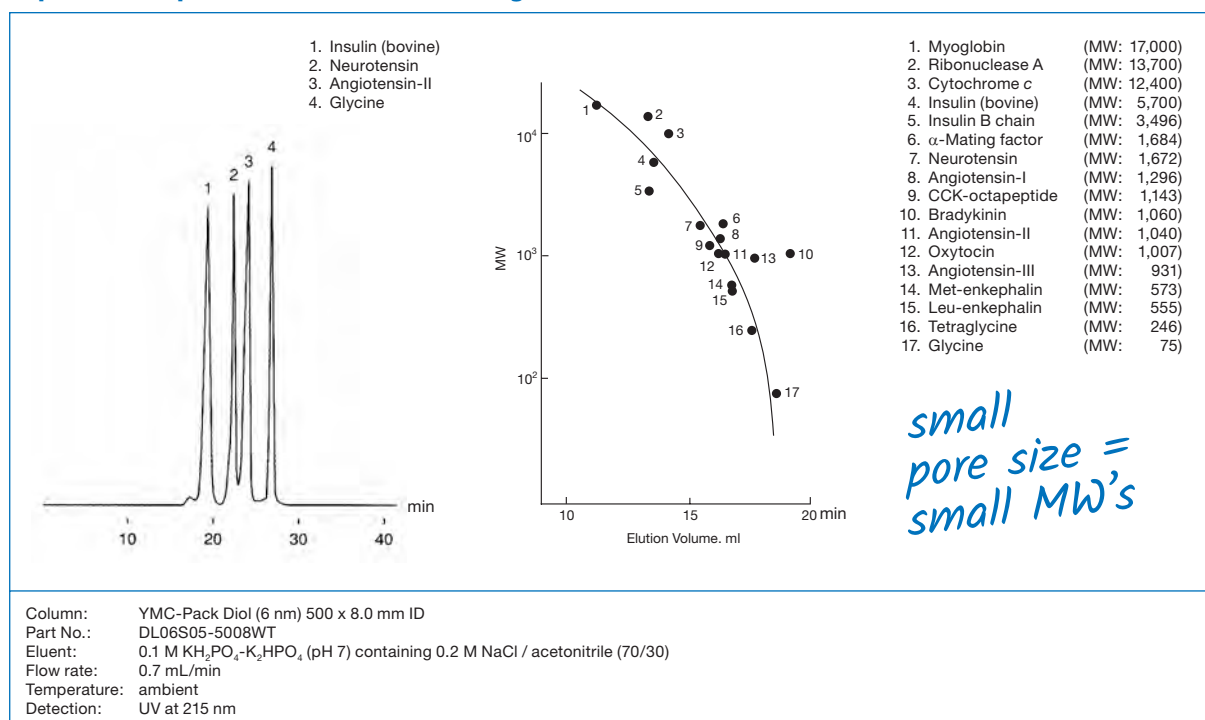
SEC – YMC-Pack Diol: Phase selection for proteins

Phases for different MW ranges

For separation of proteins with molecular weights from 10,000 to several 100,000 Da



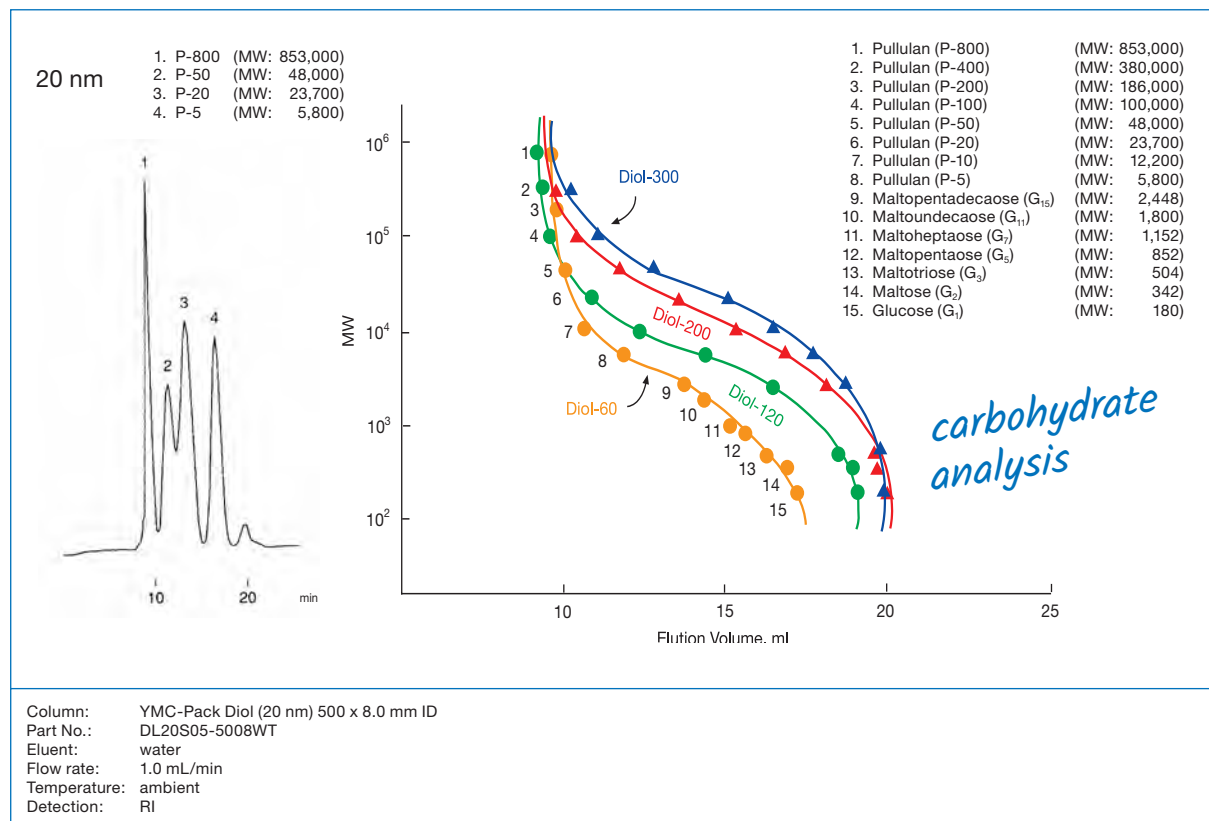
Separation of proteins with molecular weights below 10,000 Da



SEC – YMC-Pack Diol: Phase selection for carbohydrates

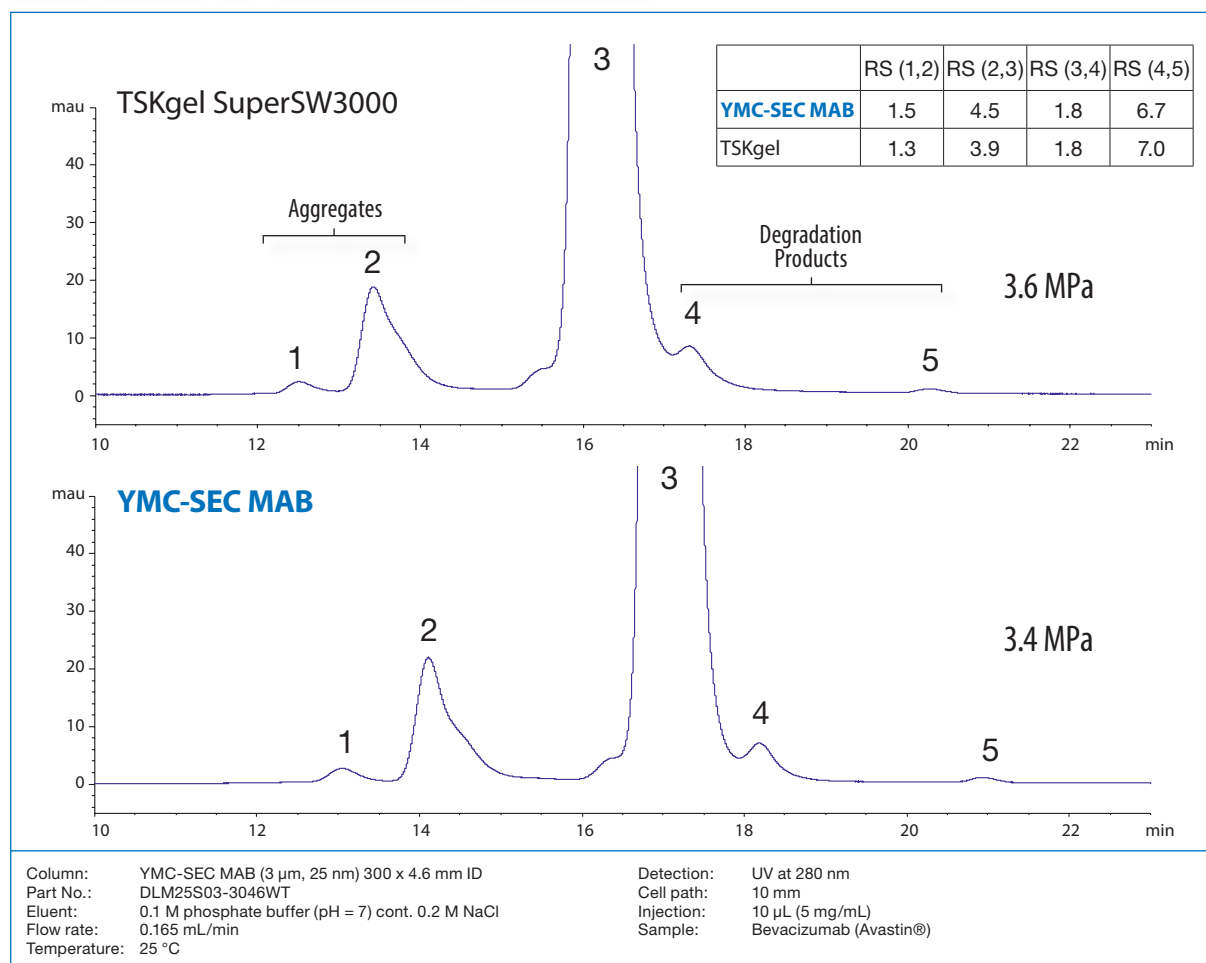
Phases for different MW ranges

For molecular weight determination of oligosaccharides and polysaccharides

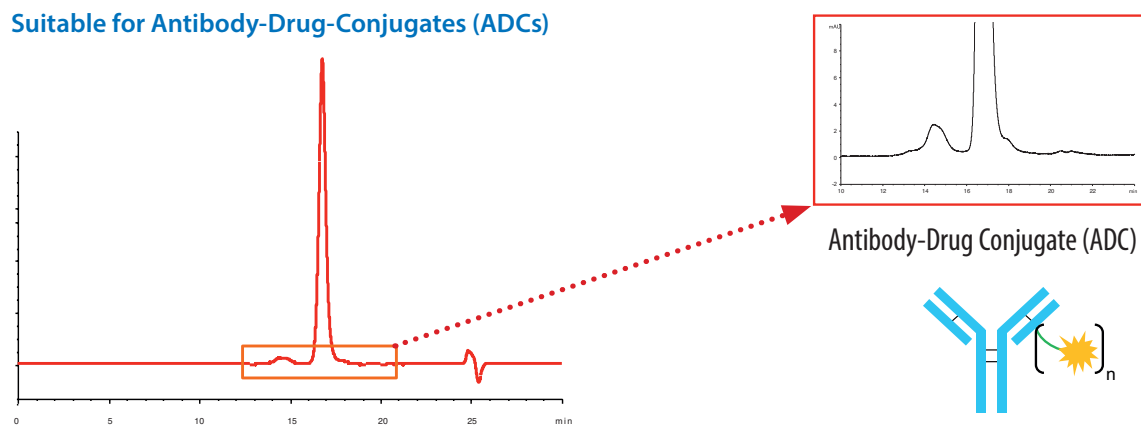


SEC – YMC-SEC MAB: MAb & ADC analysis

Ideal choice for monoclonal antibodies



Suitable for Antibody-Drug-Conjugates (ADCs)



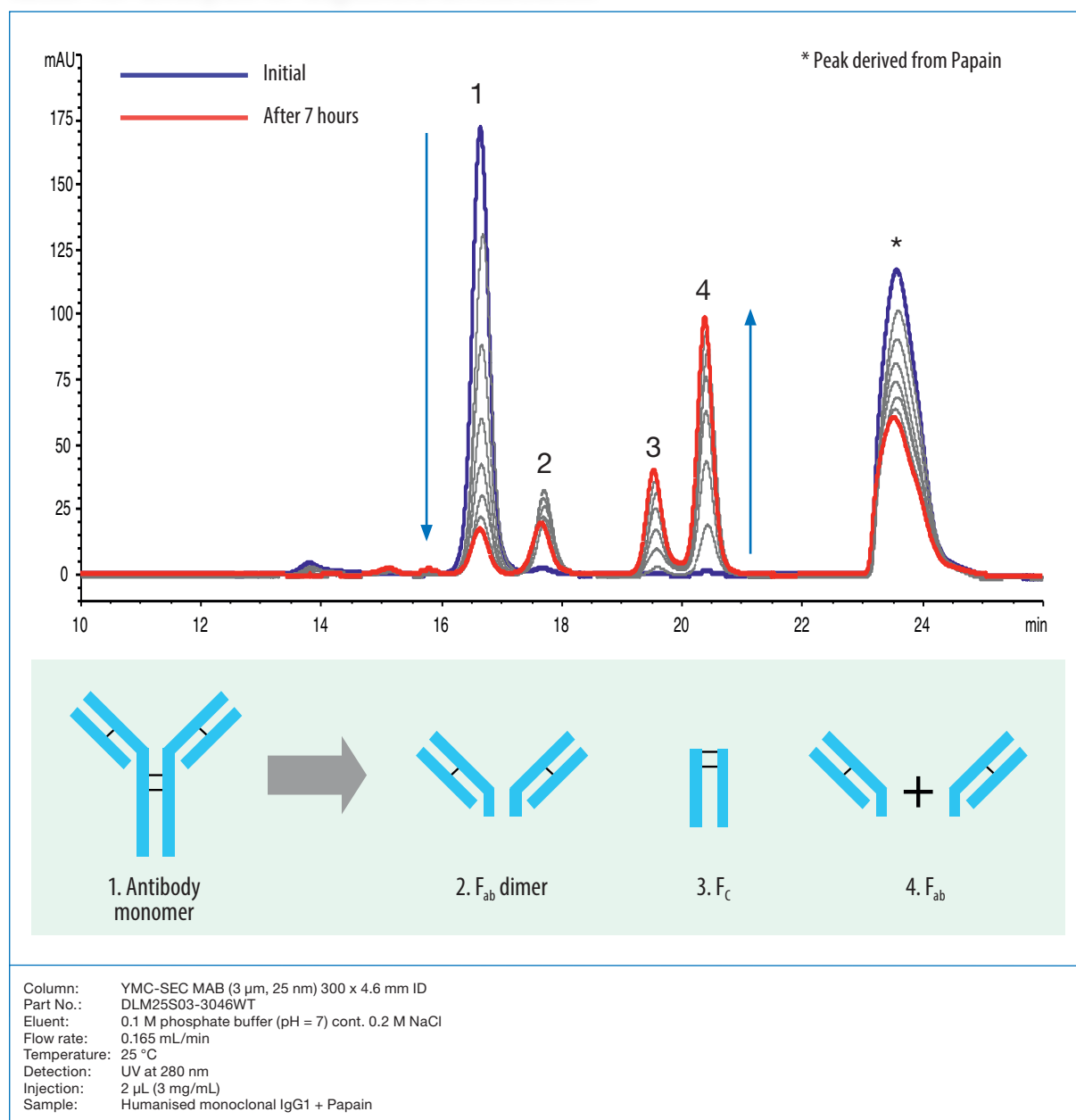
Column: YMC-SEC MAB (3 μ m, 25 nm) 300 x 4.6 mm ID
 Part No.: DLM25S03-3046WT
 Eluent: 0.1 M phosphate buffer (pH = 7) cont. 0.2 M NaCl /
 2-propanol (85 / 15)
 Flow rate: 0.165 mL/min

Temperature: 25 °C
 Detection: UV at 280 nm
 Injection: 4 μ L (2.5 mg/mL)
 Sample: SigmaMAb Antibody Drug Conjugate Mimic

YMC-SEC MAB is also suitable for the analysis of Antibody-Drug Conjugates (ADC). The addition of an organic solvent to the mobile phase can improve the results obtained for ADC analysis.

SEC – YMC-SEC MAB: Digest monitoring

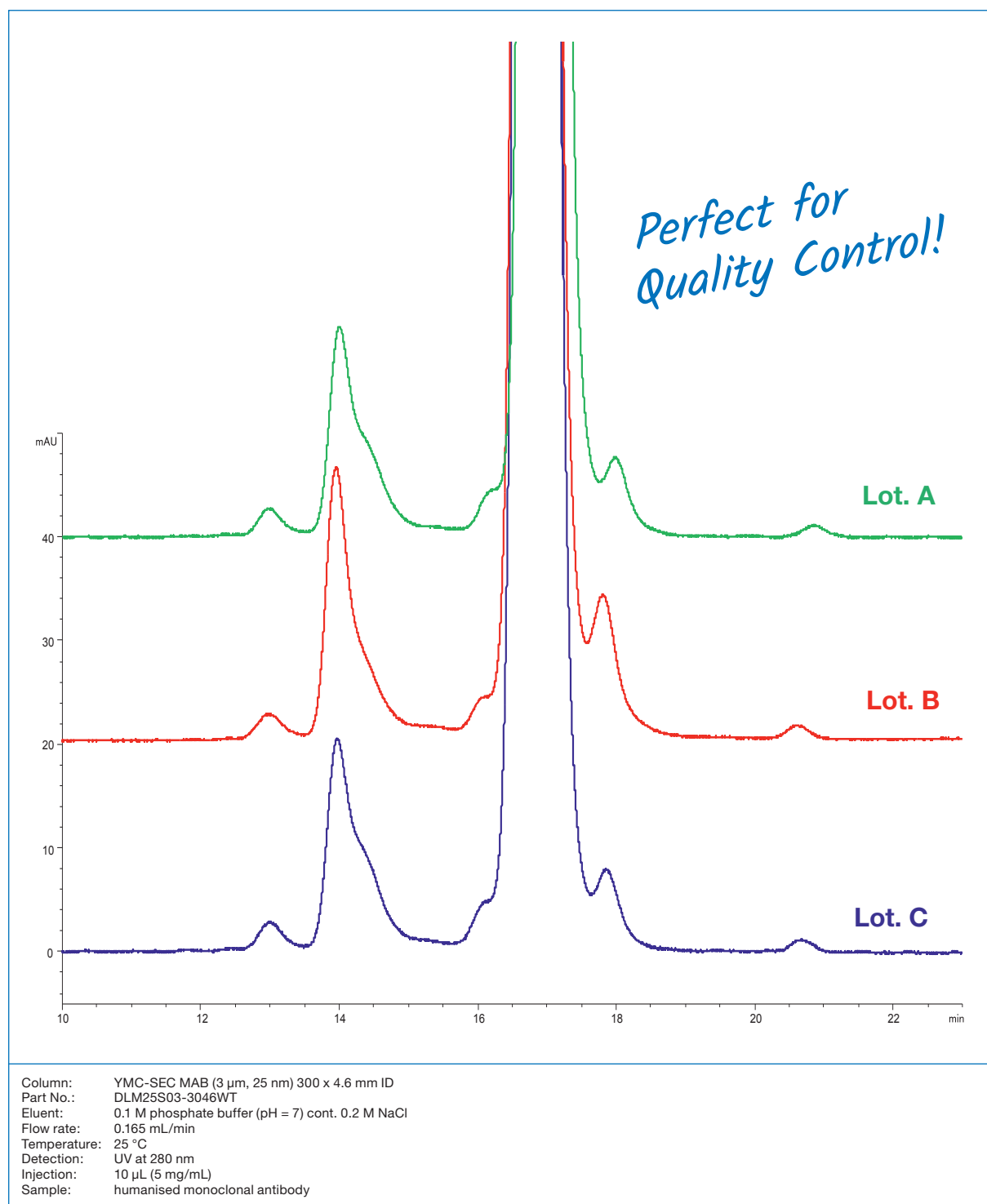
Ideal for analysis of digested antibodies



Ideal for the analysis of fragments/degradation products of antibodies: Digestion of a monoclonal antibody with papain was monitored for 7 hours. The peak of the monomer decreased as digestion proceeded, while peaks for degradation products increased.

SEC – YMC-SEC MAB: Reproducibility

Excellent lot-to-lot reproducibility

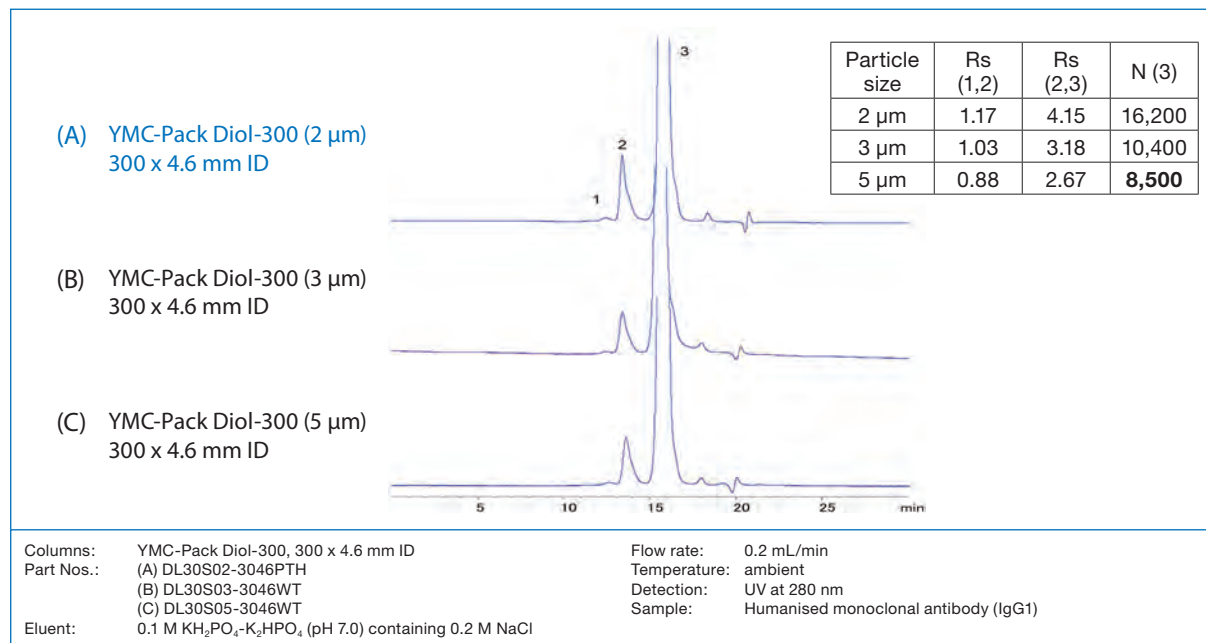


YMC-SEC MAB provides excellent reproducibility for the separation of monomer and aggregates as well as for monomer and their fragments, making it very effective for quality control of antibody drugs.

SEC – YMC-Pack Diol: Resolution & throughput

Benefits of using smaller particles

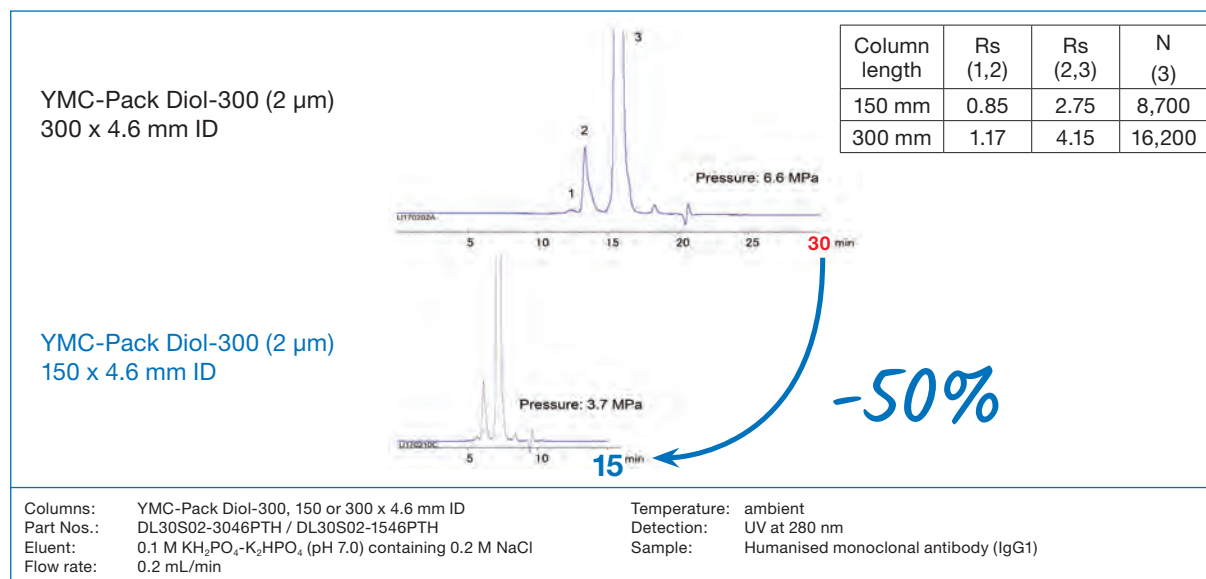
Higher resolution for analysis of monoclonal antibodies



All three particle sizes show identical separation patterns for monoclonal antibody analysis. This allows easy method transfer between HPLC and UHPLC. A method developed using conventional HPLC can be directly transferred to UHPLC using a 2 μm YMC-Pack Diol

column. YMC-Pack Diol UHPLC columns greatly improve the resolution between aggregates and the monomer peak. In addition, a shoulder peak which can be observed after the monomer peak can be partially separated using the 2 μm column.

High throughput analysis of monoclonal antibodies

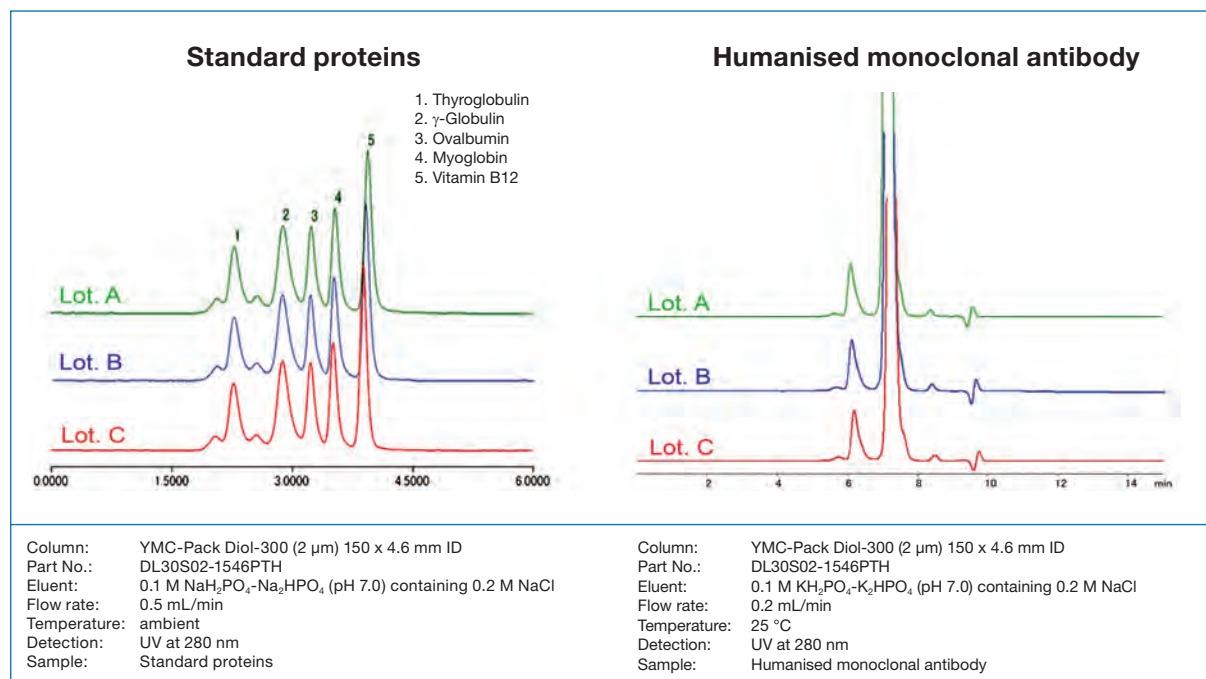


By using a 150 mm length column, 50% shorter run times can be achieved with the same resolution as for a 5 μm 300 mm length column (compare upper and lower chromatograms). This allows an increase in throughput to be achieved. The backpressure is only 6.6 MPa, even for the 300 mm column. Therefore, YMC-Pack Diol 2 μm columns can be used with both UHPLC and HPLC systems.

SEC – YMC-Pack Diol: Reproducibility & stability

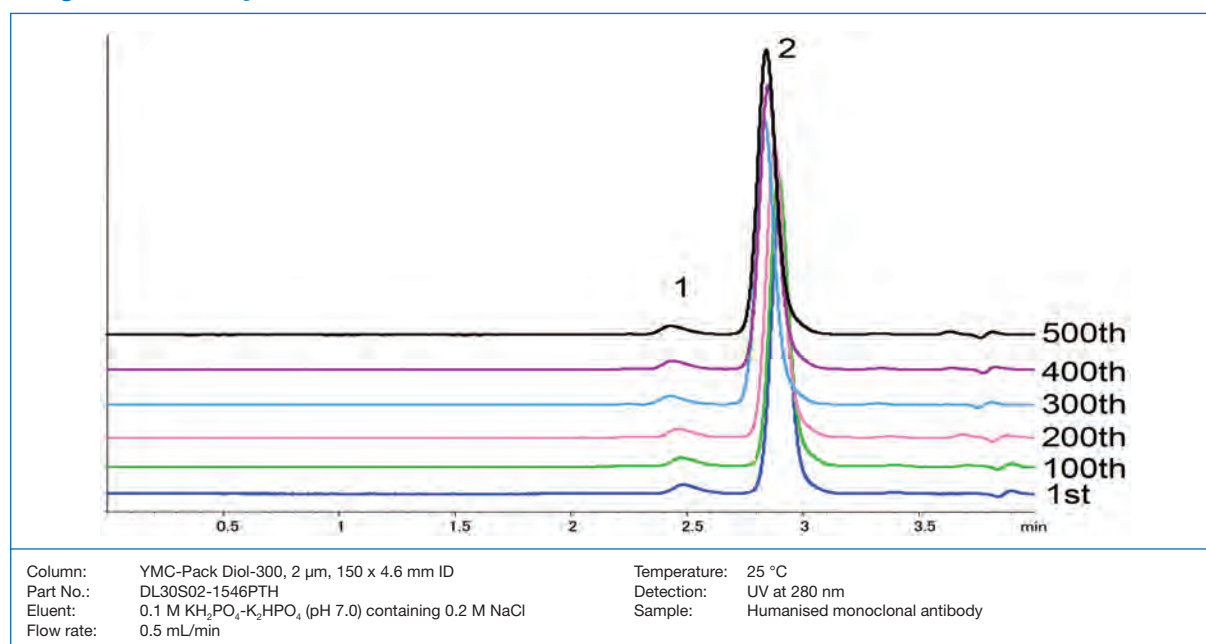
Reproducibility and stability data

Excellent batch to batch reproducibility



YMC-Pack Diol UHPLC columns have excellent batch-to-batch reproducibility. This makes YMC-Pack Diol 2 μ m columns the ideal choice for the quality control of bio-based drugs including monoclonal antibodies.

Long-term stability



YMC-Pack Diol UHPLC columns maintain their performance for more than 500 injections of sample during monoclonal antibody analysis. This ensures reproducible and reliable quality control of bio-based drugs including monoclonal antibodies.

SEC – Ordering information

2 µm UHPLC columns

Phase	Column ID [mm]	Column length [mm]		Guard cartridges* with 10 mm length
		150	300	(pack of 5)
YMC-Pack Diol-200	4.6	DL20S02-1546PTH	DL20S02-3046PTH	DL20S02-0104GC
YMC-Pack Diol-300	4.6	DL30S02-1546PTH	DL30S02-3046PTH	DL30S02-0104GC

*Guard cartridge holder required, part no. XPGCH-Q1

3 µm HPLC columns

Phase	Column ID [mm]	Column length [mm]			Guard cartridges* with 10/30 mm length
		150	250	300	(pack of 5)
YMC-SEC MAB	4.6	DLM25S03-1546WT	–	DLM25S03-3046WT	DLM25S03-0104GC
	6.0	–	–	–	–
	8.0	–	–	DLM25S03-3008WT	–
YMC-Pack Diol-60	4.6	DL06S03-1546WT	DL06S03-2546WT	DL06S03-3046WT	DL06S03-0104GC
	6.0	–	–	DL06S03-3006WT	–
	8.0	DL06S03-1508WT	–	DL06S03-3008WT	DL06S03-0310WTG**
YMC-Pack Diol-120	4.6	DL12S03-1546WT	DL12S03-2546WT	DL12S03-3046WT	DL12S03-0104GC
	6.0	–	–	DL12S03-3006WT	–
	8.0	DL12S03-1508WT	–	DL12S03-3008WT	DL12S03-0310WTG**
YMC-Pack Diol-200	4.6	DL20S03-1546WT	DL20S03-2546WT	DL20S03-3046WT	DL20S03-0104GC
	6.0	–	–	DL20S03-3006WT	–
	8.0	DL20S03-1508WT	–	DL20S03-3008WT	DL20S03-0310WTG**
YMC-Pack Diol-300	4.6	DL30S03-1546WT	DL30S03-2546WT	DL30S03-3046WT	DL30S03-0104GC
	6.0	–	–	DL30S03-3006WT	–
	8.0	DL30S03-1508WT	–	DL30S03-3008WT	DL30S03-0310WTG**

*Guard cartridge holder required, part no. XPGCH-Q1

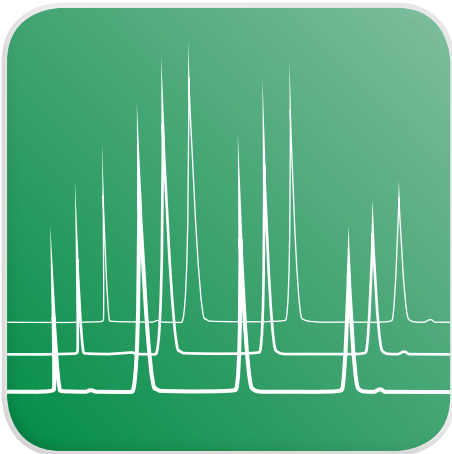
**no holder required for 30 x 8 mm ID guards

5 µm HPLC columns

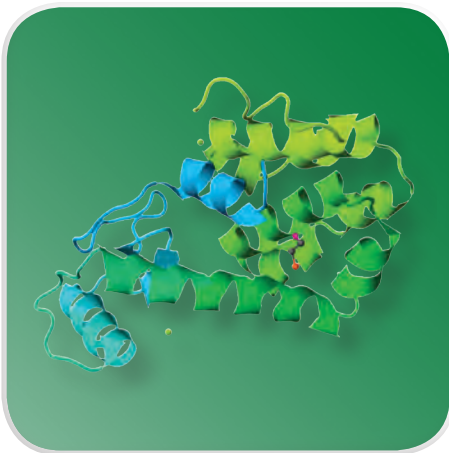
Phase	Column ID [mm]	Column length [mm]			Guard cartridges* with 10/30 mm length
		250	300	500	(pack of 5)
YMC-Pack Diol-60	4.6	DL06S05-2546WT	DL06S05-3046WT	–	DL06S05-0104GC
	6.0	DL06S05-2506WT	DL06S05-3006WT	DL06S05-5006WT	–
	8.0	–	DL06S05-3008WT	DL06S05-5008WT	DL06S05-0308WTG**
	10.0	DL06S05-2510WT	DL06S05-3010WT	DL06S05-5010WT	DL06S05-0310WTG**
YMC-Pack Diol-120	4.6	DL12S05-2546WT	DL12S05-3046WT	–	DL12S05-0104GC
	6.0	DL12S05-2506WT	DL12S05-3006WT	DL12S05-5006WT	–
	8.0	–	DL12S05-3008WT	DL12S05-5008WT	DL12S05-0308WTG**
	10.0	DL12S05-2510WT	DL12S05-3010WT	DL12S05-5010WT	DL12S05-0310WTG**
YMC-Pack Diol-200	4.6	DL20S05-2546WT	DL20S05-3046WT	–	DL20S05-0104GC
	6.0	DL20S05-2506WT	DL20S05-3006WT	DL20S05-5006WT	–
	8.0	–	DL20S05-3008WT	DL20S05-5008WT	DL20S05-0308WTG**
	10.0	DL20S05-2510WT	DL20S05-3010WT	DL20S05-5010WT	DL20S05-0310WTG**
YMC-Pack Diol-300	4.6	DL30S05-2546WT	DL30S05-3046WT	–	DL30S05-0104GC
	6.0	DL30S05-2506WT	DL30S05-3006WT	DL30S05-5006WT	–
	8.0	–	DL30S05-3008WT	DL30S05-5008WT	DL30S05-0308WTG**
	10.0	DL30S05-2510WT	DL30S05-3010WT	DL30S05-5010WT	DL30S05-0310WTG**

*Guard cartridge holder required, part no. XPGCH-Q1

**no holder required for 30 x 8 mm ID guards

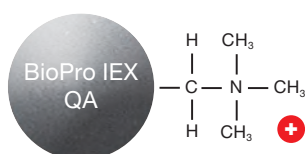


IEX

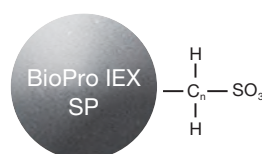


IEX – Bio Pro Series

- porous or non-porous hydrophilic polymers
- high binding capacity and recovery of biomolecules
- very high resolution
- low nonspecific adsorption
- excellent reproducibility



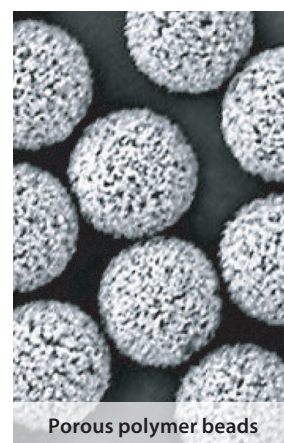
**strong anion
exchanger**



**strong cation
exchanger**

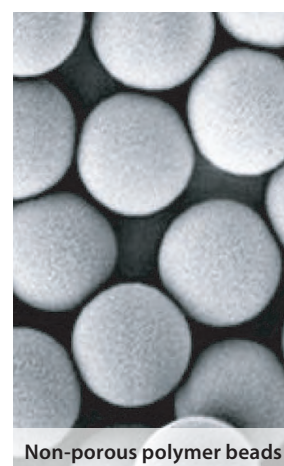
	BioPro IEX QA	BioPro IEX SP
Matrix	hydrophilic polymer (polymethacrylate)	hydrophilic polymer (polymethacrylate)
Particle size / μm	5	5
Pore size / nm	100	100
Charged group	$-\text{CH}_2\text{N}^+(\text{CH}_3)_3$	$-(\text{CH}_2)_3\text{SO}_3^-$
Counter ion	Cl^-	Na^+
Available pH range	2.0 – 12.0	2.0 – 12.0
Temperature range	4 – 60 °C	
Pressure limit	2.5 – 3.5 MPa (360 – 510 psi)	
Column hardware	PEEK	

Also available in 10, 20, 30 or 75 μm for preparative scale



Porous polymer beads

	BioPro IEX QF	BioPro IEX SF
Matrix	hydrophilic polymer (polymethacrylate)	hydrophilic polymer (polymethacrylate)
Particle size / μm	3; 5	3; 5
Pore size / nm	non-porous	non-porous
Charged group	$-\text{CH}_2\text{N}^+(\text{CH}_3)_3$	$-(\text{CH}_2)_3\text{SO}_3^-$
Counter ion	Cl^-	Na^+
Available pH range	2.0 - 12.0	2.0 - 12.0
Temperature range	4 – 60 °C	
Pressure limit	3 μm : 25 MPa (3,625 psi) 5 μm : 6 – 12 MPa (870 – 1,740 psi)	
Column hardware	PEEK	



Non-porous polymer beads

General

YMC's BioPro IEX series of ion exchange columns are available in QA and SP chemistries, based on 5 μm porous (QA or SP columns) or on 3 or 5 μm non-porous (QF and SF columns) hydrophilic polymer beads.

The porous materials offer excellent binding capacity with exceptionally high efficiency and low operating pressure, whilst the non-porous particles offer high efficiency, very high resolution and low operating pressures.

IEX – BioPro IEX: Reproducibility & DBC

High binding capacity and high recovery for porous type

The porous versions of YMC's BioPro IEX show high dynamic binding capacity and excellent recovery, making them useful for semi-preparative separations of proteins and antibodies.

Comparison of dynamic binding capacity (DBC) for BSA

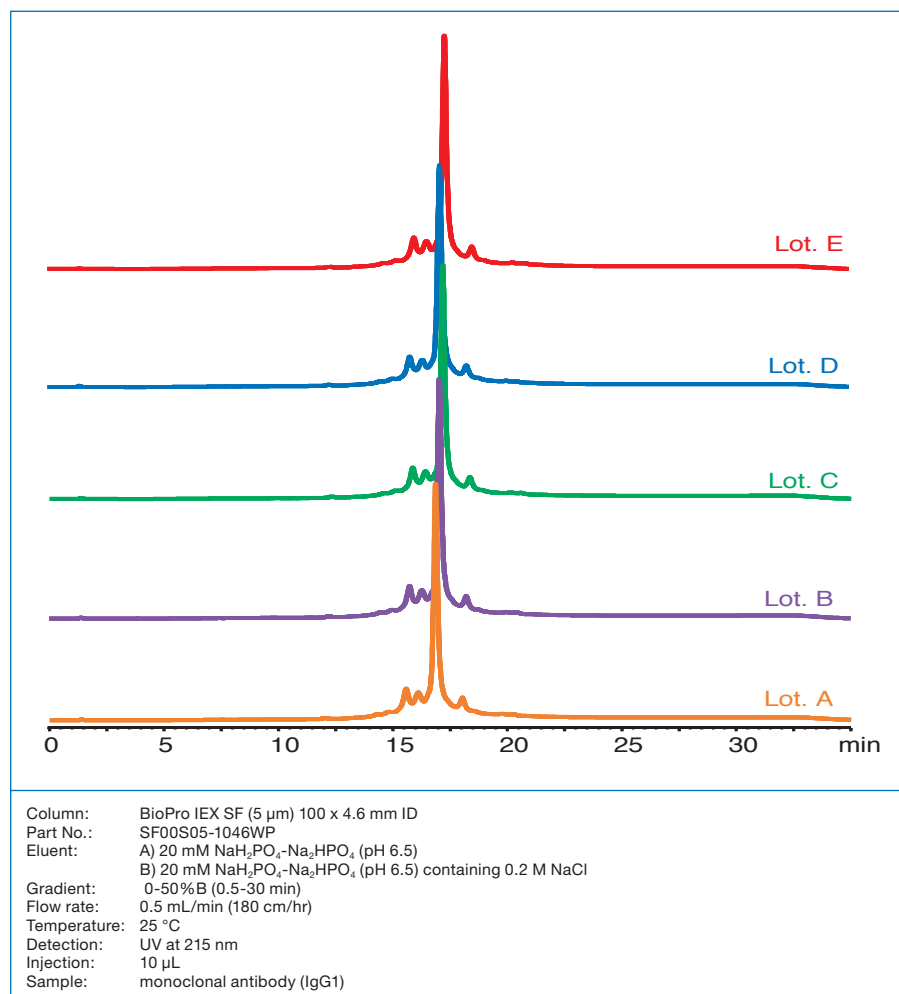
	Dynamic binding capacity (mg/mL-gel, 10% breakthrough)	Eluted amount (mg/mL-gel)	Recovery* (%)
BioPro IEX QA	126	120	95
Mono Q (GE Healthcare)	100	35	35
BioAssist Q (Tosoh Bioscience)	73	58	79

* Recovery: (Eluted amount/Dynamic binding capacity) x 100

*High
recovery
rates for
BioPro IEX*

Compared with conventional porous polymer anion exchange columns, BioPro IEX QA provides higher DBC and recovery rates. This indicates that BioPro IEX has a much lower nonspecific adsorption compared to conventional columns.

Excellent batch-to-batch reproducibility

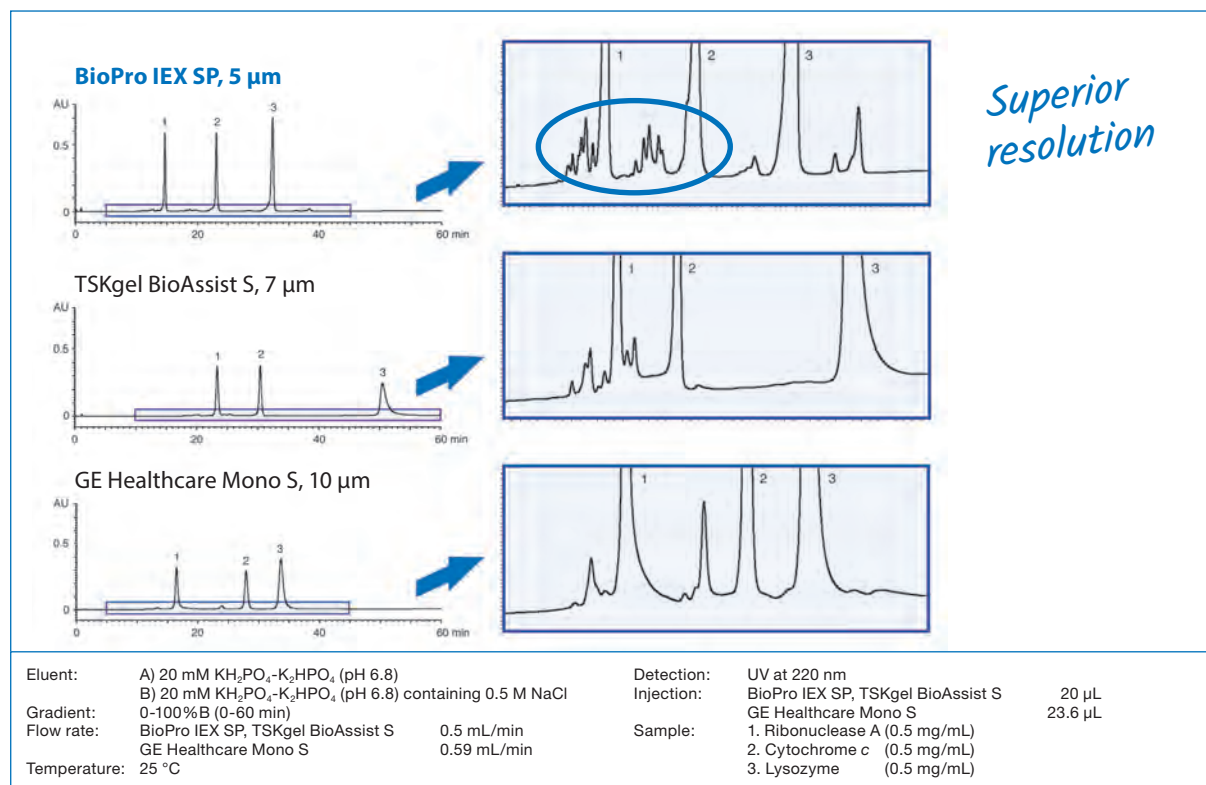


BioPro IEX SF columns exhibit excellent batch-to-batch reproducibility for MAb analysis with resolution of peaks for small charge variants. All gel batches are inspected by rigorous quality control tests, including HPLC analysis of MAb, and must meet the required criteria before release. BioPro IEX columns are the best choice for the quality control of MAb and other biopharmaceuticals.

IEX – BioPro IEX: Resolution & throughput

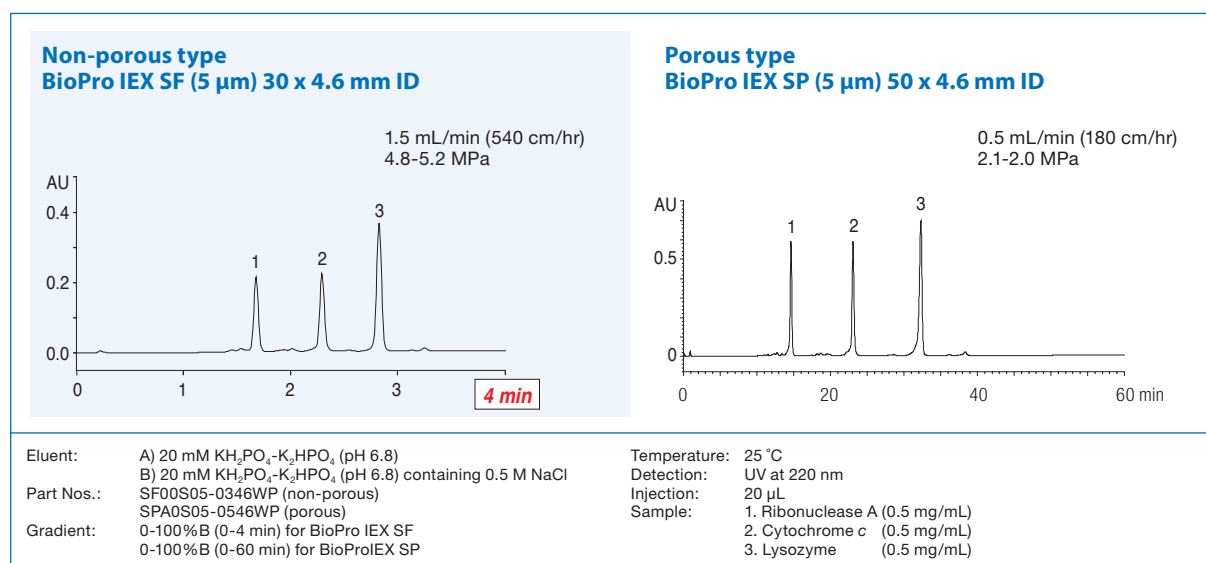
Superior resolution

Comparison of standard protein separation on BioPro IEX SP and commercial S type products



Only BioPro IEX is available in the smaller particle size and is therefore able to provide superior resolution.

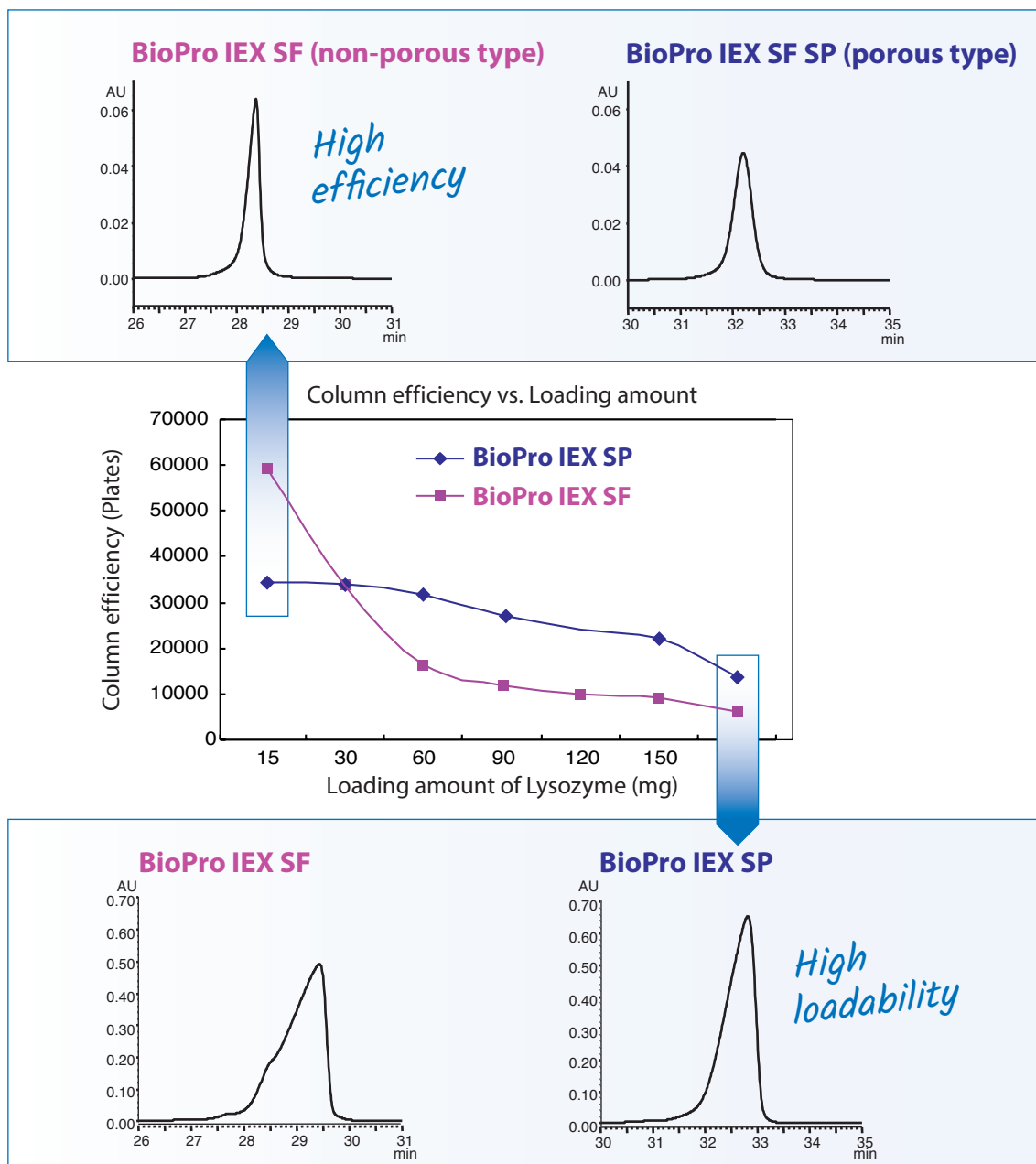
Ultra-high-throughput analysis with non-porous BioPro IEX



The high mechanical stability of non-porous polymer beads and the short column length allow faster elution of proteins at a higher flow rate without any loss of resolution.

Column efficiency and loadability

When to use porous and non-porous BioPro IEX



Eluent: A) 20 mM NaH_2PO_4 - Na_2HPO_4 (pH 6.8)
 B) 20 mM NaH_2PO_4 - Na_2HPO_4 (pH 6.8) containing 0.5 M NaCl
 Gradient: 0–100% B (0–60 min)
 Flow rate: 0.5 mL/min
 Temperature: 25 °C

Detection: UV at 280 nm
 Injection: 100 mL
 Sample: 1. Ribonuclease A
 2. Cytochrome c
 3. Lysozyme

BioPro IEX SF offers outstanding column efficiency at small amount of sample loading. Non-porous type of BioPro IEX columns are especially suitable for microscale analysis which requires higher resolution.

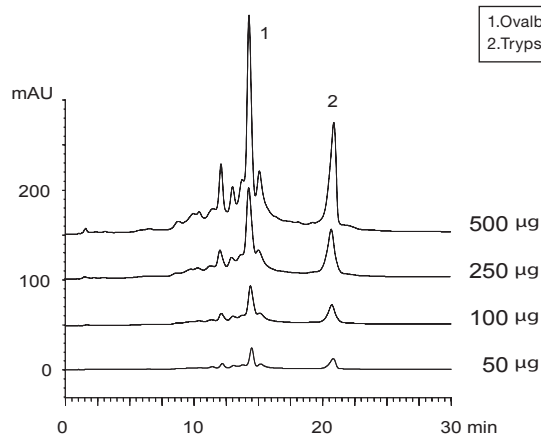
BioPro IEX SP maintains the good peak shape even when the loading amount increases. Porous type BioPro IEX columns with high capacity are useful for high-load analytical separations and laboratory-scale purification.

IEX – BioPro IEX: Loadability & recovery

High loadability

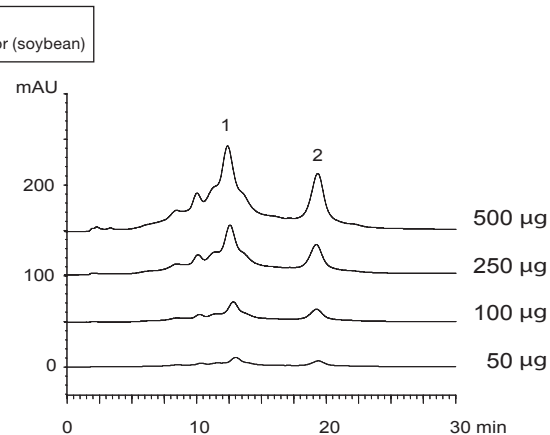
Loading study for BioPro IEX QA (porous)

BioPro IEX QA (5 μ m) 50 x 4.6 mm ID



Part No.: QAA0S05-0546WP
 Eluent: A) 20 mM Tris-HCl (pH 8.1)
 B) 20 mM Tris-HCl (pH 8.1) containing 0.5 M NaCl
 Gradient: 10-80%B (0-30 min)
 Flow rate: 0.5 mL/min
 Temperature: 25 $^{\circ}$ C
 Detection: UV at 280 nm
 Injection: 100 μ L

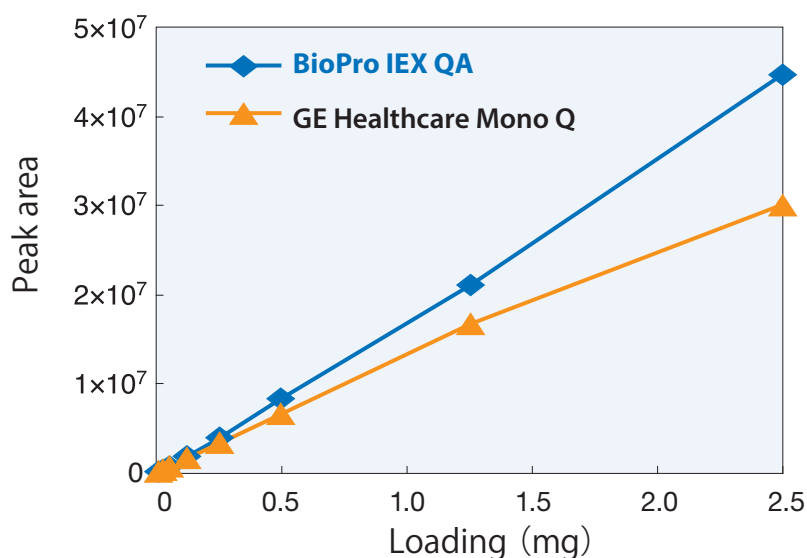
GE Healthcare Mono Q (10 μ m) 50 x 5.0 mm ID



Eluent: A) 20 mM Tris-HCl (pH 8.1)
 B) 20 mM Tris-HCl (pH 8.1) containing 0.5 M NaCl
 Gradient: 10-80%B (0-30 min)
 Flow rate: 0.5 mL/min
 Temperature: 25 $^{\circ}$ C
 Detection: UV at 280 nm
 Injection: 100 μ L

High recovery

Recovery of trypsin inhibitor



Eluent: A) 20 mM Tris-HCl (pH 8.1)
 B) 20 mM Tris-HCl (pH 8.1) containing 0.5 M NaCl
 Gradient: 10-80%B (0-30 min)
 Flow rate: 0.5 mL/min (180 cm/hr for 4.6 mm ID,
 150 cm/hr for 5.0 mm ID)

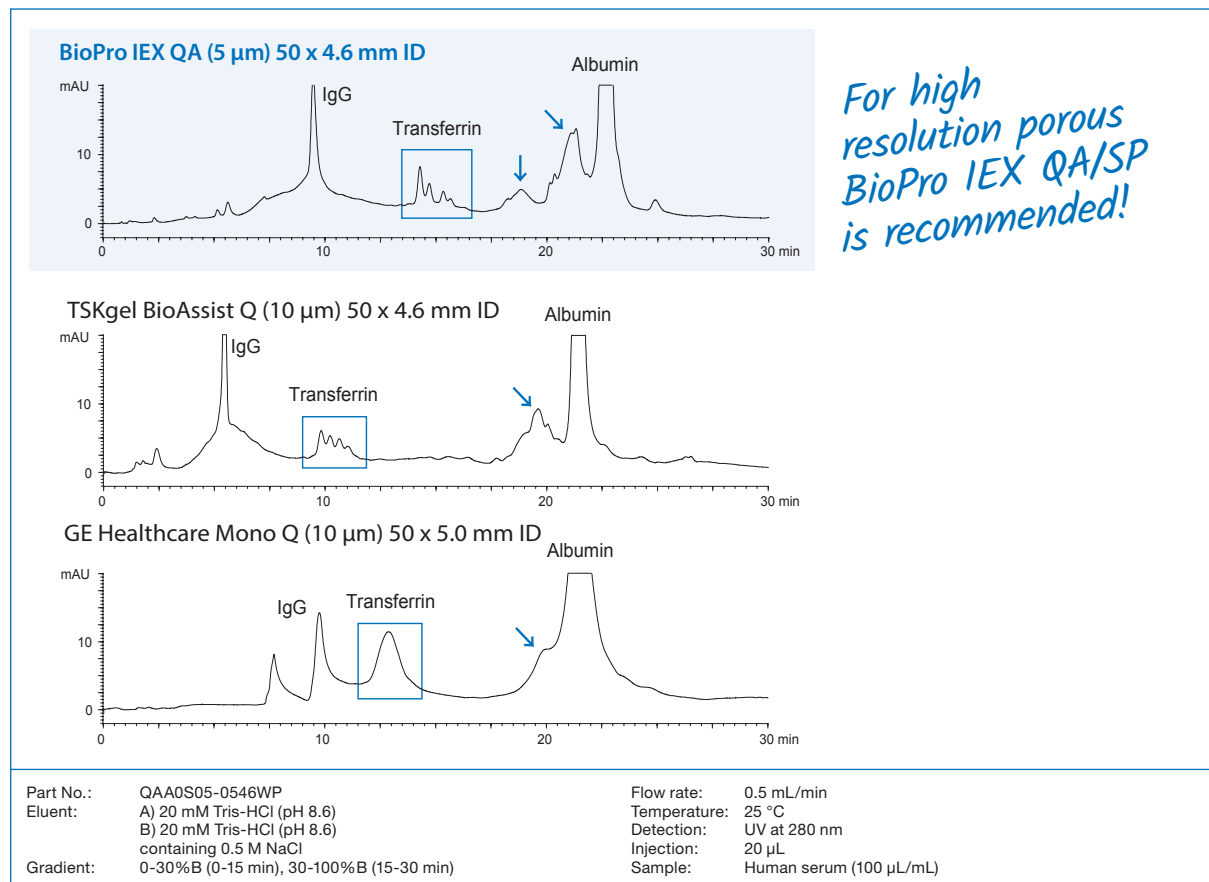
Temperature: 25 $^{\circ}$ C
 Detection: UV at 280 nm
 Injection: 100 μ L

BioPro IEX QA shows excellent resolution and peak shapes even when the loading amount is increased. The porous type BioPro IEX columns are suitable for laboratory-scale purification of proteins.

IEX – BioPro IEX: Challenging separations

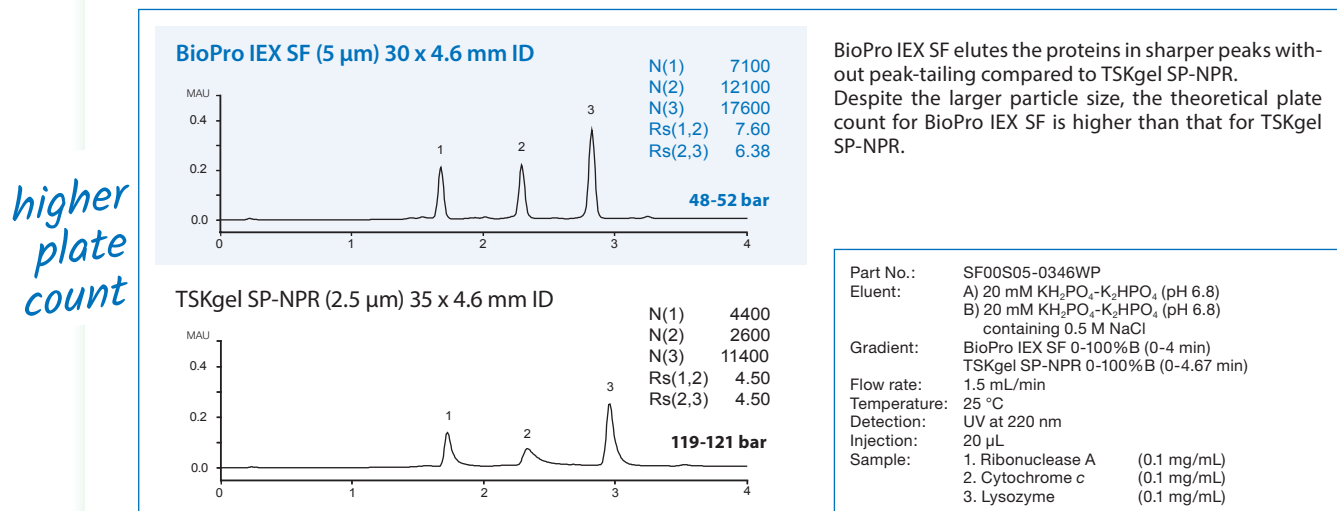
Protein separation in challenging matrices

Separation of proteins in human serum on BioPro IEX QA and commercial Q-type products



Better performance at lower backpressure

Comparison of standard protein separation on BioPro IEX SF and a commercial SP-type product

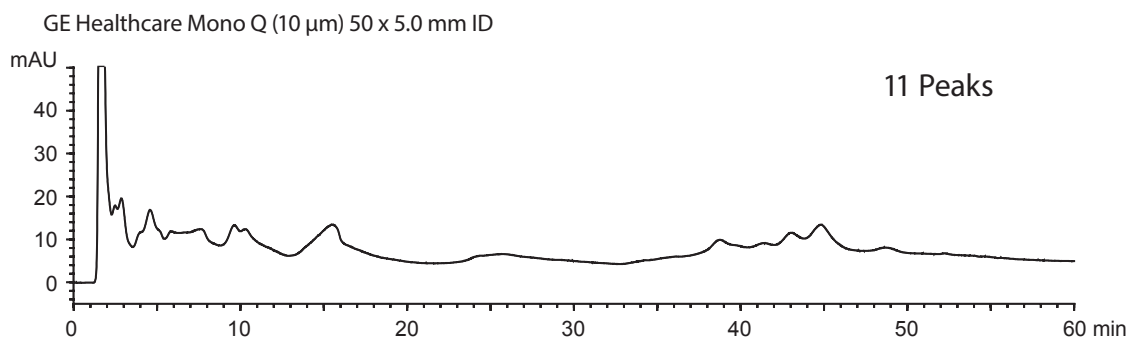
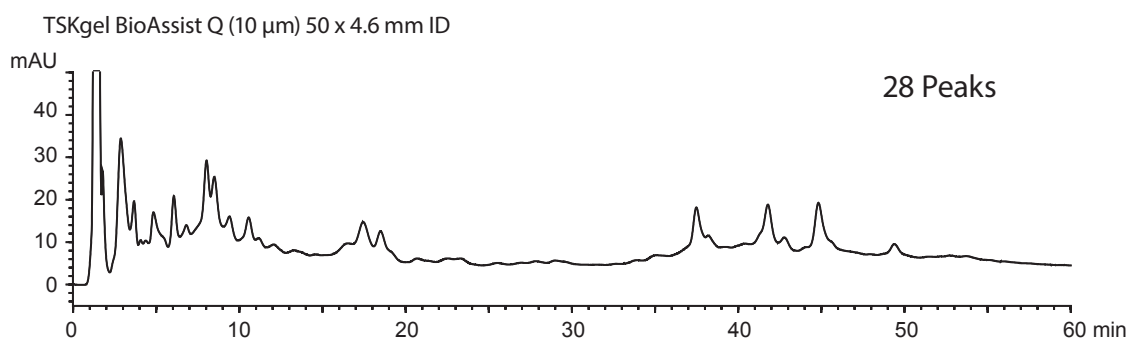
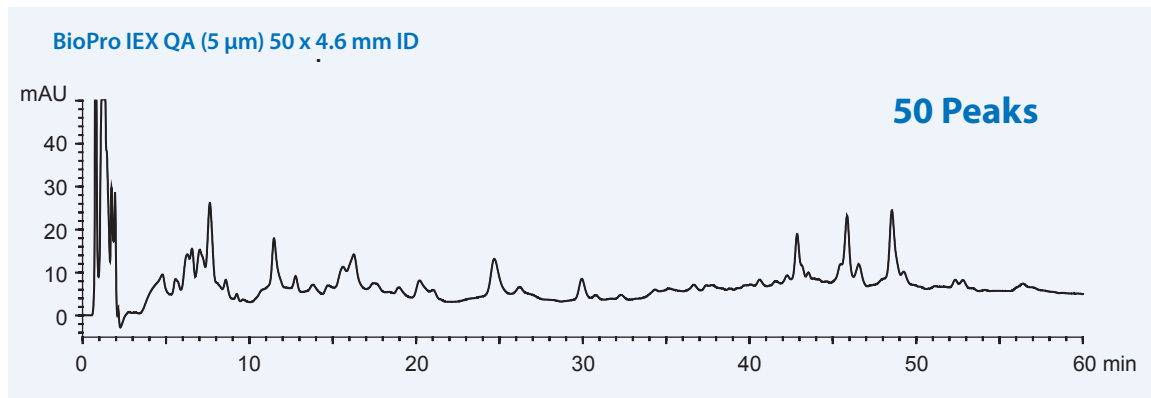


Compared to the competitor's column, BioPro IEX SF gives higher theoretical plate counts, excellent peak shapes, and lower backpressures. This makes BioPro IEX SF most suitable for high-throughput analysis.

IEX – BioPro IEX: Peptide mapping

Peptide mapping

Peptide mapping of tryptic digests of BSA with enhanced sensitivity

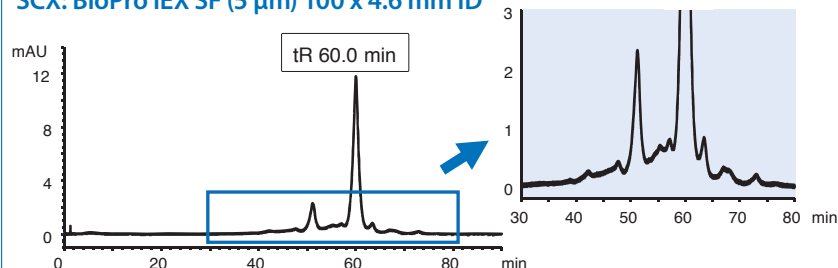


Part No.: QAA0S05-0546WP
 Eluent: A) 20 mM Tris-HCl (pH 8.6)
 B) 20 mM Tris-HCl (pH 8.6) containing 0.5 M NaCl
 Gradient: 0-15%B (0-30 min), 15-60%B (30-60 min)
 Flow rate: 0.5 mL/min
 Temperature: 25 °C
 Detection: UV at 220 nm
 Injection: 20 μ L
 Sample: Tryptic digest of BSA

IEX – BioPro IEX: Antibody analysis

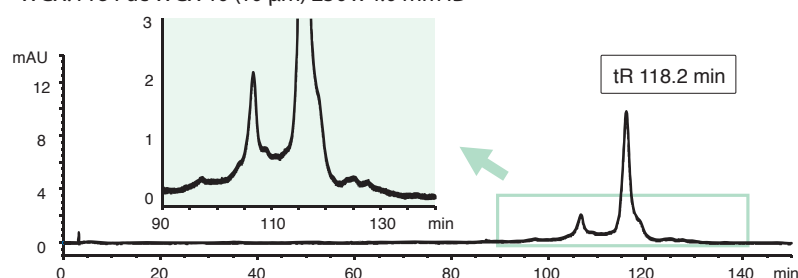
Monoclonal antibody analysis with non-porous cation exchange columns

SCX: BioPro IEX SF (5 µm) 100 x 4.6 mm ID



Higher Resolution!

WCX: Pro Pac WCX-10 (10 µm) 250 x 4.0 mm ID

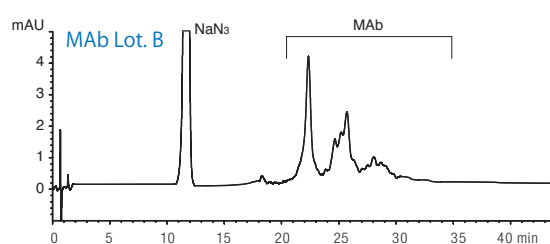
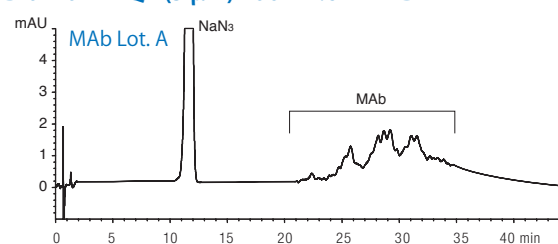


Part No.:	SF00S05-1046WP	Temperature:	30 °C
Eluent:	A) 20 mM MES-NaOH (pH 5.6) B) 20 mM MES-NaOH (pH 5.6) containing 0.2 M NaCl	Detection:	UV at 280 nm
Gradient:	35%B (70 mM NaCl)	Injection:	10 µL
Gradient slope:	0.25%B/min (0.5 mM NaCl)	Sample:	Humanized monoclonal IgG1 (1 mg/mL)
Flow rate:	180 cm/hr (0.5 mL/min for 100 x 4.6 mm ID, 0.378 mL/min for 250 x 4.0 mm ID)		

The separation of MAb is compared using a strong cation (BioPro IEX SF) and a weak cation exchange column (ProPac WCX-10) under the same gradient conditions at pH 5.6. BioPro IEX SF can achieve a higher resolution of MAb than the competitor's column in a shorter analysis time.

QC of monoclonal antibodies with non-porous BioPro IEX QF

BioPro IEX QF (5 µm) 100 x 4.6 mm ID



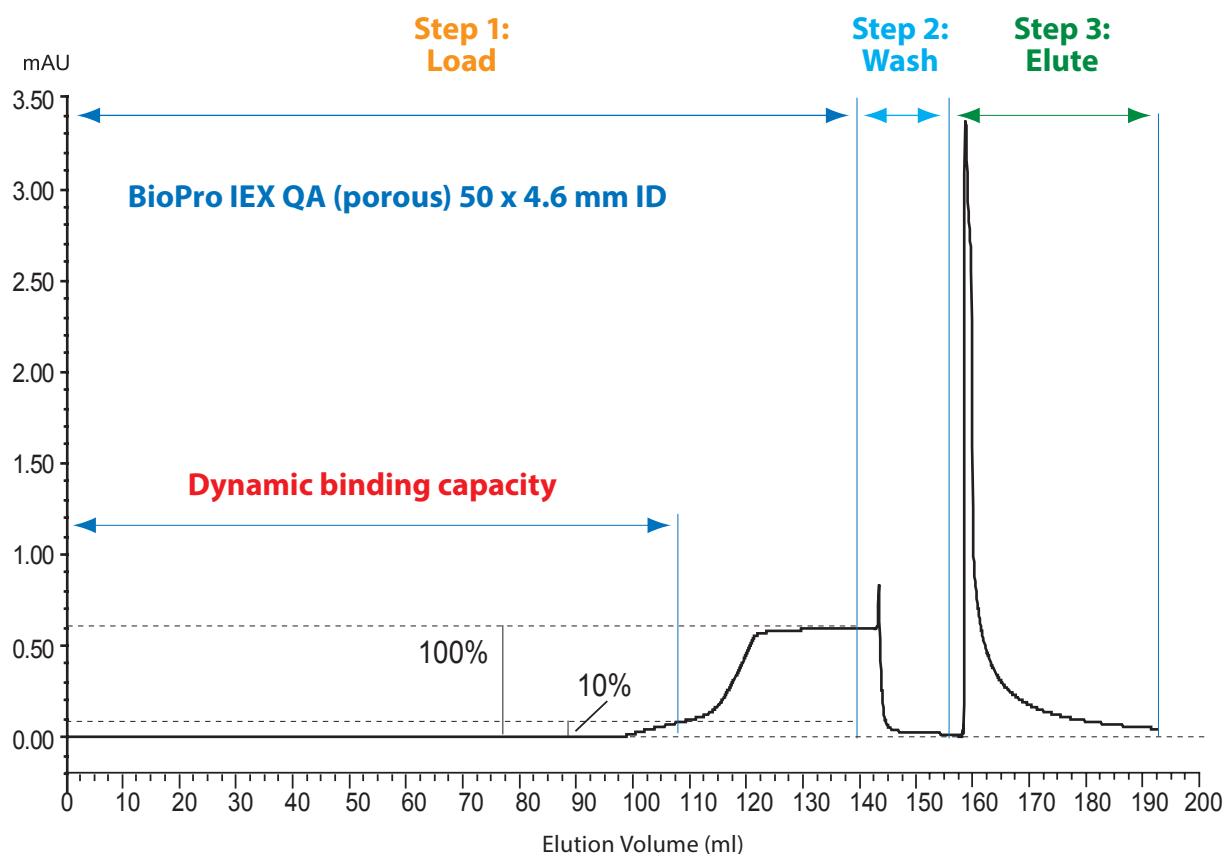
Part No.:	QF00S05-1046WP	Temperature:	25 °C
Eluent:	A) 20 mM Tris-HCl (pH 8.1) B) 20 mM Tris-HCl (pH 8.1) containing 0.5 M NaCl	Detection:	UV at 220 nm
Gradient:	10-25%B (0-60 min)	Injection:	14 µL (0.1 mg/mL)
Flow rate:	1.0 mL/min (360 cm/hr)	Sample:	Mouse monoclonal IgG1 anti-human IgG4 (Purified by DEAE chromatography, containing NaNa3)

Two different batches of commercially available MAb purified by DEAE chromatography were analyzed on a BioPro IEX QF column (100 mm length).

The MAb was separated into several peaks, and the batch-to-batch variability is observed. The BioPro IEX QF/SF 100 mm length columns, which have high efficiency, are ideal for characterization of glycoproteins, such as monoclonal antibodies, and for quality control assessment of biopharmaceuticals.

IEX – Expert Tips: DBC

Determination of Dynamic Binding Capacity*



Before determination, equilibrate the column with equilibration buffer.

Step 1: Load

A protein solution of known concentration is continuously loaded onto the column at the desired flow rate and the absorbance of the eluate is monitored until full saturation is achieved (100% UV absorbance of the pure sample solution).

Step 2: Wash

Wash the column with equilibration buffer until no more protein elutes (0% UV absorbance).

Step 3: Elute

The DBC of the medium is a measure of the volume of protein solution that has been applied up to a specific breakthrough point (usually 5 or 10%).

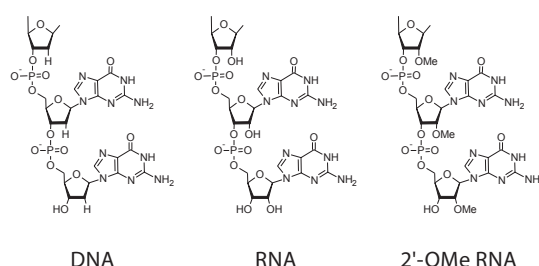
* Application data by courtesy YMC Co., Ltd.

Optimisation of oligonucleotide separations on ion exchange chromatography

Non-porous anion exchange column is generally suitable for analysis of oligonucleotides. For optimisation of single-stranded DNA and RNA of about 20 mer some conditions such as type of mobile phase and column temperature can be changed.

Sample Group 1 (Phosphodiester oligonucleotides; PO)

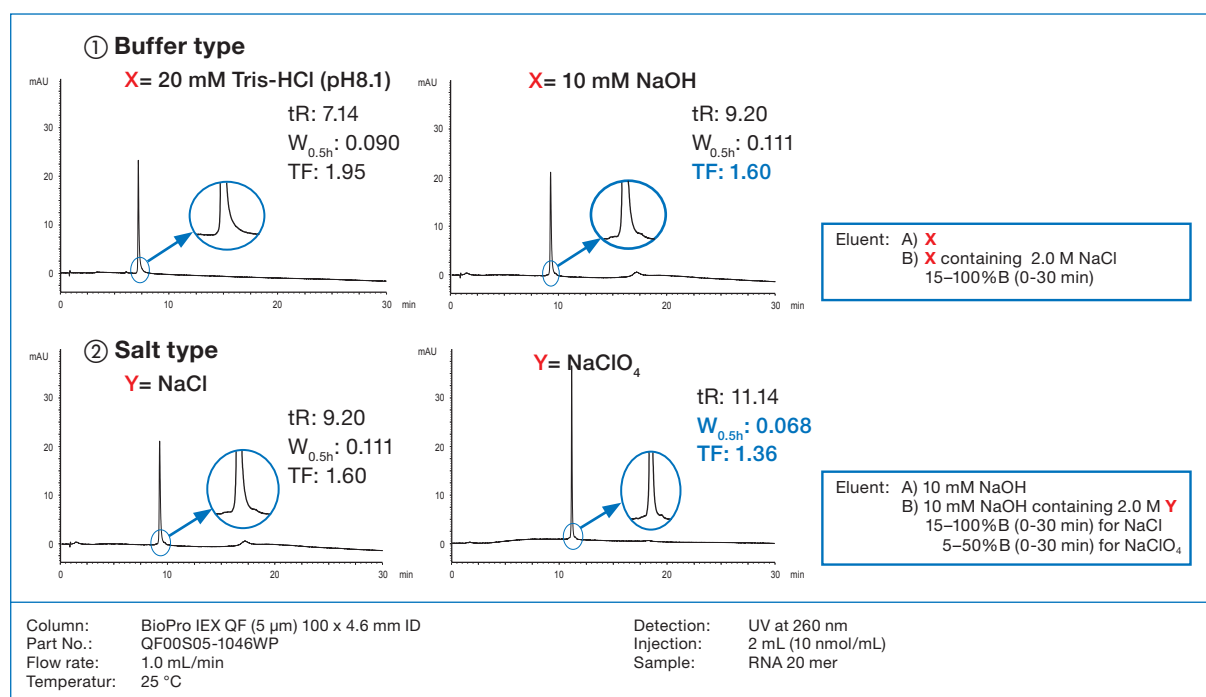
1	Single-stranded DNA (ssDNA)	5'-TCATCACACTGAATACCAAT-3' (DNA 20 mer)
2	Single-stranded DNA (ssDNA)	5'-GTCATCACACTGAATACCAAT-3' (DNA 21 mer)
3	Single-stranded RNA (ssRNA)	5'-UCAUCACACUGAAUACCAU-3' (RNA 20 mer)
4		5'-GUCAUCACACUGAAUACCAU-3' (RNA 21 mer)
5		5'-U(M)C(M)A(M)U(M)C(M)A(M)C(M)A(M)C(M)U(M)G(M)A(M)A(M)U(M)A(M)C(M)C(M)A(M)A(M)U(M)-3' (2'-OMe RNA 20 mer)
6		5'-G(M)U(M)C(M)A(M)U(M)C(M)A(M)C(M)A(M)C(M)U(M)G(M)A(M)A(M)U(M)A(M)C(M)C(M)A(M)A(M)U(M)-3' (2'-OMe RNA 21 mer)



N(M)=2'-OMe RNA

1 Improvement of peak tailing

By changing the buffer from 20 mM Tris-HCl (pH 8.1) to 10 mM NaOH, the tailing factor for an oligonucleotide is reduced. Furthermore, the peak tailing is further suppressed when NaClO₄ was added to 10 mM NaOH instead of NaCl.



2 Improvement of carry over

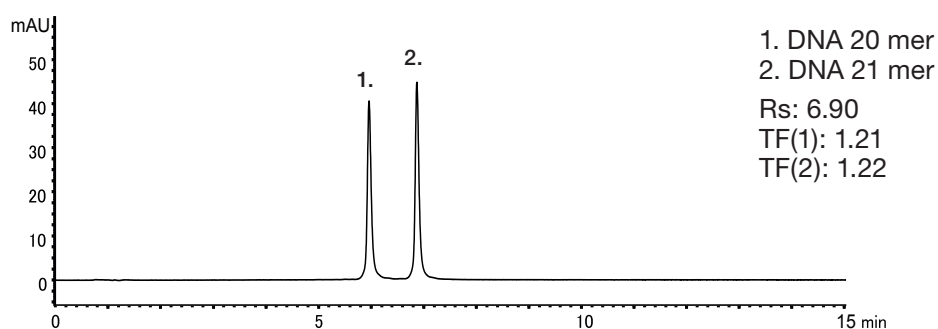
When the initial gradient concentration of NaCl is low (ex. 50 mM), carryover is observed. By increasing the initial gradient concentration of NaCl up to 400 carryover can be avoided with good reproducibility.

IEX – Expert Tips: Oligonucleotides

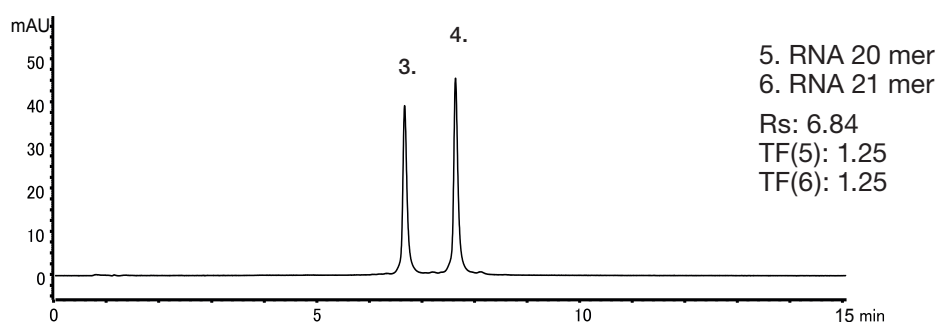
3 Examples under optimised conditions

Each ssDNA, ssRNA and 2'-OMe ssRNA with single-base difference in length can be separated under the optimised condition.

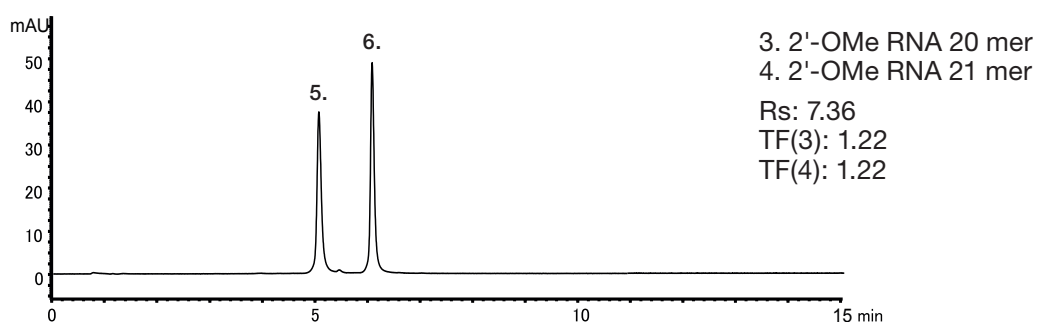
Single-strand DNA



Single-strand RNA



Single-strand 2'-OMe RNA



Column: BioPro IEX QF (5 μ m) 100 x 4.6 mm ID
Part No.: QF00S05-1046WP
Eluent: A) 10 mM NaOH
B) 10 mM NaOH containing 1.0 M NaClO₄
Gradient: 25–55%B (0–15 min), 100%B (15–20 min)

Flow rate: 1.0 mL/min
Temperature: 25 °C
Detection: UV at 260 nm
Injection: 4 μ L (5 nmol/mL each)

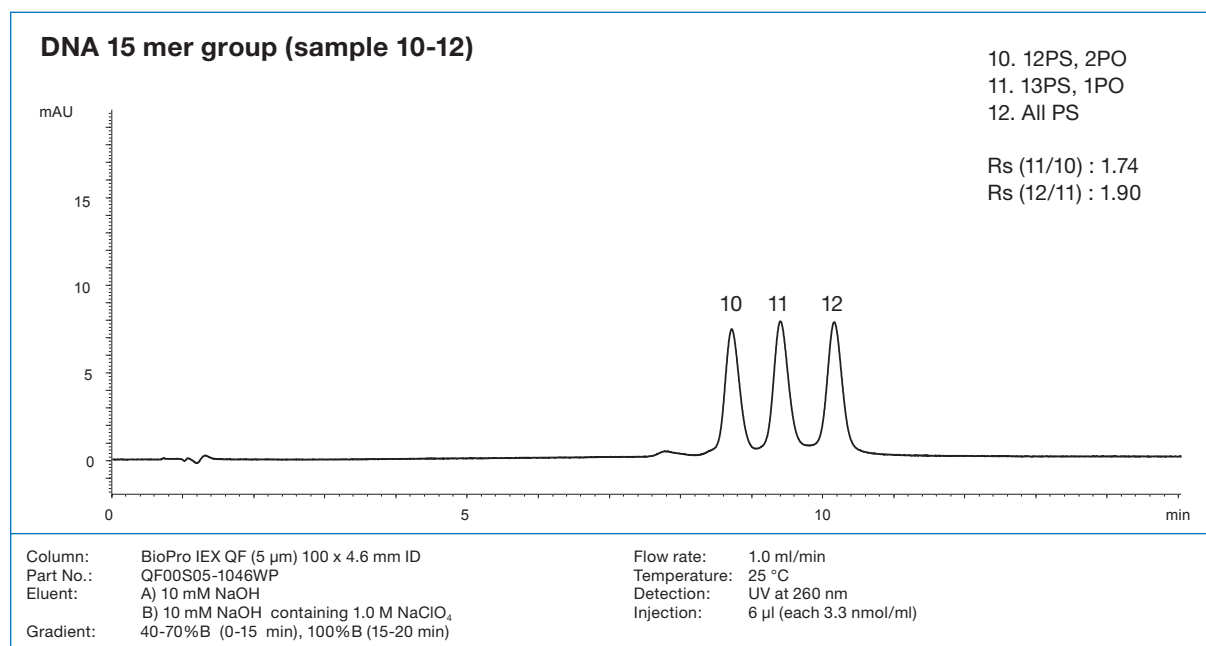
4 Optimum conditions

10 mM NaOH (as buffer solution), NaClO₄ (as salt) and higher initial gradient concentration of salt are preferable to suppress carryover and peak tailing.

IEX – Expert Tips: Oligonucleotides

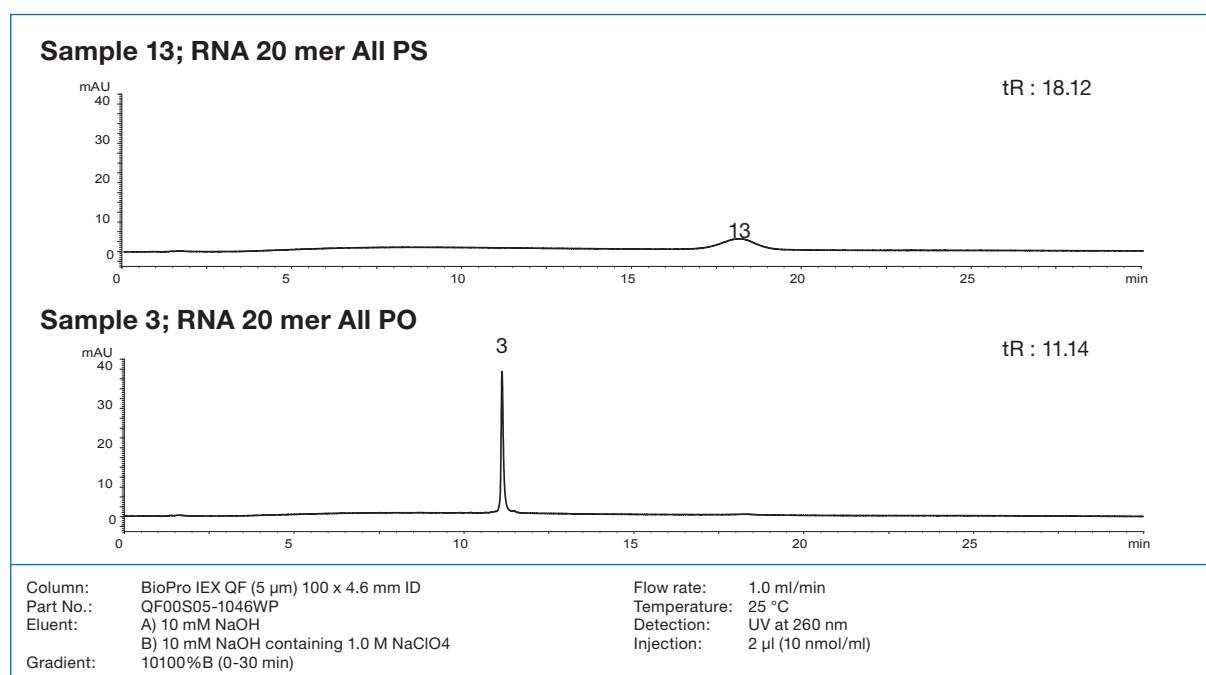
6 Resolution of phosphorothioate oligonucleotides with different degrees of thiolation

Using the optimised conditions described, [all PS], [13PS, 1PO] and [12PS, 2PO] of DNA 15mer (samples 10–12) were clearly separated by ion exchange chromatography.



7 Difference in required salt concentrations for eluting modified RNA (all PS) and normal RNA (all PO)

The separation of RNA 20mer all PS is compared with RNA 20mer all PO under the same conditions. Since acidity of all PS is much higher than all PO, a higher salt concentration is required for elution. The peak of all PS is much broader because it is thought that all PS contains as many as 219 (524, 288) stereoisomers.



8 Separation of optimisation trial of phosphorothioate oligonucleotides with single-base differences in length

A steeper gradient curve, increasing column temperature and adding organic solvent can improve peak separation. However, increase of organic solvent ratio gave little improvement in peak separation.

X/Y=100/0
32–80%B (0–24 min)
= $\Delta 20 \text{ mM NaClO}_4/\text{min}$
25 °C



Steep gradient curve
X/Y=100/0
32–80%B (0–8 min)
= $\Delta 60 \text{ mM NaClO}_4/\text{min}$
25 °C



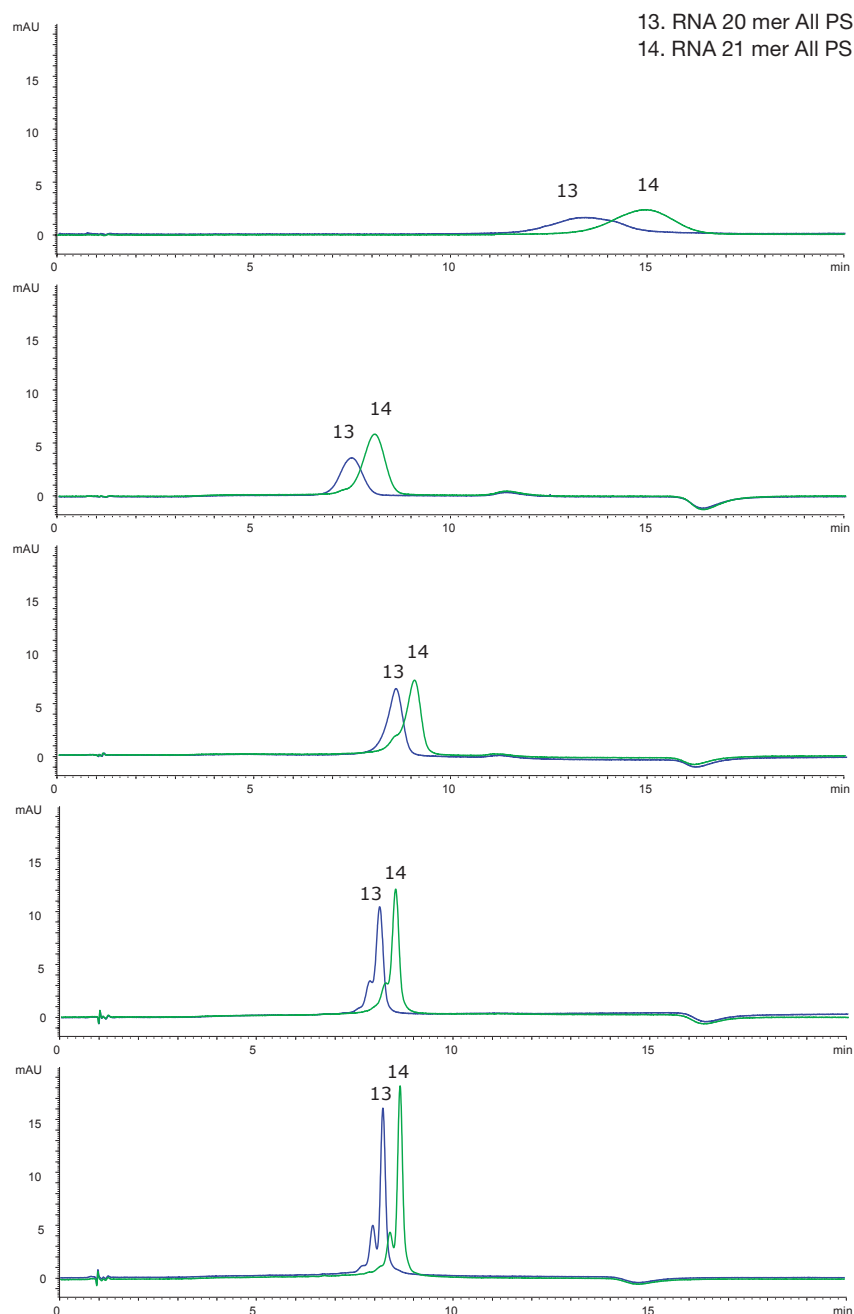
Raising column temperature
X/Y=100/0
32–80%B (0–8 min)
= $\Delta 60 \text{ mM NaClO}_4/\text{min}$
60 °C



Addition of organic solvent
X/Y=80/20
40–100%B (0–8 min)
= $\Delta 60 \text{ mM NaClO}_4/\text{min}$
60 °C



Increasing ratio of organic solvent
X/Y=70/30
40–100%B (0–6.3 min)
= $\Delta 60 \text{ mM NaClO}_4/\text{min}$
60 °C



Column: BioPro IEX QF (5 μm) 100 x 4.6 mm ID
Part No.: QF00S05-1046WP
Eluent: A) 10 mM NaOH/methanol (X/Y)
B) 10 mM NaOH containing
1.0 M NaClO_4 /methanol (X/Y)
Gradient: 1.0 ml/min
Flow rate: UV at 260 nm
Detection: 2 μl (10 nmol/ml)
Injection:

IEX – Ordering information

3 µm non-porous analytical columns in PEEK hardware (max. pressure 250 bar)

Phase	Column ID [mm]	Column length [mm]			Precolumn filter 2 µm*
		30 (250 bar)	50 (250 bar)	100 (250 bar)	(pack of 5)
BioPro IEX QF	4.6	QF00S03-0346WP	QF00S03-0546WP	QF00S03-1046WP	XRPRCP25
BioPro IEX SF	4.6	SF00S03-0346WP	SF00S03-0546WP	SF00S03-1046WP	

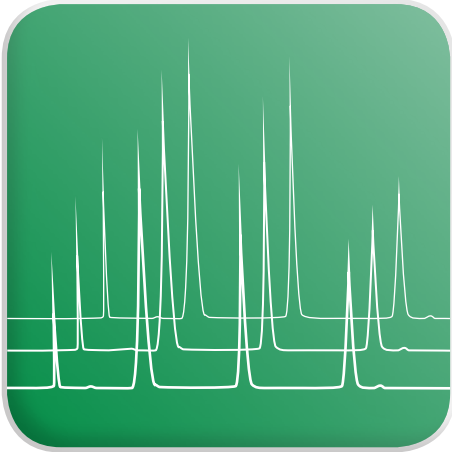
5 µm non-porous analytical columns in PEEK hardware (max. pressure 60–120 bar)

Phase	Column ID [mm]	Column length [mm]			Precolumn filter 2 µm*
		30 (60 bar)	50 (100 bar)	100 (120 bar)	(pack of 5)
BioPro IEX QF	4.6	QF00S05-0346WP	QF00S05-0546WP	QF00S05-1046WP	XRPRCP25
BioPro IEX SF	4.6	SF00S05-0346WP	SF00S05-0546WP	SF00S05-1046WP	

5 µm porous analytical columns in PEEK hardware (max. pressure 25–35 bar)

Phase	Column ID [mm]	Column length [mm]			Precolumn filter 2 µm*
		30 (25 bar)	50 (30 bar)	100 (35 bar)	(pack of 5)
BioPro IEX QA	4.6	QAA0S05-0346WP	QAA0S05-0546WP	QAA0S05-1046WP	XRPRCP25
BioPro IEX SP	4.6	SPA0S05-0346WP	SPA0S05-0546WP	SPA0S05-1046WP	

Other dimensions on demand
 * Holder required, part no. XRPRCP02



HIC

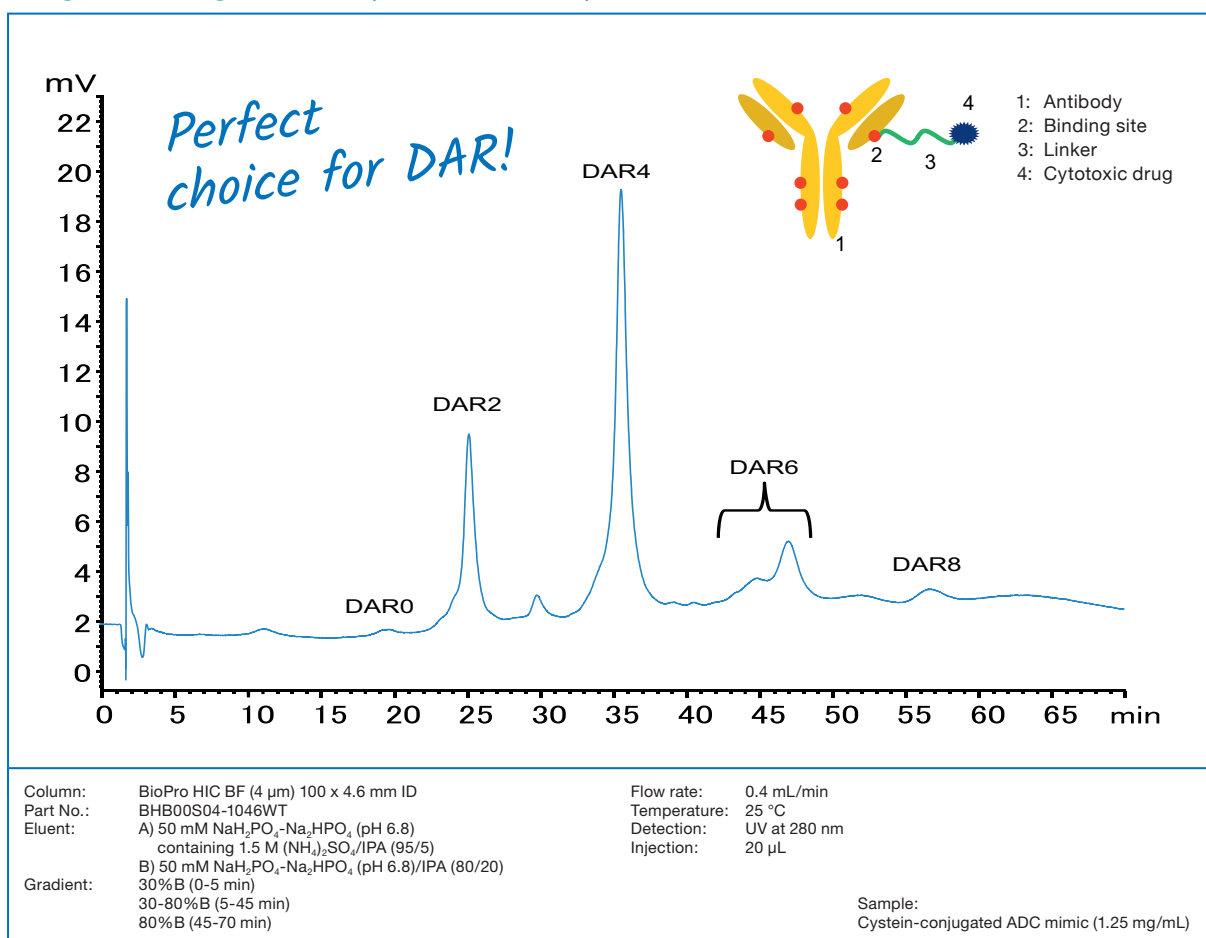


HIC – BioPro Series

- Specifically designed for antibodies, antibody-drug conjugates (ADCs), proteins
- High performance separation
- High throughput analyses
- Excellent lot-to-lot reproducibility
- Long term stability
- Virtually no carryover effects

	BioPro HIC BF
Base particle	hydrophilic polymer (polymethacrylate)
Particle size	4 µm
Pore	non-porous
Functional group	butyl
pH range	2–12
Temperature range	10–60 °C

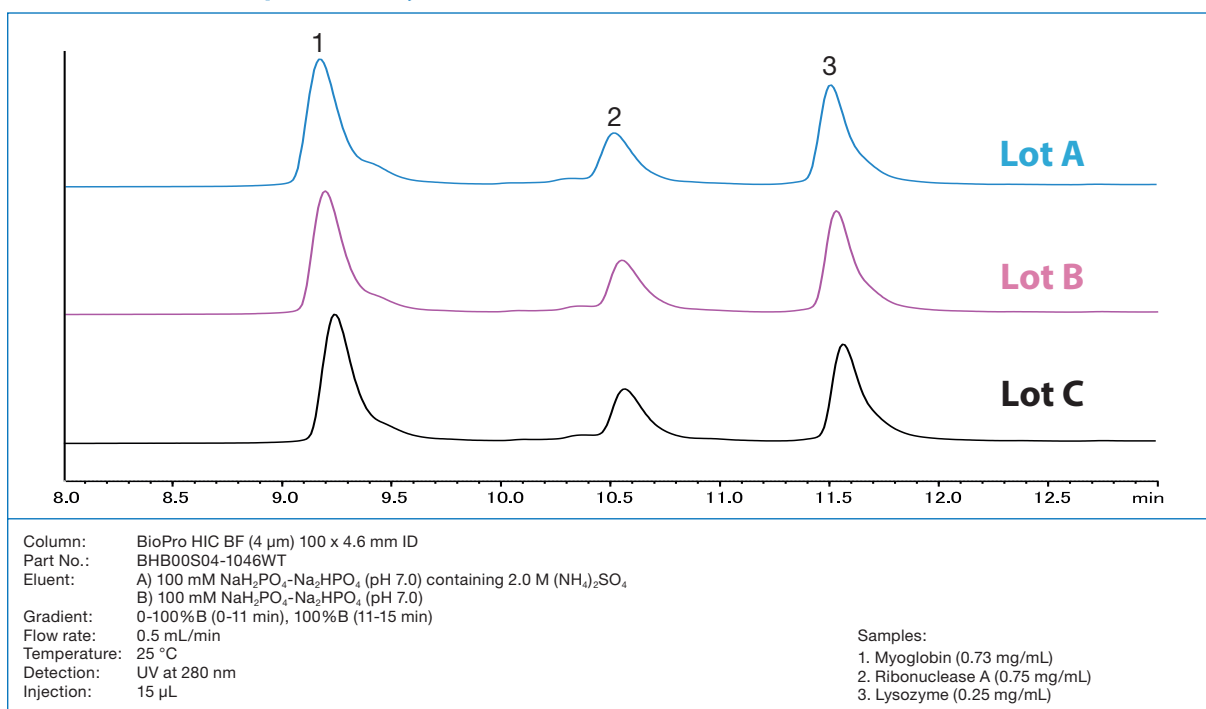
Designed for Drug-to-Antibody Ratio (DAR) analysis of ADCs



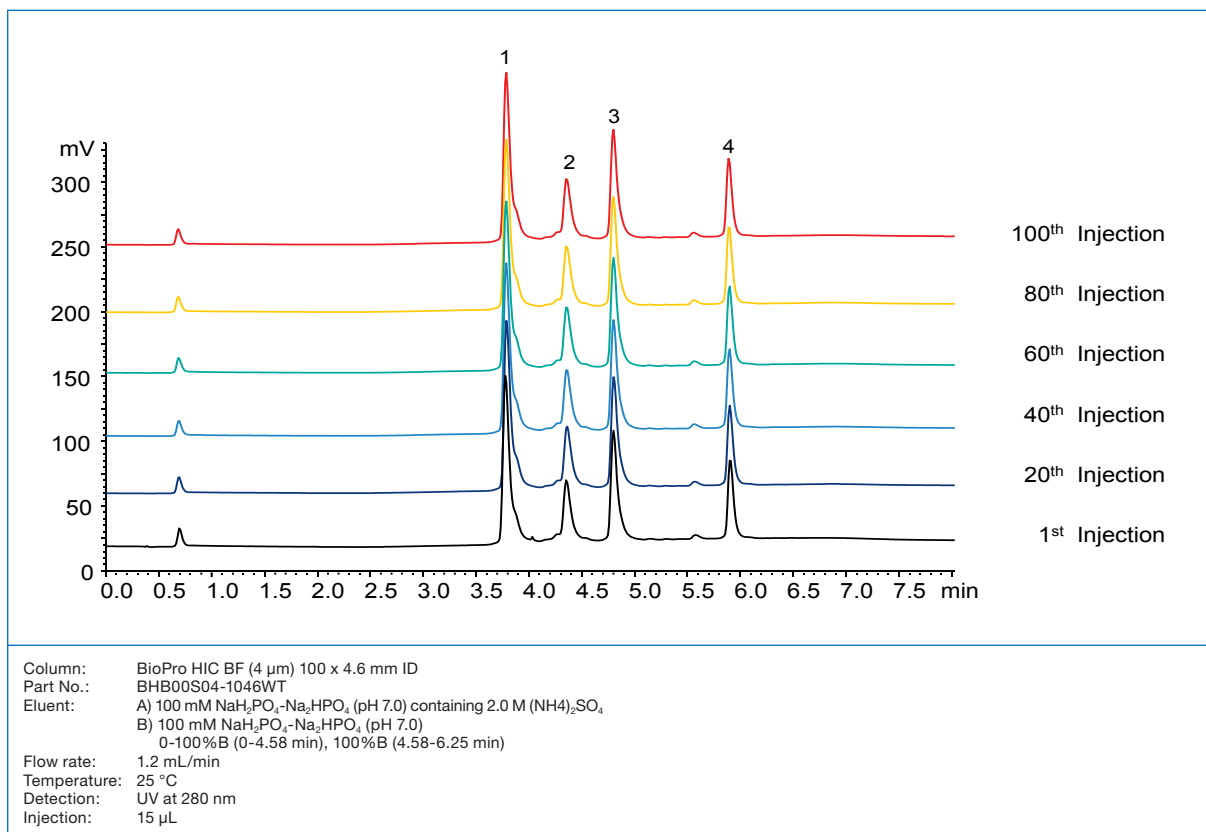
BioPro HIC BF is suitable for ADC (antibody-drug conjugate) analysis, and is especially effective for monitoring of DAR (drug-to-antibody ratio).

HIC – BioPro HIC: Reproducibility & stability

Excellent lot-to-lot reproducibility



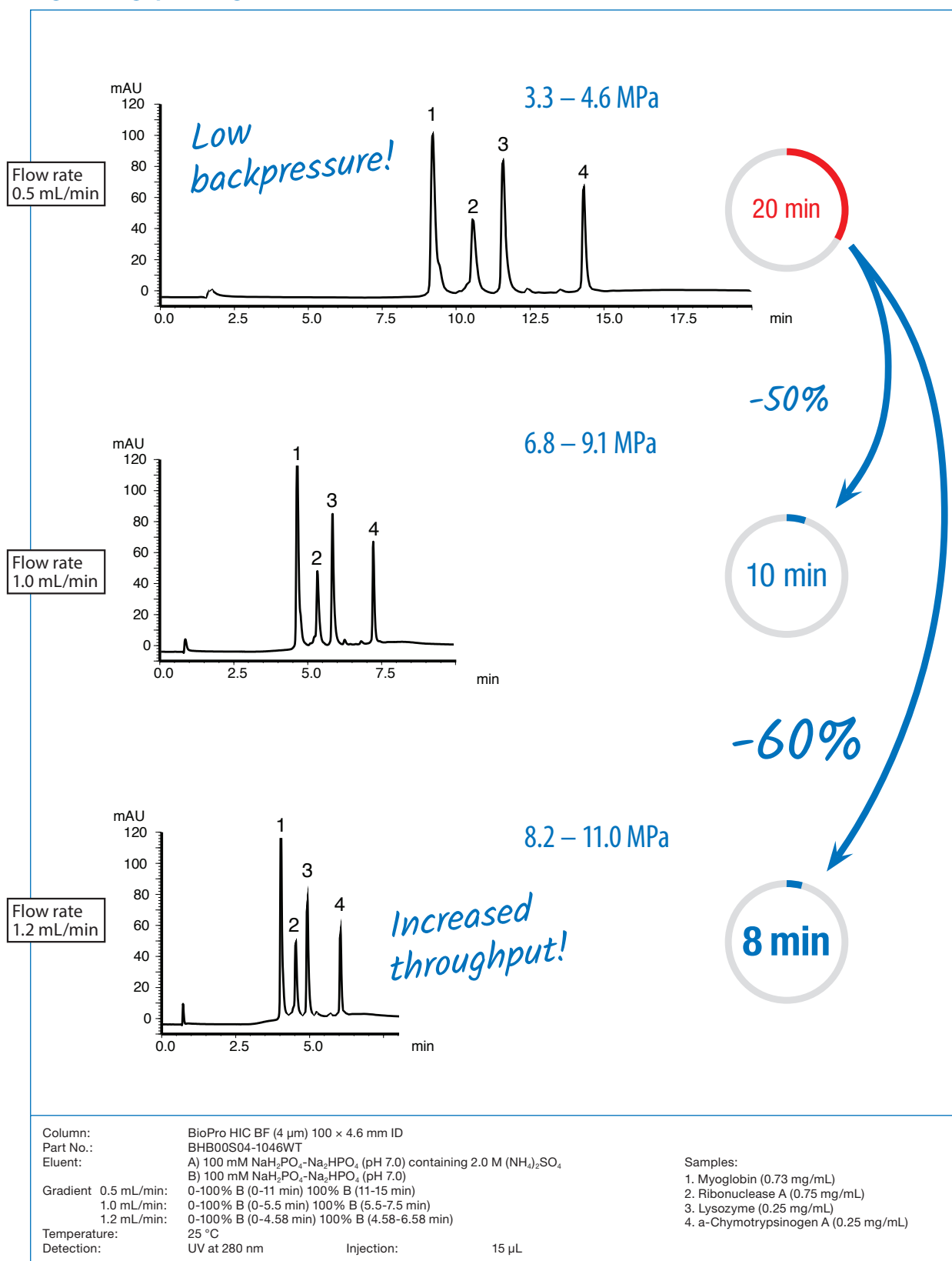
High column stability



Optimised column packing method results in highly stable performance over a long time.

HIC – BioPro HIC: Throughput & resolution

High throughput at high resolution

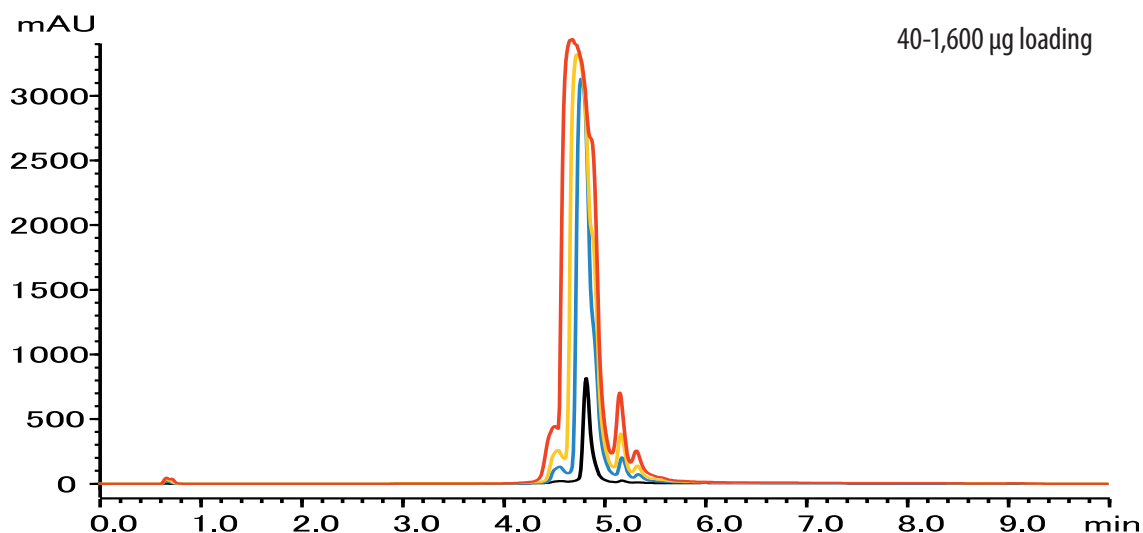


Due to its intermediate particle size, BioPro HIC BF can improve analysis throughput.

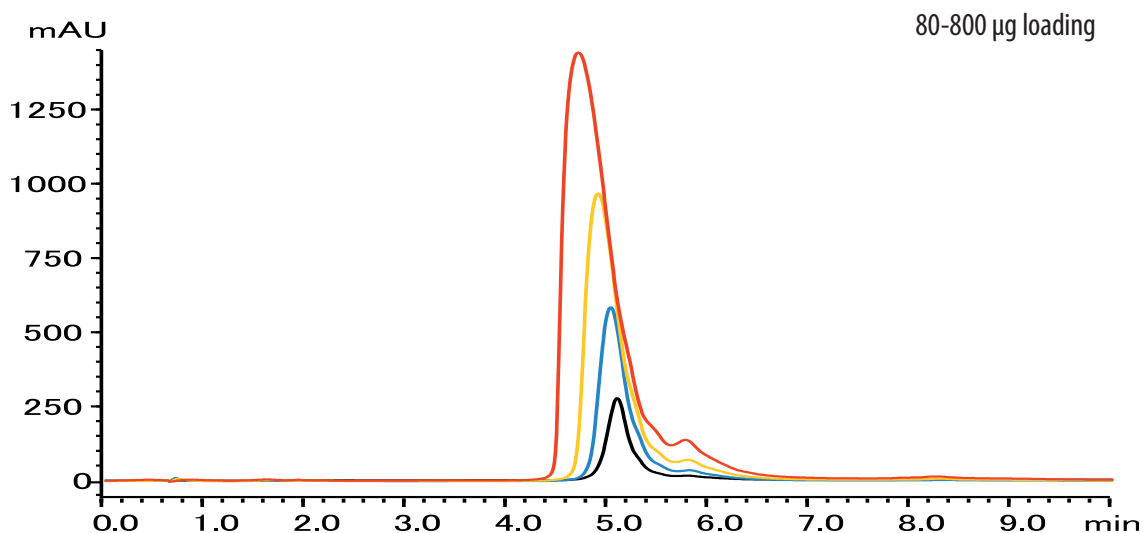
HIC – BioPro HIC: Loadability

High loadability

Lysozyme



Humanized monoclonal IgG



Column: BioPro HIC BF (4 µm) 100 x 4.6 mm ID
 Part No.: BHB00S04-1046WT
 Eluent: A) 100 mM NaH₂PO₄-Na₂HPO₄ (pH 7.0) containing 2.0 M (NH₄)₂SO₄
 B) 100 mM NaH₂PO₄-Na₂HPO₄ (pH 7.0)
 Gradient: 0-100%B (0-4.58 min), 100%B (4.58-6.58 min) for lysozyme
 60%B(0-0.5 min), 60-100%B (0.5-7.5 min), 100%B (7.5-10 min) for IgG
 Flow rate: 1.2 mL/min
 Temperature: 25 °C for lysozyme
 30 °C for IgG
 Detection: UV at 280 nm
 Injection: 4, 40, 80, 160 µL for lysozyme
 32, 80, 160, 320 µL for IgG

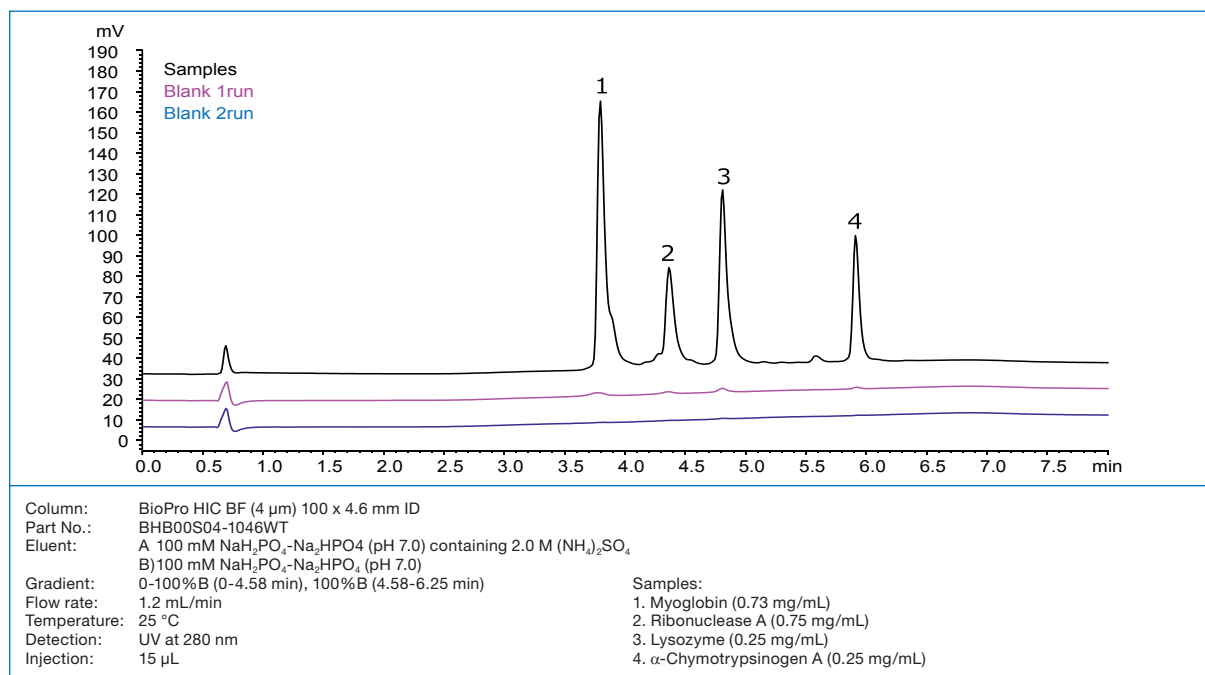
Sample: Lysozyme (10 mg/mL)
 Humanized monoclonal IgG (2.5 mg/mL)

BioPro HIC BF provides superior peak shapes, even under high loading conditions. This allows detection of very tiny amounts of impurities in the sample.

In addition, it can be used for lab-scale purifications e.g. for isolation of variants for various research requirements (e. g. structural analysis).

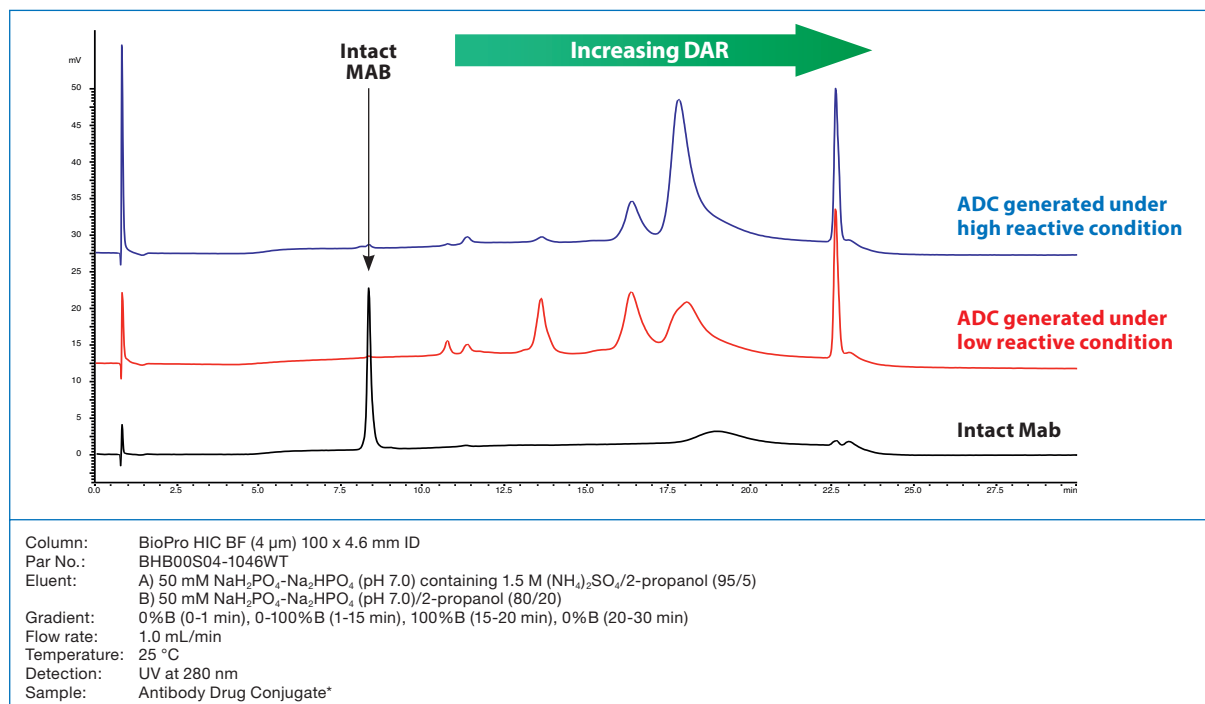
HIC – BioPro HIC: No carry over & ADC monitoring

Suppressed carryover



Carryover between analyses is minimized on BioPro HIC BF. This contributes significantly to reproducibility of analyses.

Characterisation and reaction monitoring of ADCs



ADCs generated under various reaction conditions are shown. BioPro HIC BF is suitable for characterisation as well as reaction monitoring of ADC production.

The influence of salts in HIC separations

The technique known as hydrophobic interaction chromatography is a mode of chromatography that separates proteins by differences in surface hydrophobicity.[1] This method utilises reversible interactions that occur between protein molecules and hydrophobic stationary phase ligands attached to the particle surface.

Certain non-denaturing salts are used to improve the hydrophobic interactions between proteins and the stationary phase. The mobile phase is typically an aqueous solution of salts such as ammonium sulfate or sodium chloride and a buffer to control pH (usually phosphate

buffer between pH 6 and 7). The Hofmeister series of lyotropic and chaotropic ions shown below in Fig. 1 provides a template for salt selection. High concentrations of salt, particularly ammonium sulfate, may precipitate proteins; therefore, solubility should be checked under the initial gradient (binding) conditions. The strength of the interaction between the protein and stationary phase decreases with decreasing salt gradient (see Fig. 2). Another option is a change of pH which results in an increase in the charge on the protein due to the ionisation of acidic groups.

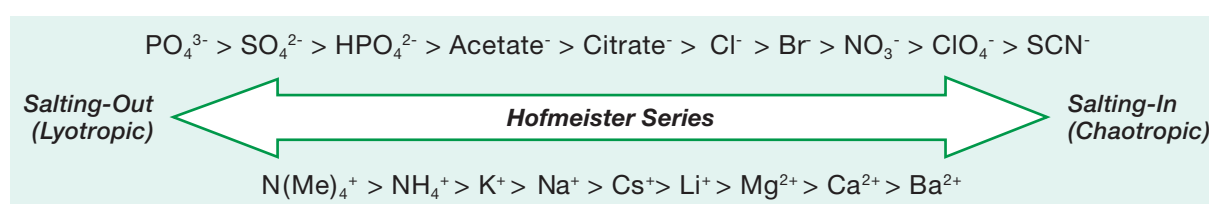


Fig. 1: The Hofmeister Series of lyotropic and chaotropic ions.

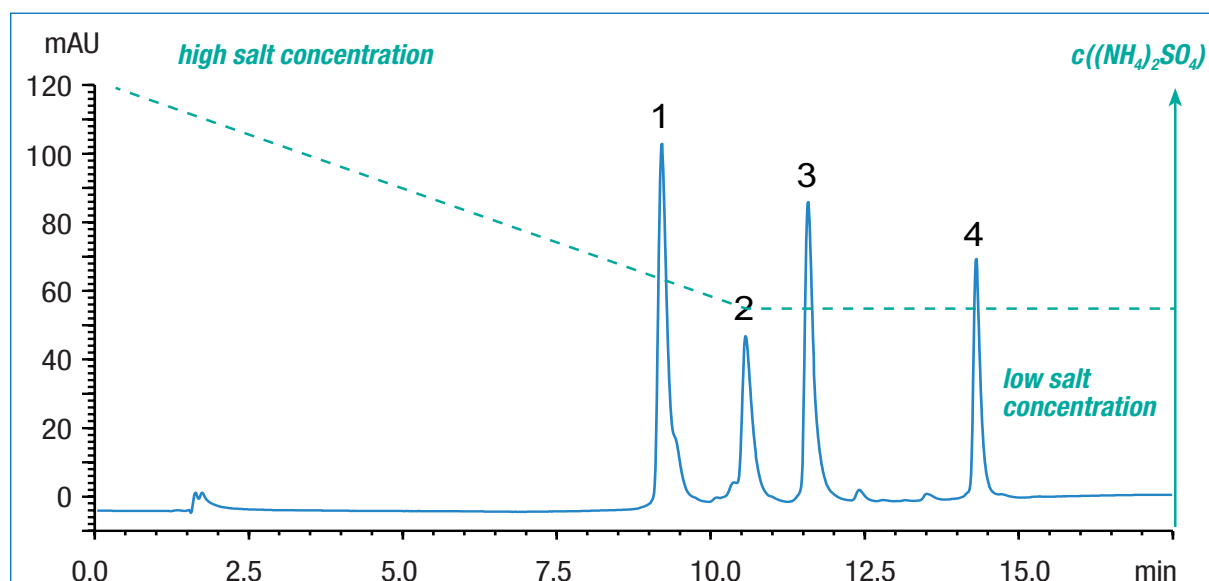


Fig. 2: Method with decreasing salt gradient.

Column: BioPro HIC BF (100 x 4.6 mm ID)
 Part No.: BHB00S04-1046WT
 Eluent: A) 100 mM NaH₂PO₄-Na₂HPO₄ (pH 7.0) containing 2.0 M (NH₄)₂SO₄
 B) 100 mM NaH₂PO₄-Na₂HPO₄ (pH 7.0)
 Flow: 0.5 mL/min
 Gradient: 0-100% B (0-11 min) 100% B (11-15 min)
 Temp.: 25 °C Detection: UV at 280 nm
 Injection: 15 µL

Samples: 1. Myoglobin (0.73 mg/mL)
 2. Ribonuclease A (0.75 mg/mL)
 3. Lysozyme (0.25 mg/mL)
 4. α-Chymotrypsinogen A (0.25 gm/mL).

HIC is particularly effective when used to separate proteins and monoclonal antibodies. The separation of monoclonal antibodies (MAb), MAb aggregates and glycosylated MABs can be achieved due to their specific hy-

drophobic properties. It also provides an excellent method for determination of drug-to-antibody ratios (DAR) in antibody-drug conjugates (ADCs).

[1] Queiroza, J.A.; Tomaza, C.T.; Cabral, J.M.: Hydrophobic interaction chromatography of proteins, J Biotechnol. 2001, 87, 143-159.

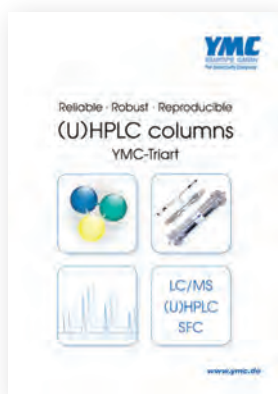
HIC – Ordering information

4 µm HPLC column

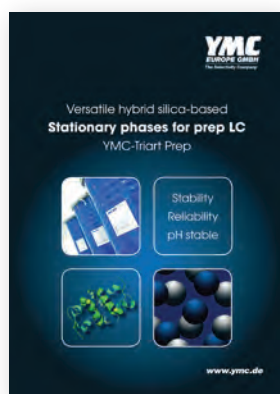
Phase	Column ID [mm]	Column length [mm]	Precolumn filter 2 µm
		100	(pack of 5)
BioPro HIC BF	4.6	BHB00S04-1046WT	XRPRCP25

Holder required, part no. XRPRCP02

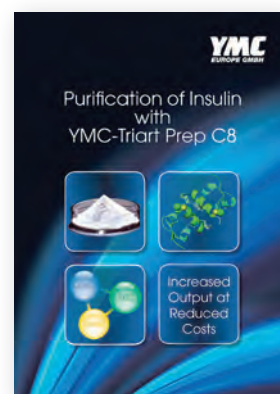
Other Catalogues / Brochures Available



**(U)HPLC columns
YMC-Triart**



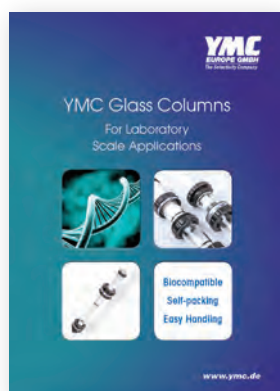
YMC-Triart Prep



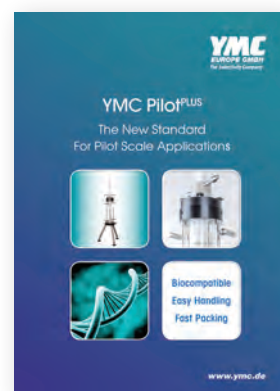
**Purification of Insulin
with YMC-Triart Prep C8**



BioPro IEX Resins



YMC Glass Columns



YMC Pilot^{Plus}

Application data mainly by courtesy of YMC Co., Ltd.

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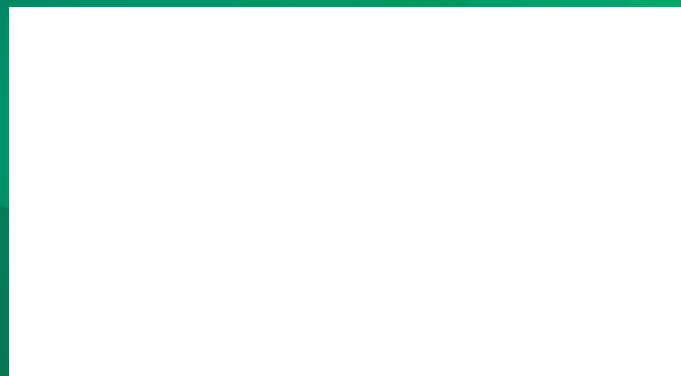
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